CALIFORNIA ENVIRONMENTAL CONTAMINANT BIOMONITORING PROGRAM (BIOMONITORING CALIFORNIA)

SCIENTIFIC GUIDANCE PANEL MEETING

CONVENED VIA WEBINAR BY: OFFICE OF ENVIRONMENTAL HEALTH
HAZARD ASSESSMENT

CALIFORNIA ENVIRONMENTAL PROTECTION AGENCY
STATE OF CALIFORNIA

TUESDAY, JULY 14, 2020 10:00 A.M.

JAMES F. PETERS, CSR CERTIFIED SHORTHAND REPORTER LICENSE NUMBER 10063

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PRESENTERS:

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James McCord, PhD, Center for Environmental Measurement and Modeling, United State Environmental Protection Agency

Rachel Morello-Frosch, PhD, University of California, Berkeley

Jon Sobus, PhD, Center for Computational Toxicology and Exposure, United State Environmental Protection Agency

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PROCEEDINGS

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MS. ZALAY: Good morning, everyone. This is Marley Zalay. It's 9:55, so I'd like to run through some logistics before we get started.

I would like to welcome you to Biomonitoring California's first virtual meeting of the Scientific Guidance Panel. Today's meeting is being held through the GoToWebinar platform only and is being recorded and transcribed.

All attendees are automatically muted when joining the meeting and there will be several options for providing comment during the meeting, which our Chair Meg Schwarzman will be explaining a bit later. If you will be speaking during the meeting, please use a headset, if possible, and remember to speak directly into your microphone and introduce yourself before speaking. This is for the benefit of the transcriber and everyone else on our webinar.

Meeting materials are available to download from our website www.biomonitoring.ca.gov on the July SGP meeting page. If you have technical difficulties during this webinar, please send an email to biomonitoring@oehha.ca.gov and we will have someone from our IT department assist you.

We will have a break at 12:30 p.m. for one hour

for lunch. And we will begin the meeting promptly at 10:00 a.m., so until then we'll just have a few -- a few minutes of everyone getting set up.

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DIRECTOR ZEISE: So shall we start, Sara?

MS. HOOVER: Lauren, what we want to do is start promptly at 10:00, so Marley was dong the gathering at 9:55.

DIRECTOR ZEISE: Okay. I was just checking.

Okay.

MS. HOOVER: Thank you for checking. And she will introduce you at about one minute to 10:00.

DIRECTOR ZEISE: Okay. Very good.

MS. HOOVER: Actually, while we have one minute to go, I will remind that everyone who's joining, if you're a speaker, you don't have to show your webcam until you're actually speaking. So the panelists will always be showing. Thank you.

The panelists will always be showing their webcam, unless they pause. And you can feel free to pause at any time, panelists, and Lauren. And then your name will just appear in the box.

DIRECTOR ZEISE: Okay.

MS. HOOVER. Everyone else will share their cam when they're actually speaking.

MS. ZALAY: Okay. As we approach 10:00 a.m., I

would like to introduce Lauren Zeise, at this time, the Director of the Office of Environmental Health Hazard Assessment.

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DIRECTOR ZEISE: Okay. Thanks, Marley. Good morning, everybody. I'd like to welcome the Panel and the audience to this virtual meeting of the Scientific Guidance Panel for the California Environmental Contaminant Biomonitoring Program, also known as Biomonitoring California. So thank you all for participating and sharing your expertise.

The Panel last met on March 4th, 2020. Just to briefly recap that meeting. After the Program update, the Panel discussed the California Regional Exposure Study, or the CARE study. Unfortunately, due to the COVID-19 emergency, the CARE Study has been suspended.

The remainder of the meeting focused on the SGP's consideration of the class of quaternary ammonium compounds, or QACs, as potential designated chemicals. The Panel deliberations were informed by presentations from two distinguished guest speakers, a guess discussant, and public commenters. The Panel voted unanimously to recommend that QACs be added to the list of designated chemicals for Biomonitoring California, which means that any QAC could be included in a future Program study.

In making this recommendation, the Panel members

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highlighted the rapidly increasing production and use of QACs, significant data gaps in QAC exposure information, high exposure potential for the public and occupational groups, such as custodians and hospital staff, known human health effects including asthma associated with QAC exposure.
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So a summary of the input from the March meeting, along with a complete transcript, is posted on the March SGP meeting page on biomonitoring.ca.gov.

So since we're meeting virtually today, I'd like to have the SGP members introduce themselves. So starting with our Chair, Meg, you want to introduce yourself and give your affiliation.

CHAIRPERSON SCHWARZMAN: I'm Meg Schwarzman from UC Berkeley.

DIRECTOR ZEISE: Okay.

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PANEL MEMBER McKONE: I'm Tom McKone. I'm just going across the picture.

DIRECTOR ZEISE: Yeah, that's great. Why don't we just go across the top.

PANEL MEMBER McKONE: Kind of like we would at a podium.

DIRECTOR ZEISE: Yes.

PANE MEMBER McKONE: Tom McKone, retired from Lawrence Berkeley National Laboratory. I'm still an

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affiliate there and then also Professor of Emeritus at the University of California Berkeley.
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PANEL MEMBER LUDERER: I'm Ulrike Luderer. I'm the Director of the Center for Occupational and Environmental Health at the University of California, Irvine.

PANEL MEMBER FIEHN: Oliver Fiehn.

PANEL MEMBER QUINTANA: I'm Jenny Quintana.

Sorry.

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DIRECTOR ZEISE: Go ahead, Jenny.

PANEL MEMBER FIEHN: Oliver Fiehn, California -- University of California, Davis.

PANEL MEMBER QUINTANA: I'm Jenny Quintana,

Penelope is my real name, and I'm at the San Diego State

University School of Public Health.

DIRECTOR ZEISE: Great. So, Eunha, do you want to...

PANEL MEMBER HOH: I'm Eunha Hoh, School of Public Health, San Diego State University.

DIRECTOR ZEISE: Great. Veena.

PANEL MEMBER SINGLA: Good morning. Veena Singla with the Natural Resources Defense Council in San Francisco.

DIRECTOR ZEISE: Okay. Carl.

PANEL MEMBER CRANOR: Carl Cranor from University

of California, Riverside.

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DIRECTOR ZEISE: Okay. So I think we got everyone, is that right? I think on -- perhaps the orders aren't the same on everyone's screens.

So anyway, welcome, Panel. And with that, I'll turn the meeting over to our Chair Meg Schwarzman of UC Berkeley, who will provide more details about today's meeting.

Meg, on to you.

CHAIRPERSON SCHWARZMAN: Thanks, Lauren. And welcome, everybody, to this new format. Missing seeing you all in person, but it's nice to see your faces here. And I'm just sorry we can't see the other attendees. I want to start by announcing the goals for today's meeting as we usually do. We'll first hear a Program and laboratory update and have a chance to provide input and ask questions about that. We'll review some of the latest developments in non-targeted analysis, and we'll do that through presentations by guest speakers. And then we'll have an open discussion with the guest speakers and the audience to consider next steps for the Department in the area of non-targeted analysis.

So because we're in this new format, some notes on how this meeting will work, which is different than we usually do. During the question periods after each talk,

we're asking that the speakers who presented that talk remain unmuted with their webcam showing so they can respond to questions from the Panel and the audience.

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For SGP members, for panelists, if you want to speak or ask a question, please just raise your hand and I will be watching the webcams and call on you. At the appropriate time, you can unmute yourself after I call on you and ask your question or provide your comment.

If webinar attendees have questions or comments during the question periods after each talk, you can submit them via the question feature of the web platform -- the GoToWebinar platform or you can email them to biomonitoring@oehha.ca.gov. And a reminder to please keep your comments brief and focused to the items that are being discussed at the time.

We'll read relevant comments allowed and paraphrase them, if necessary. We can also receive oral comments from webinar attendees during the public comment periods in both the morning and the afternoon, if you'd rather speak them yourself and during the discussion session that's in the afternoon. So if you want to speak, please use the raise-hand function. Since you don't have your webcam on, we can't see you raise your hand. Use the raise-hand function or the question feature in GoToWebinar, and I'll call on you at the appropriate time.

So next I want to introduce Nerissa Wu who will give our first -- who will give our Program update.

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Nerissa Wu is Chief of the Exposure Assessment Section in the Environmental Health Investigations Branch in the California Department of Public Health. And she's overall lead for Biomonitoring California.

Nerissa will be providing our Program update.

(Thereupon an overhead presentation was presented as follows.)

DR. WU: Hi, everyone. This went so well in practice. Give me a second to get my screen shared here.

Okay. Does everybody see my slides?

(Thumbs up.)

DR. WU: I, of course, can't hear anybody.
CHAIRPERSON SCHWARZMAN: Yes.

DR. WU: So welcome, everybody. I will be giving the Program updates for today. I first want to start with a kudos to the OEHHA crew for getting this all working. It's amazing to be able to see you all. I'm going -- going to be going quite fast through our Program updates and I apologize for that, but I have a lot of material to cover in 16 minutes.

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DR. WU: First, I want to talk about some staff transitions. We have a number of them. First of all,

welcome to two new OEHHA staff, who are joining us to work on the AB 617 projects. Both of them may be familiar to many you. Julia Varshavsky who comes to us from the Program on Reproductive Health and the Environment at UCSF, and Susan Hurley who has worked at CDPH, Cancer Prevention Institute, and UCSF. Both are joining us with enormous amounts of expertise and experience, so we're really excited to have the opportunity to work with them.

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Also, this isn't strictly Biomonitoring
California, but EHIB has a new Branch Chief, Dr. Michael
Wilson, who sat on the Scientific Guidance Panel for many
years, also known as Mr. Meg. And so it's great to have
somebody who really knows and supports our Program.

Sadly, we also have a few goodbyes from our team. Judy Balmin, who was our outreach coordinator has moved on to the Safe Cosmetics Program and Robin Christensen, who has been such a key part of this program, since she joined in 2009, is moving on to be a Branch Chief of the Substance Addiction and Prevention Branch at CDPH.

I also want to just take a minute to remember Reber Brown, who passed away in May. Reber was a long-time staff member at ECL. He did extensive work for this Program, particularly in POPs analysis and he will be missed.

And finally, another note on personnel, a lot of

our staff here at CDPH have been involved with COVID-related activities. And while they're all working really hard to try to cover biomonitoring work, as well as COVID work, our availability as a crew to cover biomonitoring work has been very much impacted, and that's likely to continue for the foreseeable future.

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DR. WU: So when we last met, we were just gaining momentum with our sample collection in San Diego and Orange counties. And we thought we'd be out in the field until the end of May. And we were starting to pay attention to COVID, of course, wondering if maybe we should go to a urine-only protocol or if there are ways we could change our protocol to limit face-to-face contact. But by the middle March with the State going into shelter-in-place, we knew we had to shut down our study.

So we took a very concentrated four days of contacting all of our participants who had been enrolled or had been invited to let them know that we were putting the study on hold. At that point, we had invited 526 participants, including a batch of invitations that went out right before shelter-in-place started.

About 64 percent of those invited responded, meaning that they at least logged on and activated their account. And we were able to complete sample collection

from 90 participants. But 245 people were in the process somewhere and had to be told that we would not be collecting their samples. We got a lot of feedback that people were disappointed and asked to be put on a mailing list. If we were ever able to get back out in the field again, they would like to be part of the study.

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DR. WU: For CARE-2, we had our results go out to individual participants in February. So we're now focused on summarizing statistics and conducting demographic analyses. And we're hoping to have data posted on the web in the next month, and hold a public meeting or webinar at sometime in the near future.

So I'm going to show tables comparing the results of CARE-2 to CARE-LA and talk about some of the demographic associations that have been identified for CARE-2.

But as we look at the tables, I just want to put the reminder out there that we need to keep the issue of generalizability in mind. This is surveillance. So the goal of sampling is to obtain a participant pool that resembles our overall population as closely as possible.

CARE is not based on probabilistic sampling as NHANES is, but we put a lot of effort into matching our region's population the best we can through things like

randomized postcard mailings and quota samplings, which is -- which are designed to match census data,

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DR. WU: And our sampling goals are based on race and gender. And we do a pretty good job of meeting the racial breakdown of each region.

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DR. WU: Our participants do skew a little female more so in L.A. than in Region 2. And we don't select participants based on education and income. And you can see our participants have skewed towards a more highly educated group compared to the overall population. And why does this matter?

Well, there are demographic trends for many of our analytes, meaning that if levels are -- levels might be higher or lower for a particular group. And, for example, if we see that the levels of mercury are higher in Asians than for other groups, which is seen in the literature, if we under-enroll Asians, we might end up with an underestimate for the mean mercury level for the region.

There are lots of ways beyond demographics that study participants might be different from people who do -- who don't decide to enroll in a study and they often can't be measured or quantified the way demographics can.

And this is actually true even if your study does use probabilistic sampling, because if your participation rate is low, probabilistic sampling starts to look a lot more like convenience sampling.

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So it's just a reminder of that participation rate and how -- how closely you are mirroring your region or your overall population is just something to keep in mind as you compare and interpret results.

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DR. WU: So on to the analytes. Here are blood metal results. Detection frequencies were close to a hundred percent for all of the blood metals. You see that the geometric means are somewhat lower in CARE-2 than they were in CARE-LA. We have seen some associations between race and metals. This may be reflective of the overall demographics of the region.

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DR. WU: For urinary metals, and these are creatinine corrected numbers, these are the urinary metals that were detected in at least 65 percent of participants. So antimony, manganese, and uranium are not on this table. And as with blood metals, the urinary metals are a little bit lower in CARE-2, notably arsenic and mercury. And as in the previous slide, it may be reflective of the region's demographics, specifically the lower proportion

of Asians in the region.

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DR. WU: For all the findings I'm presenting, these are results of multi-variable analyses adjusted for other demographic parameters. And for today, I'm only presenting arsenic, mercury, lead, and cadmium. We'll come back another time to talk about the other metals. And there are a few things we can say about demographics and metals. Female participants have higher urinary mercury and cadmium, as well as blood cadmium, but lower levels of lead than male participants.

Levels of arsenic, lead, and cadmium increased with participant age. And there were some race associations with black participants having a higher blood cadmium level than white and Hispanic participants, but Asian participants having a higher urinary cadmium level than white and Hispanic participants. We also saw that urinary mercury levels were higher in participants who chose to participate in Spanish as compared to English. And that's something that we did not see in CARE-LA. So we will continue to investigate it.

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DR. WU: For PFASs, we measure 12 of the PFASs.

All but one participant had at least one PFAS in their serum. And on average, each participant had seven PFASs

detected. Detection frequencies and geometric means were generally lower in Region 2 than L.A., notably, Me-PFOSA's 44 percent lower and PFNA was 32 percent lower. The racial breakdown that I just described earlier might explain some of this, but there's also the well-established temporal trend or the decline of PFAS over time that might account for this. The two exceptions PFHxS went up for CARE-2 and PFOS was -- P-F-O-S was also higher.

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DR. WU: As far as demographic trends, men were higher than women for PFOS, PFOA, and PFHxS, which has been seen in the literature. PFAS trends -- PFAS levels increased with age, with an 8 to 20 percent increase per decade. And there were the racial trends that have been seen in literature in CARE-LL previ -- CARE-LA previously, with Asians being generally higher, sometimes quite a bit higher than White and Hispanic participants. And Hispanic participants being generally lower than other race groups.

Although, the trends were somewhat less pronounced in CARE-2, and this might be because of some of the -- there were small cells for some of the racial groups, so statistical significance is impacted.

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DR. WU: We also looked at 1-nitropyrene, the

biomarker of diesel exposure. And we were able to analyze 160 samples for 1-NP, including only samples that were collected in February and March to avoid the seasonality issue that we saw in CARE-LA. So this table presents the two 1-NP metabolites that Chris Simpson's lab reports to us. 8-OHNP was similar for both regions, but 6-OHNP was quite a bit higher in CARE-2.

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DR. WU: Hispanic ethnicity was associated with higher levels, 30 to 35 percent higher, of both metabolites, but the differences were not statistically significant. And again, we did have small cell sizes for some -- for some race groups. And because there are only 160 samples, as opposed to the 359 of the whole CARE Study, that was particularly a problem for these subset groups.

Age was inversely associated with 8-OHNP. And working with diesel equipment in the three days prior to sample collection was also associated, but with 6-OHNP not 8-HO -- 8-OHNP. In CARE-LA, we found an association between smoking and 1-NP metabolites, but this association was not identified in the CARE-2 results.

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DR. WU: And our last panel was phenols. And this panel usually includes three parabens, bisphenol A,

and its analogs BPS and BPF, triclosan, triclocarban, and benzophenone-3. For CARE-2, butylparaben and BPF were not reported. So of the seven analytes that were measured, only three hit the 65 percent threshold for calculation of a geometric mean and demographic analyses. And I have not included CARE-LA numbers here, because the CARE-LA subset was -- it was 60 people, it was women only, and we selected those samples equally across racial groups rather than in accordance with the region's demographics.

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Because there's so much association between race, and gender, and phenols, it really wouldn't be a valid comparison to take the CARE-LA subset and compare it to the CARE-2 subset.

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DR. WU: So levels of methylparaben and BP-3 were significantly higher in female participants, three times higher, than in men. The levels of BPA were also higher in women but to a lesser extent. We did not find significant differences by race and ethnicity for any of the three analytes. And in literature, there has been seen an association. But with 151 samples in the phenols subset, we had some very small cells. And again, that is something that really impacts our ability to identify a significantly -- a statistically significant difference.

Age was associated with increased levels of

methylparaben and BPA. And in CARE-2, we asked questions about product use in the six hours preceding sample collection. And levels of methylparaben were 91 percent higher among people who reported using lotion in the six hours before sample collection.

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Methylparaben levels were also associated if you had used body wash in the six hours preceding sample collection, but that association was not significant.

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DR. WU: So in summary for each of these panels, this was just our first pass through the CARE-2 data. As we've outlined in these meeting before, we start off with a look at each analytical panel and the demographic parameters and then we start going into exposure data, geography, residence, and other factors like seasonality. So there will be more to report on each of these panels in future meetings.

So what does the future of CARE look like? Well, even before COVID, we had talked about taking a break from field work for the coming year to give us a chance to catch up on analyses and evaluate how CARE was going. At this point, there's even more uncertainty about field work and how we can conduct sample collection in a way that's safe for participants and staff.

It's also really likely that our workforce and

budget will continue to be impacted for some time. So for now, we are not planning field work in the next year still. And we will take this opportunity to continue work on analyzing our CARE data, as well as data that we've accumulated from ACE, and FREES, and other studies. So I want to take my remaining few minutes to highlight two other biomonitoring activities.

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DR. WU: First, the AB 617 projects, the Community Air Protection Program is continuing to move forward. You've heard about the two new staff who have been brought on to work on this study. And OEHHA has contracts in place to start designing studies and get one of those studies launched in this year. Staff are working on identifying relevant biomarkers to include. And like the East Bay Diesel project, the 617 projects are going to include complementary approaches, like air monitoring and ultrafine particle analysis to help us interpret the biomonitoring data.

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DR. WU: We're also working with Dr. Libin Xu who was here at our March meeting to present his work on QACs. It was a very timely topic. With all the disinfectant use these days, widespread exposure to QACs is really a concern. So we are working with Dr. Xu towards a urinary

method to measure QACs exposure. We have urine samples that we've collected through this intra-laboratory pilot project, which is a protocol to collect a convenience sample. We have samples and exposure information from participants in 2018 and we have gone back to those same participants and collected another set of samples and exposure information during the shelter-in-place time period. So we're hoping to use the samples and the exposure information to hopefully get a urinary method working that we'll be able to use in future studies.

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DR. WU: Finally, one recent piece of news, leg report number five, which covers biomonitoring activities from 2016 to '17, it has been approved for distribution. So it's currently on the EHIB website and it will be posted to the Biomonitoring site shortly.

And just in closing, I do want to just thank all of our staff. As always, they're very hard working. Everyone is impacted by COVID and shelter-in-place in different ways. And our staff, without missing a beat, took their work home and have continued to move the Program forward despite all the various challenges that we face.

And with that, I will close up and take questions.

CHAIRPERSON SCHWARZMAN: Thank you, Nerissa.

We have -- you actually moved really efficiently. And we have a few minutes for clarifying questions from the Panel. And there will be a discussion session after our two subsequent lab updates. So questions for Nerissa. And a reminder to panelists, just raised your hand and I will call on you.

Oliver.

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PANEL MEMBER FIEHN: Okay. How interesting.

Thank you for the -- for the report. It's very good to see that there's a lot of progress made and, of, course sad that some of the field work had to be stopped. I'm also sad for the participants.

Now -- now, this -- this situation is going on for many technical studies as well. And many people face the same problem. So have you considered using other types of sampling, for example, in-house sampling, where you could send out, you know, devices from -- I don't want to say dry blood spots, but there are other devices that can also be sampled easily, at least for some contaminants or concerns. Is that something you have discussed or considered?

DR. WU: Yes. And, in fact, in the week preceding our shutdown, we talked a lot about whether or not we could use a urine kit, something we did for follow

up for participants in the BEST study where we mailed a kit with instructions and participants could freeze it and have it overnighted back to us.

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We'd be able to do our phenols, and 1-NP, and urinary metals. But PFASs is a huge interest to the State right now and we would unfortunately not be able to look at PFASs.

It's something that I think we need to hold open as a possibility, not only in the time of COVID, but also we are facing budgetary limits as we always have. And one of the things that's really expensive for us is going out in the field to remote locations. So it's something that we do continue to explore. I think before we invested fully in something like that, we would want to think a lot about the validity and the instructions. It's very hard to get -- for quality assurance purposes to leave the sampling up to, you know, everybody out in the field to follow. And we have found in a number of studies that we have to keep the instructions very, very simple.

And, of course, validity of the sample would be an issue if the instructions aren't followed. But for sure, I take your point, it's something we have to consider.

CHAIRPERSON SCHWARZMAN: Yeah, Carl.

PANEL MEMBER CRANOR: Nerissa, thank you for your presentation. Would you say a little bit more about what you're looking for vis-à-vis the air pollution. I think that's terribly important. I'm just curious where it's going.

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DR. WU: This is actually a question for OEHHA. They are leading the 617 study, so maybe Sara or Marley could answer that question.

MS. HOOVER: Hello. This is Sara. Carl, I'm sorry, could you repeat that? I was doing some behind-the-scenes logistics when you asked.

PANEL MEMBER CRANOR: Just the question of I think air pollution is terribly important. I just wondered what you were -- what you were looking for or where you think that was going.

MS. HOOVER: Oh, so that's a work in progress. We did give -- we've given a few updates on AB 617 work and all the research we're doing. So Susan and Julia, which is great, fantastic addition to our team, they're currently delving into all kinds of different options.

You know, in the past, we measured 1-NP. We're probably looking for other options, more broadly looking at PAHs. We're looking at diagnostic ratios of PAHs. We're looking at biomarkers of effect. We're -- and as Nerissa mentioned, and which is going to be really

critical, we're going to be doing complementary measurements as well in order to help us, you know, actually see if what we're seeing in biomonitoring is linked -- linked to the air.

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So we -- it's quite -- it's going to be quite a complicated design and we're just embarking on that. We just -- just got the contract approved with UC and we're really in the research phase.

CHAIRPERSON SCHWARZMAN: Jenny, go ahead.

PANEL MEMBER QUINTANA: Hi. Jenny Quintana.

Just to follow up on that, if we wanted to give input or look at that, we would then write to you individually. Would that be the best way to do that?

Because I didn't see, for example, black carbon being measured, which would be very critical for 1-nitropyrene.

MS. HOOVER: Whoopsie. Sorry. Little logistical problem there. We do have -- we did measure black carbon in East Bay Diesel. And certainly that's on the table. So we're looking at that as well. Feel free to chime in with suggestions now or, yes, you can always email me. One-on-one is never a problem with the SGP, so always feel free to email me or the Biomonitoring California email, if you have input not during an SGP meeting.

CHAIRPERSON SCHWARZMAN: Any other questions for Nerissa? And actually, I would invite a little bit more

of this conversation about the 617 study, if folks have thoughts, during the short discussion period that follows our next two updates.

Any other questions for Nerissa?

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In that case, we can move on. And I just want to thank Nerissa and really applaud the team for the way that you have so quickly -- everything is moving and changing so fast, and you've really sort of made the most out of the opportunities that still remain, despite the kind of new limitations under -- under the shelter-in-place and COVID-19 situation. So thank you for that update and maybe we'll have time for a little more discussion after the next updates to come.

So thank you, Nerissa. And I want to introduce our next speaker. Dr. Jianwen She is Chief of the Biochemistry Section in the Environmental Health Laboratory Branch at CDPH. And he's going to provide an update on the lab activities.

(Thereupon an overhead presentation was presented as follows.)

DR. SHE: Thank you, Meg for the introduction. Good morning, SGP members and audience. Today I will update SGP what the Environmental Health Laboratory has done since March 2018.

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DR. SHE: My talk today, I will focus on the following activities: completed projects and ongoing projects, publications, semi-targeted analysis, ongoing method development.

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DR. SHE: In 2018, we completed 180 samples from analysis of metals in blood and in urine for Northern California firefighter studies. We also completed 430 sample analysis for the metals in blood and urine and also for 60 samples for environmental phenols for CARE-LA studies.

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DR. SHE: Between 2019 and 2020, we completed three projects. We finished 66 samples for Camp Fire firefighters for analysis of metal in blood and urine. We also completed 1,000 samples to analyze cotinine in serum for a study we called MACOTA study.

Last project we complete is CARE study Region 2. We finished 359 samples for metals in blood and urine, and also among the 359, we finished 151 samples for environmental phenols in urine.

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DR. SHE: In the middle of the year, we plan to complete three projects. One is 66 samples for PAH metabolite in urine for Camp Fire firefighter studies, 90

samples for both metals, phenols. Metals only in blood and urine, but phenols in urine only for CARE-3 study to complete this -- like a submission to stop the project.

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And we also have a three-year grant to complete 1,800 samples for a study, short as, PRECATO. That means each year we finish 600 samples for analysis of cotinine in archived serum samples.

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DR. SHE: At the same time, we published two analytical methods, number one and number three; collaborated with our -- with our collaborators, we published two study findings that listed here as published number two and number four.

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DR. SHE: Now I'd like to talk about a little bit of targeted, semi-targeted and non-targeted analysis.

For targeted and semi-targeted analysis, we usually use different data acquisition and data flow approach. So on the top of this slide, I put the targeted and semi-targeted analysis together since they are more similar. So because we have some pre-conception about what we are looking for, we can select the precursor ions. We have -- can compare to an inclusion list to make sure some chemical we are not interested in, like drugs, are not included. We call this approach data-dependent

analysis, DDMS or information-dependent because of targeted or semi-targeted we know something.

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For the full -- fully non-targeted analysis, we use technology data independent or information independent analysis. So we cannot pre-selection pre-cursor ions, but we -- technically, we need to narrow down our search space, so use a small mass-to-charge range. We still can use the exclusion list to make sure we don't find information people are not interested.

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DR. SHE: The examples. I use a group of carbazole chemicals to demonstrate the technique we are using could be used for targeted, or non-targeted, or semi-targeted. Although, I focus on the semi-targeted analysis.

Carbazole is a group of chemicals often found in the environmental samples at high levels. Think it would be interested to know if they ever get to human body, their analysis are good examples of targeted, semi-targeted, and non-targeted analysis.

Basically, we try to find their fragmentation profile that bear the relationship between the fragmentation profile in our analytical machine to their structure. So they applied to other three different current analysis.

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DR. SHE: In this slide is a spectrum of 3-chlorocarbazole. We run the EI-MS. The structured is showed here. And also in accordance with Huckel's rule, these chemicals, the structure we see, is aromatic structure -- aromatic structure. If a compound is aromatic, it means they're very stable. If they're very stable in the machine, you see a very high molecular ion class, which show the most right side of the screen 201, 203. Also, 201, 203 shows the chlorine present.

Next(inaudible)166 further confirm the chlorine present, molecular ions of chlorine. When...these chemicals -- this group of chemicals will lose group CNH chlorine. And then because our structure very stable, they can form double charged ions. You can see that half mass 100.6 on most of left side of spectrum. But we call this a fragmentation profile we can link to the structure.

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DR. SHE: Another example I use 3-bromocarbazole, other homologue that demonstrates a similar fragmentation profile.

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DR. SHE: If you go further compare this chemical with one the(inaudible)chemical, benzene, chlorine, dibenzofuran, which in the 1980 to 1990s, EPA and CDC put

a lot of effort to study it. Even today, we are still looking for.

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If you compare carbazole and the furan, because they structurally follow the same Huckel's rule. So their fragmentation profile is similar.

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DR. SHE: So with this information, we can definitely say carbazole unfortunately or fortunately is not in human bodies.

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DR. SHE: And targeted, semi-targeted, and non-targeted analysis are complex and curable research topic. I called it forever research topics. In the past, we proposed three separate but still related approaches.

In 1980s, we developed a system called automatic structure elucidation systems with mass spectrometry data. In this system, we use library search and we also tried to spectra interpretation. If the -- if the chemical is not in the library, we try to use knowledge-based engineered to us. We call automatic substructure search if the total structure not there. So fragmentation profile is one of the features we are looking for.

In 1990s, we developed isotope profile match approach. And then because of, as you know, a lot of the or organic chemicals composed of elements, like oxygen --

oxygen, halogens, and sulfurs, they all have very distinguished -- different isotope profile. Specifically, the isotope profile then you can also elucidate the structure of chemicals.

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Three years ago, we developed another approach, because we(inaudible) technology we can't total accurate mass-based isotope profile recognition program. In these systems, basically now we have accurate mass, so we could build a database. And also, we have spectra, we build a library. We also can use in silica metabolite predictions of this information. Together, we can more accurately predict or identify the structure.

So there are related -- separate systems, but related. So they all can be used to guide us to do the targeted, semi-targeted, and non-targeted analysis.

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DR. SHE: This year, we have limited time to work on-site, because we are encouraged to work at home. Here, a few methods we plan to work on in next few months. And then we like develop metabolite in urine of a VOC, which we didn't finish yet.

We also tried to improve our phenol panels, which include the metabolite of BP-3 and the phenols. And there's even the metabolite of 13 other chemicals, like triclocarban and parabens.

Also, we like investigations of deconjugation step to make sure bisphenol A assay is accurate, cover the total bisphenol A. Some questions there I think triggered us to do this investigation.

We also can like to work on the -- I called the semi-targeted analysis and to automate -- automation of data analysis, because like the machine we use is advanced, which cover with -- come with some very advanced program concept, like XTK include, .com included, so which will allow us to do automation.

And then we also plan to investigate -investigation of fragmentation profile for other selected
chemicals of interest for Biomonitoring California.

Thank you. Now, I have time for some clarification questions.

CHAIRPERSON SCHWARZMAN: Thanks for that. We have time for clarifying questions from the Panel.

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PANEL MEMBER McKONE: I'll make sure I'm unmuted. Thank you. Very interesting set of things going on.

I had a question related to exposure potential. So your -- in order to understand how to analyze carbazoles in the environment, you have to do some sort of structure activity analysis, right, I mean, in order to know what fragments and what methods to use. Has there

been a thought about using some of that structure activity research that you've done to maybe look at a possibility of estimating exposure potential, so you might know what ranges we're looking for when we do the biomonitoring?

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DR. SHE: Yeah, that's a very good point. One approach I remember -- if I understand your question practically, one approach we called effect-based analytical approach. Based on the structure and the activity relationship, we group the chemical together. And then with this group within this -- if there are -- that have the same response mechanisms and then we can -- based on this phase, we can have the biological approach. We call it the effect-based approach.

Plus, the instrument analysis can further help with targeted analysis or even improve our sample flow in the laboratory. We haven't done it yet, but metabolite -- based on metabolite prediction can be think as one part of this approach.

PANEL MEMBER McKONE: Thank you.

CHAIRPERSON SCHWARZMAN: Okay. Thank you so much Jianwen.

I'm going to introduce our next speaker. This is our third update. June-Soo Park is Chief of the Biomonitoring Branch in the Environmental Chemistry Laboratory at the Department of Toxic Substances Control.

And he'll provide an update on the non-targeted analysis work that's conducted by his laboratory.

(Thereupon an overhead presentation was presented as follows.)

DR. PARK: Hi. Hello, everyone. Can you hear me okay?

(Thumbs up.)

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DR. PARK: Okay. Thank you. So the -- you know, the -- I'm glad to see you all here again. I think that's the number one priority now days. So I'm going to give our DTSC lab update regarding non-targeted analysis, non-targeted study. This going to be overview for the -- of the project we've been conducting.

So I'm going to call NTA hereafter, in short.

You see the -- our NTA team, Miaomiao Wang, team leader.

I hope she can chime in for some technical question, if I can't answer. Also our chemist, Ting Jiang, Christopher Ranque, and myself.

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DR. PARK: Our NTA workflow hasn't been changed much since the -- my last presentation here a couple of years ago. So we still maintain similar sample prep and instrumental analysis, and the data acquisition. Still doing the feature extraction and alignment. For your recollection, feature was a potential compound of a not

yet identified, but still contained some chemical information like accurate mass, retention time, and the peak intensity.

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We still trying to get rid of some -- the small noise-looking peaks, also the -- some inconsistently detected peaks, some duplicate or triplicate injections, some laboratory background interference, then send them for the realignment. And I have them ready for the identification. It's called suspect screening analysis, so -- by comparing to the database.

So here, we have several database, but here it's human contaminant database. I believe this one has been the most ever gone through most dramatic changes from the last time, because it has grown from 700 chemicals to more than 3,000. Also, they match -- then we resend them to the further confirmatory process by using fragmentation and the reference standard. If they not match, they have to wait for the unknown analysis workflow.

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DR. PARK: This is how the human contaminant database look like. As you can see, consumer product is the major contributor to the growth of this database. Right now, it has close to 1,300. Most of them came from EPA CPDat. CPDat is the chemical in the product database developed in U.S. EPA. Also, it has about 300 PFAS, 200

environmental phenols, pesticide, personal care, food additives, and packaging, and 88 flame retardant plasticizers, phthalates, many other industrial chemicals.

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DR. PARK: I'd like to share our challenges and lesson -- lessons we learned so far from one of our major human NTA studies. It's called, Discovery of Novel Chemicals in 300 pairs cord and maternal serum collected Bay Area. This was funded by NIH. We are in collaboration with Tracey Woodruff's group in UC San Francisco.

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DR. PARK: So as I mentioned, we extracted thousand of features, then identified less than 15 percent at best. Even with that -- the identified features, we are not sure if we identified correctly. In other words, we not sure if the compound was the compound that is supposed to be identified.

So anyway, the people using confidence levels for identification, Schymanski divided into five groups, starting from the lowest confidence level when each -- when only exact mass information available, all the way up to the level one highest confidence level, when you have reference standard to compare available.

So if you apply this confidence levels to our

compounded list, you can see most of them are falling into the confidence level 3 to 4, of course, sometimes 5. Only some of PFAS we can claim as a confidence level 1, because we have analytical standard available in-house.

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DR. PARK: Having that said, the -- we are sending identifiable -- identified features to our confirmatory processor. But we cannot do that all, because it's just simply too many, wasting too much -- or taking too much resources and time.

And that's why we prioritized them, particularly our colleague all-in-one data prioritization, which compounded to go faster in this MS/MS fragmentation experiment. The first thing we -- the compound we collected, the first criteria is the detection frequency, the selected compound showed 100 percent detection frequency throughout the sample, also very abundant, because we don't want to lose signal in the MS/MS fragmentation.

We also selected a compound showing any demographic differences, like education, and race, and ethnicity, and household income is, et cetera. And lastly, the -- among compounds showing correlations between two matrices, maternal and cord serums, we selected compound show that -- one matrix showed a much

higher intensity than the other.

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DR. PARK: Then we come up with the number of target features, 208 for initial MS/MS test. So Ting Jiang, our chemist, conducted an MS/MS experiment. So 15 percent. She come up with a 15 percent as an MS/MS matching rates. That means about 30 compound out of 208 been confirmed by MS/MS. It sounds a little lower than expected. I know. But even these matching rates will also match through not only our work, experimental MS/MS fragmentation peaks, but also the -- some matched through the -- some online tools like experimental MS/MS database, and some computational modeling like in silico.

Ting presented this work at ASMS last month. Of course, it was all online, order of presentation. With this low matching rate, we may not the only one, because we found other study showed the similar matching -- rating the wastewater. Of course, this is the -- you may say the matrix are quite different between wastewater and human. But this is kind of all we can find -- all Ting can find to compare, because it was very rare, very hard to find any human study that went beyond the suspect screening. That's why the -- we brought this Schymanski's group's paper published 2015.

So one thing I'd like to mention, so comparing to

their study it's kind of given us some -- the -- you know, the idea that this kind of low matching rates may not be the very far from the reality, I guess.

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DR. PARK: Anyway, these are the example features confirmed by our MS/MS matching. Some used as food additives and preservatives, like paraben. The -- you can count about 13. Plus, we have 17 more I didn't show you here. We already purchased the -- excuse me -- analytical standard. For instance, a standard or designated data available in the market. But a few of them we are still struggling to get access to them -- to the standard in the market. So they are -- they are currently being evaluated for final confirmation.

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DR. PARK: So we are currently applying very similar NTA workflow to our other ongoing human biomonitoring project, NTA project studies that include some EJ communities, like Fresno that we've been working with Lauren Zeise group, and OEHHA. We analyzed the Fresno samples, 70 pairs of cord and maternal serum and currently compare it to the Bay Area data I just presented before.

You know, another study, we are working with Rachel Morello-Frosch team at UC Berkeley. We are

comparing chemical profiles in others, some women firefighters, nurses, and office workers.

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DR. PARK: We currently have NTA-related manuscripts in progress, on number two and three, related what I already presented earlier. And they're are -- they are the human data. Number one is the lead my Miaomiao Wang. And this is regarding the data from EPA NTA studies, NTA....studies that we have participated in a couple of years ago.

So I got five minute warning already.

Okay. So and number four is -- this is also the non-targeted PFAS studies investigation in wastewater.

Number five is the -- regarding stormwater runoff following 2017 wildfires in Northern California. We are working as two-track redoing(inaudible) and she's took a part -- took a role for some volatile and semi-volatile persistent chemicals. So we are doing some polar compound -- some polar non-target compound.

And six, seven, and eight, I think they are much slower progress like the cat and the firefighters exposed to AFFF and even metabolomics, but we'll get there. We'll get there.

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DR. PARK: So we'll continue testing our

analytical standard for final confirmation. And then we're going to select 5 to 10 chemicals to quantitate using a calibration curve. Of course, we will continue our effort to improve our identification rate and confidence.

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Here's two of our biggest challenges. You heard a lot about it. I don't want to say any more, but -- you know, that we cannot control. We are understaffed. So this unknown identification workflow is kind of on hold, even though we have more than 85 percent of extracted features not identifiable by database.

Also, some non-targeted volatile and semi-volatile analysis using GC/Q-TOF. We just have installed in branch, but they're not -- they're not much thing I can help. But hopefully we can utilize this instrument soon.

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DR. PARK: I'd like to give many thanks to the funders and the collaborators. I just couldn't display all, because of limited space. There are many others working together, have a lot of fun conducting interesting project.

This is it. Thank you very much.

CHAIRPERSON SCHWARZMAN: Thank you for that update. I have one quick question and then we'll invite

other clarifying questions from the Panel.

DR. PARK: Sure.

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CHAIRPERSON SCHWARZMAN: That's -- you showed the correlation between the maternal and the serum blood, you call it intensity -- sort of intensity of signal. I assume that's in the non-targeted analysis of maternal and cord serum, and that 50 percent went one way and 50 percent went the other way, sort of, you know, more intense in the maternal serum compared to cord serum and 50 percent were the other way, more intense in the cord serum than maternal.

And I was wondering if you -- if it's possible to say anything about the sort of characteristics of the signals or the -- or compounds that you can identify that are consistent among the chemicals that tend to be more intensely apparent in cord blood versus maternal or the opposite.

DR. PARK: Well, I think I'm sure that we will get there. Right now, the -- we are under -- are on suspect screening analysis. It's all chemical analysis part. But we are working with Tracey Woodruff's group. Eventually, we will get there scrutinizing individual peaks, you know, that have any significant meaning. So the -- that's -- right now, the two papers is all kind of a prioritization and also chemical analysis by using the

MS/MS the fragmentation technique.

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But the follow-up manuscript, we will be more focusing on the submitting of each individual -- the chemicals we identify than from the -- based on the old demographic information and all the statistical information.

CHAIRPERSON SCHWARZMAN: Yeah. Thank you. But one of my -- one question is sort of whether even absent being able to identify individual chemicals if you can -- if you can say anything about the characteristics of the chemicals that tend to be more apparent in maternal serum than in cord serum or vice versa.

DR. PARK: Yes, probably so, just by looking with bare eyes. You know, the -- if you see some of the parents compound metabolizer tend to be existing higher in test -- in cord blood. And you may be able to tell which one distribute the higher portion each matrix. So I'm sure that you can -- you can the -- evaluate that without, you know, any further deep statistical analysis.

CHAIRPERSON SCHWARZMAN: Okay. Great. I would just be curious if you -- if you wind up with a characterization of that?

DR. PARK: You're not the only one. Trust me, you're not the only one.

(Laughter.)

PANEL MEMBER SCHWARZMAN: Oliver, you had a question.

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DR. PARK: Yeah, I saw a couple of hands raised there. Oliver, did you --

PANEL MEMBER FIEHN: Thank you. So I wondered if you have considered also collaborating with external partners in data processing? I mean, chemical laboratories, you know, similar to Dr. She's laboratory who are usually doing target analysis may have not like the necessary experience to look at non-targeted screening, using all the resources, using all the possible tools. It's a little bit overwhelming. So have you considered like contacting others, the various centers in the United States, who actually focus these days on exposome analyses and compound ID.

DR. PARK: Yeah, I think -- Oliver I think if you propose your group as a potential collaborator, yeah. Of course, yeah. Also, Jon Sobus used to help us a lot by introducing his teammates who is very expert in this area. Eunha, also the -- you know, the -- also some SFEI folks, and many others. But eventually, the -- a lot of works need to be done. That's our part. So, of course, when we need to reach out for help, you know, you will be the first one we are going to contact.

(Laughter.)

DR. PARK: I know where you live.

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CHAIRPERSON SCHWARZMAN: Eunha, I saw that you had a question, but I'm going to put that -- just a minute. I have your name written down. We need to go to public comment for a few minutes and then we'll have a chance to have a discussion based on all three of these presentations. And I have Eunha first to ask a question or start a discussion item.

So with that, I want to announce the public comment period starting. We have ten minutes allotted for public comment. And I want to just give you a reminder about how to submit comments. Webinar attendees can submit them in writing via the GoToWebinar question feature or email to biomonitoring@oehha.ca.gov. And you can also speak, if you wish. In that case, please alert us by using the raise hand or the question feature in GoToWebinar platform. And we have staff monitoring that and we will call on you.

So I'm going to start by checking in with Marley and Sara, if you know about any public comments.

MS. ZALAY: This is Marley. There are no current -- no questions currently.

CHAIRPERSON SCHWARZMAN: Sara, you're muted if you're trying to talk.

We can't hear you, Sara.

No.

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Okay. We'll have to wait for Sara to communicate in another way. She might message one of us. Inevitable hiccups. And it's okay to leave a moment here anyway, in case people didn't anticipate this moment being public comment and want to still email one or raise hand in the web platform, or anything like that for making a public comment.

There is also another public comment period in the afternoon, if you intended to, but sort of don't make this chance.

MS. HOOVER: Can you hear me now?
CHAIRPERSON SCHWARZMAN: Yes.

MS. HOOVER: Okay. I had to leave the webinar. Sorry, now I'm getting an echo. I had to leave. It would not let me unmute.

Okay. Apologies for that technical delay. I am going to go ahead and read the public comment. This is a question from Jay Murray. His question is for Nerissa. Were the urine samples spot samples or 24-hour samples? And I can actually answer that. They were spot samples. And then Jay also has a comment to share on Nerissa's talk.

He says that molybdenum is an essential element and your geometric mean values for molybdenum in urine are

consistent with what is known about dietary intake.

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And that is the only public comment that we have received.

So, Meg, you can go back to Panel questions and comments.

CHAIRPERSON SCHWARZMAN: Okay. Thank you for that. Sorry for the wrinkle.

We now have until about 11:30 for Panel discussion and any other questions that we didn't cover. So this is meant to be a discussion of all three presentations that we've heard before, the program and the two laboratory updates. A reminder to just raise your hand if you want to speak, and I'll call on you. But I want to start with Eunha who had a question or comment earlier.

PANEL MEMBER HOH: This is Eunha Hoh from San Diego State University. I have some questions first, curiosity and clarification, to June-Soo. Great work, June-Soo. It's very impressive that what you've done and presented. I would like to know about the details. A couple questions that you're talking about the uncertainty. Is that -- when you run actually acquired authentic standards is actually ran -- run those authentic standards and then compare them with your data, do you get better matching, you know, compared to your -- comparing

with the database?

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DR. PARK: Eunha, there was the million dollar question. We are working -- we are -- informed you we already purchased and injected most of the standard we were able to acquire from the market. So that's another story. So we have only 15 percent matching rate in the MS/MS fragmentation. Also, the -- in the -- so far, we have successfully acquired data from the standard injection. Also, the some -- some percentage were not matched. So that chance is kind of getting slim and slim. So I think we are getting -- realize, that we are getting into the -- we are now realizing where we are right now.

PANEL MEMBER HOH: Um-hmm.

DR. PARK: So I think, again, I have to emphasize this takes a lot of time and effort. Our particular outing done in the Bay Area study, in MS/MS, this is all manually -- you know, the -- the -- the -- have to met kind of interpret or the fragmentation patterns by just bare eyes by a synthetic chemist background. So there is no shortcut. Of course, sometimes the software can help you the kind of shortcut, but it's not all the time.

PANEL MEMBER HOH: Um-hmm.

DR. PARK: You know better than I do.

PANEL MEMBER HOH: One more question just for the -- your current human biomonitoring NTA projects, the

number two the woman firefighters and nurses, office workers, what type of samples do you use for that?

DR. PARK: That's the blood. I'm sure Rachel

Morello-Frosch will give more details this afternoon. But that was the human biomonitoring project I've been talking about is all related to the serum samples.

PANEL MEMBER HOH: Okay. Okay.

DR. PARK: Hi, Jenny.

CHAIRPERSON SCHWARZMAN: Anything else, Eunha?

That was it?

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PANEL MEMBER HOH: (Nods head.)

CHAIRPERSON SCHWARZMAN: Okay. Jenny, please.

PANEL MEMBER QUINTANA: Hi. Thank you for that presentation. I had, I guess, a comment more than a question, as well as a question. But one thing is that you said that you used a hundred percent detection as one of your criteria, which I can imagine you use, because you want to discover environmental chemicals that are very common and might be an important target for intervention. But it also occurred to me you might miss health disparities between populations by requiring a hundred percent detection. So theoretically, if you had a hundred percent detection in a disadvantage population, let's say

exposed to a lot of diesel exhaust, you might have very

low detection perhaps under your instrument sensitivity

for the other people, and so you might miss an important, you know, contaminant that would be important for health disparities. And I guess, I'm wondering if you had thought about that.

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And my second comment, just to give them all right now, is following up on Meg's comment about the cord blood. It does seem like chemicals that are more present in the cord than the mother would be the first priority for me, ones that are especially, you know, present in the cord. So thank you. Thank you again for that talk.

DR. PARK: Should I answer to question?

PANEL MEMBER QUINTANA: If you had a comment
or --

DR. PARK: No. I think -- I just have a brief comment on your first question. I -- again, I already totally agree with your second comment.

On the first one, the -- yes, we're going to miss a lot of -- but I'm -- I -- with the presentation slide, I just show you -- it's a very initial study to confirm the compound we -- also, the past compound we identified. Of course, we would -- by doing that, you know, some population disparity, that any compound related to kind of the disparity I think we're going to miss it. But we're not going to ignore those data. We will definitely go back as a follow-up study.

Right now, the MS/MS confirmation, that's the main goal of this presentation right now, also the manuscript we tried to see, you know, the -- how many compound we identify can be matched at least as a percentage. And then you -- as you can see, it just dramatically drop the matching rate in the MS/MS. So matching rate is dropped even final confirmation using the standard. So that's what the kind of reality we like to learn from this major project.

PANEL MEMBER QUINTANA: Thank you.

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DR. PARK: So one more thing. Beauty of this non-targeted, data won't go anywhere. So I think it's -- it's in the hard drive. Whatever the purpose and goal we are setting, then we will go back to the data and reinvestigate the hypothesis.

CHAIRPERSON SCHWARZMAN: Thank you for that.

Carl, you had a question or comment.

PANEL MEMBER CRANOR: A quick question. It's a technical question, but also maybe a question for Sara. In the air pollution study, is there any way to -- do you detect the particulate matter or do you just detect things that might be carried on particulate matter, if both things happen. I'm curious about that, because if you look at some of the studies out there on particular matter -- particulate matter is particularly worrisome,

but it carries other things with us.

MS. HOOVER: Yeah.

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PANEL MEMBER CRANOR: So curiosity question.

MS. HOOVER: Sorry?

PANEL MEMBER CRANOR: I'm done. That's the question.

MS. HOOVER: So as Jenny mentioned, you can measure black carbon. In terms of biomonitoring though, we don't -- and I welcome comments from anyone, but we have not identified a specific biomarker, for example, for PM2.5. So we will tackle it using a variety of tools as I mentioned. You're right, that particulate matter can be associated with certain chemicals. So we'll be looking at that option. We'll be looking at air monitoring. We'll be looking at, you know, this really cool work that a lab in DPH does, which is actually looking at particles and doing source -- exposure source work by looking at the particle itself.

The other thing I forget to mention - so thanks for bringing up the study again - we're also hoping to do some non-targeted screening actually on air samples. So as you probably are all aware, there's very specific chemicals that are monitored in air. There's not generally a really broad sweep of what's being found in air. So we don't know if this is going to pan out, but

there's certainly the capability of doing an open scan, for example, of VOCs. So we might identify a facility of concern in an AB 617 community and do some non-targeted screening work outside that facility or in a particular location in the community not necessarily tied to a particular facility. So that's another option.

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So it's very complicated. As you've probably heard in our other talks, it can be really challenging to biomonitor air pollution for a variety of reasons. But, yeah, I'm confident with the team that I have, the very excellent team that I have, and with Asa and all of your advice that we'll figure out something that's going to work.

And as I -- I think I mentioned before as well in a past meeting, we're going to try to design something that's specific. We're not going to be doing like broad biomonitoring. We don't have the resources for that, but we will definitely take into account these challenges as we design our targeted biomonitoring study.

CHAIRPERSON SCHWARZMAN: Ulrike.

PANEL MEMBER LUDERER: Yes. Thank you all for those presentations. They were all really informative and interesting. And I actually had a follow-up to the -- regarding the AB 617 study. I think Nerissa mentioned that -- you were planning on looking at some biomarkers of

effect. But I was just curious which biomarkers of effect you're thinking of measuring?

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MS. HOOVER: So same answer. We haven't determined that yet. We're obviously looking at the biomarkers of effect that are associated with air pollution in the literature. So if you have thoughts on that, we are wide open to any suggestions at this point. It's still in the early research and design phase.

CHAIRPERSON SCHWARZMAN: Yeah, Veena.

PANEL MEMBER SINGLA: Hi. Thanks for those presentations. My question was about the VOC metabolite method development. And if you could give us a reminder of what parent VOC compounds that that would be looking at and the timeline for that potential method development, and I wondered if it might be able to inform the AB 617 study at all?

DR. SHE: So for the VOC metabolite, we're looking for -- the parents could be smoking-related or organic solvent related ones. Regarding, specifically related to the AB 617, I haven't had a chance to look at a list from which one might be present. In the AB 617 is concerned list. And as I am -- but our list is basically 28 metabolites.....from -- four or five from smoking related VOCs and then a group from the solvent. That's how we come up. But....at this moment, the method

of focus on the CDC published standard operation procedure. But we always can like expand it to cover 617 concerned chemicals.

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MS. HOOVER: So I'll just chime in. This is Sara again. A couple things. The method that Jianwen just referred to, which he cited, is CDC's method. I can send you the list of the complete set of chemicals covered in that. We actually went through this with Victor De Jesús in a past meeting when we were first talking about air pollution biomonitoring. I'll also add that we've actually looked closely at the VOCs -- the parent VOCs, some of them are strongly traffic-related, like gasoline-related chemicals.

So we're actually considering -- I'm not going to say much about it, but we're considering a pilot project potentially to look at some of those stable metabolites of VOCs related to gasoline exposures. And this partially comes out of the big report that we finished on the assessment of gasoline-related exposures throughout California, the 15-year -- 18-year report I think we did. Dan Sultana in my group and I.

I can share that with you, if you're interested in it. It does talk about these issues. And so, yeah, we'll definitely be considering, you know, if we can work with Jianwen's lab, we're going to be collaborating with

him on some of those issues.

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I also wanted to chime in on a past comment and question. And Julia Varshavksy, who is listening, said that she could talk a little bit about some of the biomarkers effect -- of effect that she's been researching for 617.

So, Julia, do you want to join and give. Go ahead and share your webcam and --

CHAIRPERSON SCHWARZMAN: Hey, Sara, I'm just wondering if we could stay on this first topic for a moment and then go back to biomarkers of effect --

MS. HOOVER: Oh, sure.

CHAIRPERSON SCHWARZMAN: -- because I have a question to this one about VOCs or comment.

MS. HOOVER: Go for it. Sorry.

CHAIRPERSON SCHWARZMAN: So thank you for that comment, Veena, because I feel like it would be great to make some overlap here. And I do really appreciate that 16-year report, and making some of those connections will be great. And I just wanted to mention one more link that Jianwen might look at. This is not published yet, but we're working on -- we've been working with the California Air Resources Board consumer products database really extensively for a while trying to prioritize. So that's about consumer products used very broadly. So that's many

products that are mainly used in workplaces, but they're sold in a potentially consumer-facing way.

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And they're -- that's looking at sales volume, and also VOC, and low vapor pressure VOC content, and out of that analysis have developed some sort of priorities -- a short list of priority compounds, because of their -- either their volume in commerce or they might be lower sales volume, but higher percent in the products.

And I would be curious to kind of put our priority list, and we've prioritized them also about potency kind of against the list of potential analytes that you're looking at methods development around. And there may be great methods already and good overlap, but I would just be curious to make that connection.

DR. SHE: Great. We like to have this priority list to work with you to look for further refining and target our analytes.

CHAIRPERSON SCHWARZMAN: And we can certainly talk about it now. We're working on writing up methods and publishing it, you know, in the next few months.

DR. SHE: (Nods head.)

CHAIRPERSON SCHWARZMAN: Great. So thank you for letting me interject that into that discussion.

And then I'd love to go back to the biomarkers of effect and invite Julia Varshavsky to answer some of those

questions.

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MS. HOOVER: Great. Thank you, Meg. So, Julia, if you are online, and you can share your webcam, and unmute yourself, can you chime in?

She emailed me that she was going to do that. We'll just give her a minute.

Okay. Hang on.

CHAIRPERSON SCHWARZMAN: If anyone has another question or comment while we're waiting for Julia to get on. Yeah, Tom, please go ahead.

minute to unmute. So I actually wanted to know a little more. I'm quite interested in the halogenated carbazoles. And some of this we probably have been briefed on, but I -- it would be helpful to me if there was just a little bit of an update about, you know, sort of what the intent -- what the pathway is where we're going. I mean, the methods development looked really interesting and was, I think, very sophisticated and useful. But I'm trying to remember exactly where those fit in. I mean, they're -- they're an air -- certainly an air pollutant, but they come by other pathways too and there's fairly high volumes in some products in some workplaces. But I just wanted some more insight on where we plan to go with that, what specific efforts that's going to be included in that?

DR. SHE: Hi, Tom. Thank you for that. Is this question for me or -- sorry.

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PANEL MEMBER McKONE: Sorry, I muted it.

It probably was more a general question for the program. I don't know. I'm not sure if Sara was listening. It might be for Sara to cover about where -- you know, where carbazoles fit into the plan, and the halogenated --

MS. HOOVER: Yeah, I'm sorry. We're troubleshooting on the side, Tom, so yeah. Basically --

PANEL MEMBER McKONE: So I don't know if you heard the question. I was just trying to get more of a sense of, you know, what -- I think I understand the motivations and I know we had some discussions, but I did -- if you could just refresh us a bit on, you know, what are the next steps once we get the methods development, which looks like the methods development is moving ahead very nicely and --

MS. HOOVER: Are you specifically talking about halogenated carbazoles or something else?

PANEL MEMBER McKONE: Halogenated carbazoles.

MS. HOOVER: We have no plans to pursue halogenated carbazoles. That was a pilot that Jianwen did.

PANEL MEMBER McKONE: Okay.

MS. HOOVER: It was a laboratory study, so it's not on our radar to address those. I'm not actually sure. Maybe one of you, either June-Soo or somebody else, could say what they're used for. If you could repeat that, then I could tell you if it might already be captured in our set of chemicals.

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Jianwen, do you know what they're used for?

DR. SHE: Yes. I can give a try. Carbazoles

were found in the Great Lakes in sediment. Professor An

Li at Chicago University and others reported carbazole

levels in environment is five times higher than PBDEs.

Also, from its structure, you can see it's very similar to dibenzofuran. That means they're stable, persistent. But where are they going? They're found in environmental samples and they're found in San Francisco Bay Area in the water samples and the fish.

And then we -- we look at it basically because they're so high levels and they're persistent. And then unfortunately we use a sample is archived samples, and then Nerissa can talk about the sample we used. We look for the parents. We didn't find it. And then we talk with Anne and other group that said, oh, you may want to look for some metabolite.

So our assumption is because they persist, we should find parent in serum samples. Surprise to us we

didn't find them. So --

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MS. HOOVER: Sorry, Jianwen, let me just chime in for a second. I was asking about the sources, the exposure sources of -- not about your specific project, but where the halogenated carbazoles coming from. I see Jon is sharing his webcam. So maybe, Jon, do you have a comment in response to my question perhaps.

Jon Sobus.

DR. SOBUS: No, I don't. I was just getting prepped -- just getting prepped for the presentation. That's all.

MS. HOOVER: Okay. Great.

DR. SHE: Sara, I can answer the question quickly.

MS. HOOVER: So, Jianwen, yes, you have a comment on where they're coming from.

DR. SHE: They come -- they're coming from a byproduct from refining -- petroleum refinery(inaudible) product, because(inaudible), so the dye industry may be the major one to form the halogenated carbazoles.

MS. HOOVER: Okay. So for now, we're -- it doesn't -- it's not captured by our program. Go ahead, Tom.

PANEL MEMBER McKONE: They're also used -- I did a look. They are used in dyes in color retention. So

they might be in clothing or some consumer products --

MS. HOOVER: Okay.

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PANEL MEMBER McKONE: -- which would make them of interest, I think, to us.

MS. HOOVER: Great. So let me --

PANEL MEMBER McKONE: They may not show up as a priority, but I just thought that would be because they seem --

MS. HOOVER: I -- I'm actually --

PANEL MEMBER McKONE: -- a little bit ubiquitous and persistent.

MS. HOOVER: Yeah. My question was not to say they're not a priority. My question was at the moment, we haven't flagged them, but if you'd like to flag them as something for us to track and pursue, we can certainly do that, if that's what you're suggesting as some of your input today.

Okay. Now, I just -- I'm actually --

PANEL MEMBER McKONE: Maybe -- maybe more research on whether they should be a priority as opposed to like moving ahead with that.

MS. HOOVER: Okay. I'm actually going to invite Meg to say her comment that she just texted to me, because this is what I was going for actually, if that's true.

CHAIRPERSON SCHWARZMAN: I think -- I don't know

that much about halogenated car -- carbazoles, but what I understand is they can also come from like the environmental breakdown products essentially of halogenated flame retardants and chlorinated pesticides. I'm sure they're tracked to lots of different sources.

MS. HOOVER: So if that's true, then they are included in the -- that's kind of what I was going for, because basically we have -- as you all probably recall, we have the entire class or group of brominated and chlorinated organic compounds used as flame retardants. Anything that could be a marker for those, including an environmental breakdown product would be captured by that listing. We also have a number of chlorinated pesticides listed. So how about if we put on our to-do list, which will show up in the transcript for today, we will follow up and try to figure out whether halogenated carbazoles are already covered by our list, and if not, we'll do a little more research, as Tom has requested, to see if they should potentially be subject to like a preliminary screening. Does that sound okay with everyone?

(Nodding heads.)

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CHAIRPERSON SCHWARZMAN: Good.

MS. HOOVER: Okay. So now I'm going to hand it over to Julia. The issue was she's an attendee, so we had to unmute her in order for her to be able to speak. So go

for it Julia.

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CHAIRPERSON SCHWARZMAN: We can't hear you.

No. We can see you, but not hear you.

MS. HOOVER: Do you see the little red mic?

DR. VARSHAVSKY: (Nods head.)

MS. HOOVER: Okay. Is it turned green?

Marley, can you check if she's unmuted on your

screen?

I had this problem earlier when I was unmuted entirely and I could not speak in GoToWebinar. So there's some weird glitches happening today too. All right. How about -- well, let's see, we'll do a little more troubleshooting and see if we can get her unmuted. If you want to, in the meantime, invite other comment, Meg. You can do that.

DR. VARSHAVSKY: Can you hear me this way?

MS. HOOVER: Yes.

DR. VARSHAVSKY: Okay. Great. Sorry about that. I wasn't planning to speak, so we had to troubleshoot that. But I just wanted to reiterate what Sara said about how we're really in the information gathering stage for biomarkers of both exposure and effect related to air pollution. But I wanted to share our running list of potential biomarkers of effect, because somebody brought it up and because it would be great to get your expertise

or input along the way, so -- so that you know what we're considering. And if you have any input, we would love to hear it.

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So right now, we're looking at -- we're sort of focused on -- more on urinary measures because of the more ease of sample collection. So in that vein, we're considering oxidative stress biomarkers that have been decently well established in the literature,

8-isoprostane, 8-hydroxydeoxyguanosine, potentially paired with prostaglandin to try to get at the different lipid peroxidation versus inflamma-- inflammation pathways that may be more or less relevant for certain outcomes, like for example preterm birth.

We're also looking at biomarkers of inflammation, which as we understand it currently is more restricted to serum tissue rather than urine. And those include these panel of cytokines and/or things like plasma CRP, which has been related to air pollution as well in the literature.

We're also looking at telomere length in different kinds of human biolog -- biological tissues, so serum, and we're also exploring buccal samples as well.

And then one thing we've added to our list recently was nasal IgE antibody, which has been associated with diesel exhaust in the literature as well.

So that's sort of the rundown. We're looking at telomere length, oxidative stress, and inflammatory biomarkers. As a broad categories. So if anyone has insight or wants to talk to us -- our team about potential strategies for doing that, in an effective way, we would be more than happy to hear your input.

Thank you.

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CHAIRPERSON SCHWARZMAN: Great. Thank you so much Julia for navigating what you had to navigate to get in and tell us that. It's helpful.

It is time to move to our next talk -- Jenny is -- do you want to raise something about this and then we'll move on?

PANEL MEMBER QUINTANA: I'll be very quick. A couple comments. One was to help interpret the data, I do encourage that you measure cotinine or potentially markers of marijuana smoke exposure or vaping. It might help interpret the data, because sometimes disadvantaged communities also have a much higher rate of smoking, but also living in multi-unit apartments with neighbors who smoke. And I think it might help interpret some of the PAH data and things like that.

DR. VARSHAVSKY: Absolutely. That's great.

PANEL MEMBER QUINTANA: And the second -- second thing is if you are looking at markers of effect, we could

talk -- I can email more, but I really encourage, especially for DNA damage or things related to DNA, that you measure folate levels, if it had a huge effect on those kind of biomarkers and they -- I think the recommendation is that you always measure that if you're going to do that kind of monitoring would be the best practice.

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So thank you. I'm sorry to take the time. DR. VARSHAVSKY: Great. Thank you.

CHAIRPERSON SCHWARZMAN: Okay. So, we need to move on to our next presentation. Thank you all for your contributions to that discussion.

We're a little bit behind schedule, but I think we will make it up at some point here. I want to introduce our next speaker. We're going to hear about what EPA is doing with non-targeted analysis. So our next speaker is Jon Sobus who's a physical scientist in the U.S. EPA's Office of Research and Development, or ORD, specifically in the Center for Computational Toxicology and Exposure. And he works to develop high resolution mass spectrometry methods for characterizing contaminants of emerging concern in a variety of environmental and biological media.

He's team leader of ORD's Non-Targeted Analysis
Research Program and co-PI for EPA's Non-Targeted Analysis

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Collaborative Trial, or ENTACT. And he'll be presenting
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    findings from that trial ENTACT to us now.
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             So welcome, Jon. Thank you.
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             (Thereupon an overhead presentation was
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             presented as follows.)
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                         Terrific. Thank you for the
             DR. SOBUS:
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    introduction. Can everybody hear me okay?
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             That doesn't sound promising.
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             MS. ZALAY: Yep, we can hear you.
             MS. HOOVER: Yes. Go for it, Jon.
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             DR. SOBUS: Terrific. Can everybody see my
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   screen okay?
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             MS. HOOVER: Not yet. I think -- I think Marley
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   may need to make you the presenter, if she hasn't already
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    done so.
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             Sorry, this is Sara speaking from the...
             MS. ZALAY: Yeah. You have presenter
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    capabilities Jon. Do you see --
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             DR. SOBUS: I have shared my screen and it says I
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    am sharing.
             MS. ZALAY: Why don't -- why don't we --
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             DR. SOBUS: I'll try one more time.
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             MS. ZALAY:
                        -- try one more time.
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             Thank you.
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             PANEL MEMBER McKONE: There. Yeah.
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MS. HOOVER: And Jon, I -- Jon, I also, yes. And Jon, I want to also let you know you have your full 20 minutes, so don't worry. You don't have to rush through, because we went over.

DR. SOBUS: Terrific. And I do tend to go pretty quickly on these things. So I'll try not to go too quickly. If I can get this panel off the side of the screen, I will go into full presenter mode and we will finally be on our way.

Okay. Can you see it?

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PANEL MEMBER McKONE: Yep.

PANEL MEMBER SINGLA: Yeah, it looks great.

DR. SOBUS: Terrific. Okay. Okay. So I'm Jon Sobus. I work for the U.S. EPA. I'm going to be talking today about our most recent findings from our Non-Targeted Analysis Collaborative Trial, or ENTACT. So I wanted to first start by presenting kind of a juxtaposition between non-targeted analysis and the classical targeted analysis that we've known and used for so many decades now.

With targeted analysis, we typically know the chemicals or chemical classes that we're looking for and we build the methods specific for those chemicals or chemical classes. That's not how we do non-targeted analysis. Here, we don't pick the chemicals. We pick the samples of interest. So these can be environmental

samples, biological samples, you name it.

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We start by doing a basic laboratory preparation of those samples using liquid extraction. We often do some type of cleanup. But generally speaking, we're going to inject those process samples on a high resolution mass spectrometer. And then we are going to generate a lot of information on what we call molecular features.

Okay. So this is basically a plot of a number of different molecular features, with each feature being represented as some type of peak. We try and use some type of prioritization scheme, often using statistics, to figure out which molecular features are most interesting or most important. Then we try and propose a chemical formula for each feature. Then we try and propose a structure that corresponds to each formula. And for chemical risk assessment purposes, we might try and want to estimate some type of concentration, which is difficult when we don't have standards. And if there's a potential health issue, we might try and identify the source of that for risk mitigation purposes.

So this is just a general workflow and you're going to hear a lot of different approaches to doing NTA in the talks later this afternoon. But I wanted to point out there's at least four really good reasons to do NTA.

Number one, we can rapidly screen for known compounds. So

I think June-Soo presented a few moments ago that they screen for about 3,000 compounds. We actually screen for about 850,000 compounds right now. And some laboratories screen for in the millions of compounds. So this type of workflow allows us to do that rapid screening of knowns.

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We can also discover unknown compounds. We're going to hear about this in Dr. McCord's presentation next. These are compounds that don't exist in a database and may not even be known to exist. So this is a good mechanism for identifying those new compounds.

Also, as we heard about in one of the last presentations, we archived this data. So while we might not have the best possible methods and workflows right now, we store this data on hard drives. We can always go back in time and reassess the data and uncover historical exposures.

And finally, we can look at all this data in aggregate and we can try and identify specific source fingerprints for things like air pollution, or water contamination, or maybe even disease states. So these are four reasons that we really like doing NTA. There's many more and I'm sure you're going to hear many good examples through this afternoon's presentations.

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DR. SOBUS: So NTA has really taken off over the

course of the last say five or ten years. And there's been a lot of different commentary and viewpoint articles. I've read most of them and I found a couple quotes in these two articles particularly interesting. One quote came from an ES&T viewpoint article from Ron Hites and Karl Jobst. And they said that, "No single analytical technique is suitable for the analysis of all compounds and that successful non-targeted screening will require the development of multi-platform approaches, facilitated and validated through interlaboratory comparisons". I couldn't agree with this quote more and that's exactly what I'm going to be talking about today.

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The second article comes from Chris Higgins' group out of the Colorado School of Mines. This was published in Science of the Total Environment. And they commented that, "The novelty of non-targeted analysis, particularly its current lack of implementation by regulatory agencies, has prevented the establishment of streamlined quality assurance and quality control procedures".

So I like this quote, but I personally think it's a little bit backwards. I think it's the lack of QA/QC procedures and performance benchmarks that's caused a lack of implementation by regulatory agencies. So that's the real driver for the work that I'll be presenting on today,

trying to come up with some type of performance benchmarks in thinking about how we can get a handle on QA/QC procedures.

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DR. SOBUS: So we've been thinking about this now for over five years. And we've had several workshops at EPA. And a lot of the research that we've done and planned has focused on these five key science questions. We know that there's lots of different methods and tools for doing this type of NTA work. So how variable are the tools and how variable are the results from lab to lab.

Given that there's so many different ways of doing this, are some methods and workflows better than others? To what extent does sample complexity affect performance? Can you be really good working in one medium and perhaps not so well working in another medium? Can we evaluate and perhaps even predict the chemical space that a given method might cover? And then finally, do we have the adequate sensitivity and specificity with the current instruments and tools to identify with confidence contaminants of emerging concern?

So these five questions were the real science drives behind the ENTACT project.

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DR. SOBUS: So ENTACT is a little bit different

than many other interlaboratory trials. I think it's actually a bit complex. We've actually developed it in three parts. For part one of ENTACT, shown here on the left-hand side, we started with about 1,200 of our ToxCast chemicals. So these are chemicals that we procured in our toxicity forecaster project. There's about 5,000 chemicals in total right now. These are chemicals that are of high interest to the agency for risk purposes, either for humans or ecological species.

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Here, we took 1,200 of those 5,000 compounds, and these were the highest quality compounds. This means they were very pure and they were very stable. And we put them in ten synthetic mixtures with about a hundred to four hundred chemicals per mixture. We pulled together about 36 -- about 30 research organizations and we asked those research organizations to do a blinded analysis, using the non-targeted analysis method or methods of their choosing to try and determine what's in these samples.

Once they performed their blinded analysis and sent results back to us in a standardized template, we would unveil the chemical lists only to that group one at a time, and they would perform a final unblinded evaluation and report back to us their final findings. So this is ENTACT part one and this is really what I'm going to be speaking about today. But we realized that we have

a bit of a blind spot here, because we're working entirely in synthetic mixtures.

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So for part two of ENTACT, which I really won't have time to get into today, we're working with actual matrices. So we wanted to work with something that was well characterized using existing targeted methods. So we picked standard reference material house dust from NIST, standard reference material human serum from NIST, and then we had reference silicone wristbands made under contract with a university.

So here, we can compare the performance of a non-targeted analysis method with previous performance of targeted methods that have been used for years and reported in the literature. But we also wanted to examine things like extraction efficiency. So to do that, we took one of the synthetic mixtures and actually spiked each of these different media and then we provided laboratories with extracts of these fortified materials.

So when all is said and done, all the participating labs were given the option to get the ten mixtures, the three extracts of the reference materials, and the three extracts of the fortified reference materials. So that's ENTACT parts one and two.

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DR. SOBUS: ENTACT part three was basically, at

the time we had 4,600 ToxCast substances. We individually plated all of these substances and gave them out to a select grouping of instrument and software vendors, as well as a few select laboratories for the explicit purposes of doing MS/MS analysis or MS to the N analysis, and generating reference spectra to be made available to the public. So this is part three. It is ongoing.

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DR. SOBUS: Again, I'm going to be spending the bulk of my time talking about the mixtures today and I really wanted to spend a few moments talking about the strategy and how we designed these mixtures. So again, there are ten unique mixtures. They're numbered 499 through 508. You can see here that we have -- the first four mixtures have 95 compounds, the next two have 185 compounds, the next two have 365 compounds, and then we have these last two mixtures that I'll talk about in just a moment.

Again, we wanted to understand how the complexity of a sample affects performance. And we also wanted to understand things like reproducibility. So to get at reproducibility, we had to have a substantive compound that were spiked across many mixtures. So you can see in these tiny blue bars, there was actually five compounds that we spiked in all ten mixtures.

Then we have, what we called, Grade A replicates. This was a set of 90 compounds that were spiked in one of the mixtures with 95 compounds, one of the mixtures with 185 compounds, and one of the mixtures with 365 compounds.

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So here, we can see if the size of the mixture actually impacts performance on these compounds that were spiked across replicate mixtures. Okay. So this is starting to allow us to get at answering the complexity issue, but we wanted to make even more complex.

So we decided to include sample 507 and 508. You can see that 507 is a small mixture. It's 95 compounds, but we included lots of isomeric compounds, so compounds that share the same chemical formula, as well as isobaric compounds. These are compounds that have very, very similar monoisotopic mass. All of the first eight mixtures had very, very few isomers and isobaric compounds, whereas 507 was almost entirely isomeric and isobaric compounds.

Then we went crazy with mixture 508. This had 365 chemicals, it had tons of isomers and isobars, and it actually had lower purity compounds. So we have a suspicion that there's going to be compounds in this mixture that we did not intentionally add due to the lower purity.

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DR. SOBUS: So in addition to designing the study and managing the study, we also took part in the analysis. So we have now done several different analyses, and the work that I'm showing here is based on our first publication. So this work was entirely derived from an LC-QTOF high resolution mass spectrometer run in both positive and negative electrospray ionization.

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You can see here on the upper plot, this is what's called a bubble plot. So each little gray bubble represents an observed feature or a peak that I showed on my first slide. So the intensity of the -- the intensity of the feature or the size of the peak is actually displayed as being proportional to the size of the bubble. So the bigger the bubble, the more intense the peak. We then have the retention time of the feature on the X axis and the mass on the Y axis.

So you can see that even though we spiked 1,200 compounds, when we looked across all ten mixtures, we actually found 26,000 observed features, so 20 times more features observed than were spiked. So clearly, not all of these are real. Some are noise and artifacts and other derivative features.

So we implemented a data processing strategy.

And we tried to identify things that we believe to be noise or artifacts and things that we believe to be real.

When all was said and done, the things in gray in the middle plot are the noise or artifacts, and there was about 14,000, and the things in yellow we believe to be real. We took those 12,000 real features and compared them to the spiked substance lists and ultimately found about a thousand true positives, which leaves 11,000 other things.

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Now, we don't know if these are actually false positives or perhaps true positives that just weren't deliberately spiked, that is an unresolved question that would require much more additional analysis. But we can take the results of our investigation and we can begin to examine performance using things like a confusion matrix.

So here, we have whether the substance was spiked versus whether the substance was identified. And we can see we had a true positive rate across the ten mixtures that maximized at 65 percent and we had a false negative right -- false negative rate that got down as far as 35 percent.

So we can begin to assess performance with these two metrics, true positives and false negatives. But again, it's very difficult to get a handle on the false positives, because there may be impurities. There may be degradation products. There may be things in here that are real that are being correctly identified. But since,

we didn't put them in there deliberately, it's very hard to know whether or not they're actually there.

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And then true negatives present another tough issue. These are the things that we didn't put in that we didn't see. How do we determine what could be a true negative? Do we use a list of 5,000 compounds, 50,000 compounds, five million compounds? The size of the list is going to impact the number of true negatives.

So these two things make it very difficult to complete the true full confusion matrix.

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DR. SOBUS: So in addition to the work that we've done, as I said, there's about 30 other participants. We have six contract laboratories. We have six vendor laboratories who are analyzing the full 4,600 chemicals on multiwell plates, and then we have our general participants.

So 30 participants, very few of them are using one particular method. Many of them are using two or more methods, like we did positive and negative electrospray ionization. To date, we've gotten blinded submissions from 19 different laboratories and the full unblinded submissions from 15 laboratories.

So from the standpoint of mixtures analysis, we're about halfway there on getting the data submitted to

us. Given the timeframe and how long it's taken for these analyses to be completed, we've move forward with doing the cross-laboratory evaluation with the data in-hand.

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DR. SOBUS: So this is actually a very difficult thing to do and it takes me quite a while, even though we issued standardized templates, to process the data submitted by each individual laboratory.

The first thing that we do is I'm not trying to gauge an individual laboratory's performance. I'm trying to gauge method performance. So if a laboratory submits multiple submissions based on different methods, I'm going to treat them separately. Another thing, some laboratories submit multiple guesses per feature. I'm adamant that we get one guess per feature. So I have to do some data cleaning and ask for the best guess at the mass, formula, and compound level for each method.

Also, we've heard about the Schymanski et al. confidence levels. I have found that across many laboratories, a lot of time confidence levels are not necessarily being reported appropriately. And through discussions, we've had to revise those confidence levels on several cases.

Once I generate a clean file for each laboratory, I can then match what was submitted and cleaned against

the spiked substances at the mass, formula, and structural level. I'm here looking for three metrics. I'm looking for the number of compounds that I believe that method observed. So this is if they correctly identified the structure or correctly identified the formula when the compound was not spiked along an isomer.

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There is some evidence the compound may have been observed if we get a mass hit. But in my opinion, that's not enough evidence to support an, observation so I'm calling that -- I'm putting that in a separate bin and basically calling that undetermined for the moment. So it's observed if it was a structure match or a formula match not in the presence of a spiked isomer.

The compound was identified if it's a structure match. And the compound was reproducibly identified, if it was correctly identified at least 50 percent of the time. So this reproducibility metric is only relevant for things that were spiked more than once and correctly identified at least one time.

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DR. SOBUS: This is kind of the overall view of how the methods stack up against each other. So this is a heat map, where we've got the 1269 compounds on the Y axis, the individual methods on the X axis, and you can see here that I've grouped all the LC ESI positive methods

together. I've grouped all the ESI negative methods together. And then we have a hybrid method, which used a very different approach. And then we have a GC EI method. And you can see that the individual cells are colored purple if the compound was not observed, and yellow if the compound was observed.

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So for the ESI positive LC methods, the percent of observed compounds ranged from 42 to 69 percent. For the ESI negative, the observed compounds ranged from 21 to 39 percent. And then we have a 48 percent observation rate for the hybrid method and a 62 percent observation rate for the GI -- the GCEI method. You can see some similarities in performance when you kind of look within method grouping. The ESI positive methods do definitely have some trends. The ESI negative methods do have some trends.

But when you look at the totality of this image, you can see drastic differences in what was observed across the different methods. What's fascinating to me is to kind of look at the top purple rows. And this basically shows the number of compounds that were not observed by any method. And basically, we have five percent of the compounds overall that could not be observed by any method. So this is a fairly small number, which is -- is really good to see.

When we look across the bottom row, we can see that we have actually fewer than one percent that was observed by all 12 methods. So this is a very, very small number. When all is said and done, it was really only four compounds to date that have been observed by all methods.

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DR. SOBUS: So this is perhaps the most important slide. This is a method comparison looking at total performance across ten methods that I've fully examined to date. So there's three metrics that I want to talk about here. This is a bubble plot. So here, each individual bubble represents one method from one laboratory.

The diameter of the bubble represents the total coverage represented as a percent. So, for example, this bubble down here has a coverage of 0.69, which means they were able to observe 69 percent of the compounds. The X axis is the precision metric. This is basically how often they were able to correctly identify a compound. This is the ratio of the percent identified divided by the percent observed.

So if a particular method observed a hundred percent of the compounds and correctly identified 90 percent, the precision would be 90 percent. Likewise, if a given method observed ten percent of the compounds, and

correctly identified nine percent of the compounds, their precision would also be 90 percent. So here, I'm basically allowing precision to be compared across laboratories, irrespective of the coverage.

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The final piece is the reproducibility. This says, for chemicals that were spiked more than once and that were correctly identified at least once, how many of them were correctly identified more than half the time?

So we can look at some specific examples here. We can see, for example, in the green circle, this was laboratory seven that used the GCEI method. They had a 62 percent coverage. They had a precision of over 90 percent and they had a reproducibility of over 80 percent. So overall very strong performer.

We see similar results for lab four, LCESI positive. They had tremendous reproducibility. Slightly weaker position and slightly weaker coverage. We have two examples here, where we had good reproducibility, good precision, but you can see the coverage was at 22 percent in each case. And these were actually ESI negative methods, which is known to have less coverage.

You can also see this one example down here, where we had great coverage at 69 percent, but we had a precision of seven percent and a reproducibility of seven percent. So ultimately, we would like to see groups up

here in this upper right quadrant with as big a bubble as possible

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DR. SOBUS: So we need to think about how we give these metrics back to the participating labs and how we communicate these results to the scientific public and to the public in general. So this is something that I am proposing for the first time here today. There's basically a visual report out and then kind of a report card numerically.

So this is the same bubble plot I showed a minute ago, where we can report to a given participant their method shown in red relative to the performance of all other methods. We can simply enumerate and say you had a 95 percent precision. You had an 85 percent reproducibility and you had 62 percent coverage. But no one laboratory is going to do perfect in probably any given category.

So we want to give each lab their results in the context of the best performer in each category. So over here, just like you would get results from a medical examination or results from a GRE test, we're giving them their score relative to the maximum score. So for this particular group, their precision was 95 percent of the maximum precision, their reproducibility was 87 percent of

the maximum, and their coverage was 86 percent of the maximum. So they were not the best performer in any one category, but they were a very strong performer across all three categories.

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DR. SOBUS: So I don't have time to get into the experiments with dust today. But if you have questions afterwards or during the Panel discussion, I'm happy to address it.

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DR. SOBUS: I also don't have time to get into some of the applications with in silico spectra.

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DR. SOBUS: So I'll just wrap it up with my summary slides real quick. I hope I've shown you that NTA methods are very suitable for many ToxCast chemicals. Only five percent of the compounds to date have not been observed by any method. Multiple methods clearly are going to be required for broad characterization. There is no one-size-fits-all method. There probably never will be. And to date, fewer than one percent of the ENTACT compounds have been observed across all methods.

Because I can't complete a full confusion matrix, to date, I've examined performance across three categories. Coverage tells us the ability of that method

to observe a compound. To date, we see a range of 21 to 69 percent, which is a pretty broad range. Precision tells us the ability of the method to correctly identify compounds that have been observed. We see a massive range of 7 to 99 percent for precision. And finally reproducibility tells us the ability of the method to consistently and correctly identify the compound. And again, here we have a massive range of 7 to 97 percent.

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Although, I didn't get into it today due to timing issues, our results have shown for the SRN dust and serum that the concentration, media, and extraction techniques are going to affect performance. So while these results in synthetic mixtures are highly useful, they would be subject to change in actual media samples.

And then finally, while this cross-laboratory examination is incredibly useful, we do a lot of work with these data in-house to optimize our methods. So these data and these samples are still available for those that would be interested in testing their own methods and optimizing their methods.

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DR. SOBUS: So I'd just quickly like to thank all of the contributing researchers at EPA, all of the participating laboratories, and specifically to call out Elin Ulrich who's my co- -- my co-PI on the study, thank

her for all of her help along the way, and I'm happy to take any questions.

 $\label{eq:chargestand} \mbox{CHAIRPERSON SCHWARZMAN: Thank you so much, Jon.}$ So questions from the Panel first.

Oliver.

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PANEL MEMBER FIEHN: Thank you very much. It's very interesting. Now, as you say, the people have used different methods. Some methods are more -- have more resulting power dynamics. Like a QTOF has less resolving power than a Q Exactive. You can use ion mobility or you cannot use ion mobility. You can use two-dimensional separation or not.

Is there anything you want to say about like resolution versus precision coverage and so on?

DR. SOBUS: Yes. And I'd say it's premature to say it right now, because I basically looked at 12 methods. When all is said and done, we've got 30 laboratories, probably with a mean of two methods per lab, so that's going to be 60 data sets I have to get through. When all is said and done, I'm very comfortable we're going to be able to make recommendations like ion mobility is really helpful or resolution is really helpful.

I am starting to see trends. I don't want to necessarily speak definitively at this point, because it is a small sample set, but yes, I do believe what will

come from this -- one of many things that will from this is those hard recommendations of this is going to get you in a better place based on the performance metrics that we've laid out.

CHAIRPERSON SCHWARZMAN: Other questions from the Panel?

Tom.

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PANEL MEMBER McKONE: Thank you, Jon. That was really interesting. I'm really fascinated by the opportunity of like the non-targeted to really discover things that we weren't looking for. I guess -- the question I have is ultimately it's probably useful to know what you're looking for. And I'm kind of reminded of, there was a paper a number of years ago by Phil Howard and Derek Muir, where they went -- they just basically looked at chemicals in production and looked at chemical properties, and made estimates of chemicals that we've never put on a list that we probably should be looking for because they were persistent, you know, based on chemical property.

So it was kind a forward-looking saying this is -- and indeed, once they published the paper, people went out and looked, they did find those chemicals, because they made specific recommendations. But this idea of sort of -- sort of shining a light into saying, well,

what should be out there that we're not seeing and then doing these sort of tailoring the non-targeted analysis, the combination of the two might end up being very powerful, you know, for finding the things that we're not dealing with currently.

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DR. SOBUS: It is. And we really take that into consideration. I certainly didn't have time to get into it today, but we use a ton of molecular modeling, and exposure modeling, and pharmacokinetic modeling to try and determine compounds that are likely to be prevalent, and for which we would see higher exposures. And then we also build models to say which methods would be amenable to observing those particular compounds.

So if we screen for them on a large library and we see them as candidates, we can use that information to say this structure is potentially more likely -- based on exposure potential, this structure is probably more likely, based on the fact that its structure is amenable to the method that we're using here.

So much of what I've shown today is the tip of the iceberg in terms of what we actually use to do substance identification. But you're exactly right that this stuff for exposomics applications should not be done in a silo, thinking about mass spectrometry, it needs to consider information from the outside world.

PANEL MEMBER McKONE: Great. No, that's really good to hear. So you're not just going into the dark.

You actually have some -- some little twinkly lights that tell you where you wanted to go.

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DR. SOBUS: We try and be -- we try and be very intelligent and bring hazard data, exposure data, and use kind of ratios of the two for -- to kind of identify which chemicals are of greatest interest from a risk perspective, and then we try and, to some extent, kind of direct a non-targeted analysis towards that chemical space.

PANEL MEMBER McKONE: Really good. Thanks.

CHAIRPERSON SCHWARZMAN: Eunha.

PANEL MEMBER HOH: Eunha Hoh from San Diego State
University. Hi, Jon. I have a couple of questions
that -- it's great work. I can't imagine how much you can
present from the work. It's variable data that is
fascinating.

One question was that I was fascinated by that -the graph that you showed us the coverage, the observed
and identified. But the five percent of the compounds,
did you find them, any common things among those
compounds? And if --

DR. SOBUS: Yeah, I actually meant to. I meant to say something about that. The truth is I haven't dove

into it, because I put this together fairy quickly. But one of the things that, at least in our analyses, that showed up as being missed were the quaternary ammonium compounds that had a permanent positive charge. We typically misassign the mass, when we're using electrospray in positive or negative. And we fell victim to that, and I think several other people did, so that's one good example of a specific chemical class that people are likely to miss using LC-EIS methods.

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I think there's other similar things and then there's going to be subsets of things that either we thought they were pure and stable and they weren't or when we put them in a mixture with 364 other things, they interacted and turned into something else.

So as we -- I don't want to dig too deep right now, because we're going to keep adding results. And that number is going to keep going down. But when we finally get to that baseline number, we are going to dig deep and say why did nobody get these right?

PANEL MEMBER HOH: So second question, Jon, is more like comments. Maybe you can think -- you probably thought about that too. But, you know, the lab -- the individual lab like have specialties in certain chemicals groups as targeted analysis. They could have like more knowledge in certain chemicals probably not the other

chemicals, you know. So there might be the variability possibly to, you know, affecting the results, too.

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DR. SOBUS: Right. Right. And it's -- it's really -- there's so many layers to this that it's really -- you know, it kind of fries your brain, but your point is well taken that I've had many level one classifications submitted that were not spiked substances. So I haven't even really treated all of these other highly confident identifications that were not spiked right. But clearly, in seeing those types of results, that makes your point that different groups have different specialties.

As I go through this, I'm trying to be as fair and unbiased as possible to try and, you know, implement a universal approach. And if I get too far down and see that something that I implemented can't be universal, I have to go back and figure out something that is universal and reimplement, which I've now down four times. So it is very, very difficult to do.

PANEL MEMBER HOH: Um-hmm.

CHAIRPERSON SCHWARZMAN: John I had just one question and then we're -- we'll need to move on, which is I was really -- my ears kind of perked up at the -- your first slide, one of the four points about what is so useful about non-targeted analysis is the ability to look at historical exposures or make comparisons within

historical exposures.

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And I was just really struck by the potential for doing that and -- and whether doing this kind of analysis and storing the data might ultimately -- maybe not now, but ultimately make it possible not to even have to store the matrices.

DR. SOBUS: That's an interesting thought.

That's an interesting thought. So I would say a couple things on that. One is we have research centers that run thousands of samples, if not per month, per year, and they can only go so deep in the analysis, and that data gets stored.

Two, we do analyses very quickly. To the extent that those methods are solid and the data are collected with care and stored with care, that's a great point, that if -- if we were to come out of something like ENTACT and say this number of methods gives you appropriate coverage to get 95 percent of the chemical universe of interest for these types of applications, conceivably you could run that number of methods, run it well, have QHX store the data, and then not have to store the samples. That is conceivable.

CHAIRPERSON SCHWARZMAN: Thank you for entertaining the idea. Thank you so much for your presentation. We really appreciate it and we're going to

move on to our next presenter.

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So I want to introduce James McCord, who's a chemist in the Multimedia Methods Branch of the Center for Environmental Measurement and Modeling at U.S. EPA. His research uses non-targeted analysis and high resolution mass spectrometry to identify and characterize novel and emerging contaminants. He will be presenting on "Multimedia Exploration of Emerging PFAS and Their Sources".

(Thereupon an overhead presentation was presented as follows.)

DR. McCORD: Thank you. Thank you. Can everyone see?

CHAIRPERSON SCHWARZMAN: Yep.

DR. McCORD: All right. Excellent. So thanks for the introduction. As explained, I'll be presenting on the multimedia exploration of emerging PFAS and sources.

So I am applying non-targeted analysis as part of our broader PFAS efforts. So non-targeted analysis is, in many cases, sort of the beginning or one small part of large biomonitoring and exposure assessment activities that we're doing in association with states and regions.

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DR. McCORD: So historically, PFAS usage has been investigated by targeted LC-MS techniques. And there has

been a historical effort to ban and replace certain PFAS chemicals species with new replacement compounds with a variety of different chemistries. And non-targeted analysis allows us to expand beyond the simple targeted historical list to identify a lot of these replacement compounds as Jon just described.

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DR. McCORD: So what I mean by this, if you're looking at things like PFOS and PFOA, whose structures are shown here, these are some of the most historically prominent PFOS -- or PFAS and they've been banned in their primary application. In fluoropolymer manufacturing, PFOA has been replaced by a series of compounds. In metal plating, where PFOS was widely described, it's also been replaced.

Many of these are still perfluorinated compounds, but they introduce novel chemical moieties. In some places, fluorines are removed or replaced by hydrogens. In others, they're introducing ether linkages that are designed to make the compounds less persistent, and then they're also adding other halogen species for a variety of reasons that are discussable.

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DR. McCORD: So this, within the context of our beginning discussion, gives us a driving research question

for a lot of states and other regions, which is, in places where there's historical PFAS usage, are we seeing replacement compounds present the same sort of distribution profiles and how can we identify and monitor these compounds to help with source attribution and to sort of set the stage for follow biomonitoring and toxicity assessment.

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DR. McCORD: So to somewhat rehash what Jon went through, we see this as being a tiered approach to chemical measurement. So the classic tried and true methods are the use of targeted approaches for selected chemicals with available reference data and chemical standards. This is a straightforward method, if not always simple, but you can get good quantitative information on a list of analytes. Normally, this is using some sort of targeted LC-MS triple quad type approach.

There's also screening approaches, which have been discussed by a number of people today, when you have a very large or smaller chemical library and you apply general non-targeted type approaches and then you screen against a reference library. In this case, you're still somewhat limited by the reference library that you have and how much data is available in those reference

libraries, whether it's structural, or just a chemical formula, or sometimes MS/MS data, but you're not going to be making assumptions about exactly what you're looking for before you process the sample.

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The majority of my work is done in the sort of discovery space. So we're looking for the things that, in one of the presentations, discussed earlier the large unidentified screening space, where sizable fraction of the chemical features that are identified have no known structure or no confirmed structure. So in many cases, we're working with things that are not described at all, don't exist in reference libraries and there may be one or two references in patent literature or no references at all in order to identify that chemical feature.

So our workflow is very similar to what screening does and what Jon just described.

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DR. McCORD: So we begin with the data generation, collecting non-targeted data on normally a high res instrument. We do molecular feature extraction to identify individual molecular features. Here's a whole bunch of chromatograms stacked up on top of each other. You can pick one. You can, using high res instrumentation, do formula assignment. And from a particular formula, you can begin to either draw chemical

structures from databases or your own estimation. For example, here, we have a bunch of different compounds that all share the came chemical formula.

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And then using a list of tentative structures, you can do structural confirmation using MS/MS, MS to the third, interesting gas phase experiments, depending on your instrumentation. Then once you have a structure, you can start to think about quantitation.

Typically, for non-targeted analysis quantitation is relative. Sometimes we will do estimated absolute quantification using non-matched standards, which is always a little bit sketchy in terms of analytical reproducibility.

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DR. McCORD: So I'll talk about a few case studies. So historically, the Cape Fear River of North Carolina, which is very close to where I live and work, is a place where PFAS has been known to be a major contaminant. So this watershed affects roughly five million residents, including the City of Wilmington, North Carolina, down here at the end of the Cape Fear.

There's a couple of rivers that make it up. And this study going back to 2007 did, as I said, targeted analysis of the underlying watershed. So they're looking for a series of legacy compounds, both carboxylic acid and

the sulfonates, sampling all of the major watersheds, and identifying hot spots for PFAS levels.

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So in the Haw River in the northern part of the watershed, there is high levels of C10, 9, and 8. Towards the middle of the Cape Fear, there's a spike of C7, a little bit of C6. And then on the Little River, there's an elevated level of PFAS. And if you start to investigate the sources for all of these, they could be assigned to historical manufacturing in the Haw River and biosolids application. This location at five in the Cape Fear is associated with fluoropolymer manufacturing. And then the Little River is down water stream of both an airport and a military base. So there's historical AFFF usage in an airport.

So this is sort of how you map out historical contamination using targeted approaches. So the follow-up to that is to do non-targeted analysis and try to identify what replacement compounds or other things there might be.

So this paper in 2015 was using a TOF instrument, so a somewhat older high res instrument by just doing sampling down the river and then comparing the appearance of chemical features between different samples to assign them to a source that occurs in the middle.

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DR. McCORD: So if you sample upstream and

downstream of a factory, and you identify pronounced peaks in the downstream sample that are in the upstream sample, you can start to assign them to a source.

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So you collect a bunch of surface water, do your sample preparation, do your feature extraction as described, and in this particular instance, we were able to identify one compound with a reference standard, which is somewhat unique. This is GenX, which was a chemical that's sold by the Chemours company. And there were a number of other chemicals that didn't have any reference data or any compounds that could be used to quantify them.

Most of them, mono and polyethers, as well as some sulfonates with structures that kind of look like this. So they're perfluorinated compounds with the variations that I described in an earlier slide.

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DR. McCORD: So to follow up on that, we look at not just the service water, but also the drinking water. So here, we have three drinking water facilities along the Cape Fear River, where they're drawing their intake from the Cape Fear, and if you do targeted analysis on legacy compounds and then also the single individual compound that we were able to obtain a reference standard for, you can see upstream of the Cape Fear where you have historical contamination of legacy PFAS they're somewhat

high. But then as you move downstream, the legacy contaminants drop off and the emerging contaminants drastically outpace them.

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So this is the only compound that we could do targeted analysis for at the time. And if you do non-targeted analysis on the same sample and you compare raw spectral abundance, which is a very loose approximation of chemical concentration, you can see that very abundant targeted compound is the minority of the chemical abundance compared to other emerging PFAS in these drinking water facilities at places that are affected by fluorochemical manufacturing.

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DR. McCORD: So the State of North Carolina started an extremely intense investigation of this compound, which sort of became the face for it. It was involving both the Department of Environmental Quality and the Department of Human Health. We were brought in as sort of the technical support to do a lot of the sampling and analytical work associated with that investigation. This is both targeted and non-targeted experiments as part of an ongoing study.

So the very first thing that they did was to shut off the outfall from the specific GenX manufacturing line and then we started doing repeated continuous sampling

over the course of the next several months, developed some methods, and so on. And it's still undergoing litigation.

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DR. McCORD: So what we were able to do was, using MS/MS, we could assign structures to a number of unique chemicals that were being emitted. And it turns out that the company was aware of these chemicals and even had names for a lot of them. So some of the major versions are shown here. These are just the most abundant ones from each of the different chemical classes. And then our sampling over time, even though we can't quantify any of these species originally, we can just track their abundance in non-targeted analysis. And we could show that when you shut off one particular manufacturing line, certain chemicals would drop off, but others would either stay constant or even spike as they're changing over manufacturing lines.

And ultimately, the State decided that they needed to start sending all of their outfall to another location, because every single manufacturing process that they were performing was emitting PFAS into the river. So to reference something that Meg had asked earlier, we actually did have historical sampling from data that was collected as far as back 2010 that had been done with non-targeted analysis, and we were able to see some of

these compounds going back well before this sample, just from looking at the raw data. No stored samples.

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DR. McCORD: So following the analysis of those samples and the identification of a lot of those compounds, NC State led a collection for a bunch of serum asking if these emerging PFAS were in the exposed population in Wilmington. They were able to get 344 participants total, about 44 of whom were able to provide multiple samples. And eventually, the State was able to pressure the company in order to provide analytes so that we could do targeted analysis.

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DR. McCORD: So just doing screening on the human serum, we were able to identify three of the emerging contaminants. They were in the river water. And because we were able to obtain standards, we could quantify for them and show decreasing serum levels after the emissions were shut off.

The half-lives for many of these compounds are shorter than things like PFOA and PFOS, but still quite long, months to years.

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DR. McCORD: So in total, Chemours eventually provided the 12 novel compound standards that they had

names for to us, NC State, DEQ. And they were able to identify a drinking water target for GenX specifically based on some reference data, both in Europe and in America. NCDHHS performed that analysis. And there's a reference if anyone is interested. And there's a current consent order in Chemours where they monitor the list of compounds that were identified in Mark Strynar and my follow-up non-targeted list showing the major compounds are reduced in their permission -- in their emissions.

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And then they've installed air emissions controls as well, because we only examined water, but there is the belief that the air is providing a widespread contamination of the immediate surrounds. So that's ongoing, but it involves non-targeted analysis.

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DR. McCORD: In New Jersey, we have a similar sort of case study. So New Jersey has historical contamination of PFOA and PFNA, which are drastically higher than other places in the nation, based on UCMR data and their own studies. And it's believed to be one of the most PFNA contaminated locations in the world, because of historical manufacturing and West Deptford. So there's both air and water contamination that they performed studies on. And because they've identified legacy PFAS, we follow up with non-targeted analysis.

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DR. McCORD: So we did a non-targeted analysis multimedia sampling campaign. So in this region of south -- southwestern New Jersey, we collected soil, surface water, and well water. DEP was interested in knowing whether there was replacement PFAS, and if we could identify legacy and emerging PFAS with their fingerprints that could be used to assign a specific source.

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DR. McCORD: So we were able to identify a bunch of legacy compounds, which I won't talk about, but we did find a novel emerging contaminant, which is shown here. Structurally, we were able to identify it based on MS/MS. This is an MS/MS spectrum. Each of the compounds is a fragment of that parent molecule in the upper right, shown here with their accurate mass and the nice chlorination pattern, which indicates that they're halogenated.

And we are actually able to identify a whole family of these compounds, because it's produced by a polymer manufacturing process. So they're perfluorochloro polyether carboxylic acids, so we call CLPFECAs, and then they have a variety of internal linkers here. So they were seen in both soil and water in a mix of polymers. And we see both the chlorinated and the de-chlorinated

analogs.

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Like in North Carolina, we're unable to do quantitation. So in the domains where they asked us to give the estimated concentrations, we quantify it as PFNA, because it's something that is regulated, they have a standard for it, and their predicted physiochemical properties are fairly similar. So all the figures that I'll show are either estimated concentrations or they're just based on raw NTA abundance in the mass spectrometer, which again is just a loose parallel for concentration.

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DR. McCORD: So in the water, we are able to show that 99 percent of the abundance and the estimated concentration is associated with a single isomer. And it's found in both the tidally influenced surface water, which is everything that connects to the Delaware River. Because it's tidally influenced, you get back-flushing up even minor tributaries, but that there's also groundwater contamination in a lot of these sampling locations. And their exact locations don't matter very much. But the general trend is that things that are close to the factories that are using this compound are highly contaminated.

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DR. McCORD: In the soil, we see a very distinct

trend. So if you quantify and estimate the concentration of all of these different compounds in soil and then you do some GIS based mapping and modeling, you can show this almost bullseye pattern around a specific fluorochemical user in the location.

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So Chemours was the factory in North Carolina that was responsible for most of its contamination, here Solvay is using a different product that we were able to associate very specifically with them. And we have reason to believe that it's emitted on an air basis, or at least this pattern is from air deposition, even though there's a substantial amount of effluent that's going directly into the Delaware River and impacting it.

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DR. McCORD: So just like in North Carolina, because there's groundwater contamination, they're interested in human exposures. So because there are PFNA contaminant levels that are regulated in the state, many of the people who live in the neighborhoods and areas around these factories, they have these point-of-entry treatment systems for water treatment of drinking water.

So they have these two-stage POET systems, where it's either granular activated carbon or an ion exchange resin. And we were able to do sampling in 2019 of both the ground influent water that's going into these POET

systems, as well as the effluent water that's being delivered to drinking. And we could show that PFNA is reduced by the anticipated amount. It's not 95 percent or more.

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And then doing non-targeted analysis you can show that the abundance or estimated concentration at these influent steps decreases a similar amount. So the same treatment technologies that have been applied for PFNA in this region also seem to be good at reducing the levels of the emerging contaminants.

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DR. McCORD: So in this particular instance, we were able to show that those in-place treatment systems are protective for both PFNA and the emerging compounds. And there's ongoing litigation to try to obtain similar stock materials for quantification -- real quantification, like was done in North Carolina.

There's also litigation about the level of liability for cleanup, and the actual amount of emissions, and a number of other legal issues that's still ongoing.

Unlike in North Carolina, we haven't yet been able to institute any biomonitoring. However, it's been discussed that it will be added to an ongoing serum monitoring that's being proposed in that region. And it would also be a non-targeted serum screen.

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DR. McCORD: So overall, what I hope you've taken away from this is that non-targeted analysis allows us to investigate all sorts of different media. I've shown soil, water, serum. We've also done air and other matrices, and that it's critical in the discovery of emerging contaminants, as well as the characterization of things that aren't located in reference libraries. The non-targeted data can support early stage monitoring by identifying potential targets for follow-up experiments.

And even when it doesn't provide absolute quantification, you can still do relative quantification experiments that can be help you track things like treatment of water, or the effectiveness of air handlers.

However, ultimately, due to the current regulatory environment, chemicals standards and absolute quantification are necessary. And if you don't have a standard, it can be very difficult to perform risk assessment, unless Jon solves the problem.

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DR. McCORD: So I have to acknowledge all of the State and regional partners who are responsible for getting us all the samples and helping us set up all these studies.

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DR. McCORD: And then I will end at approximately at the same time as my one minute warning.

CHAIRPERSON SCHWARZMAN: Thank you so much,

James. Yeah. And we have permission to slightly shorten
the lunch break, so that we make sure we get your
questions answered. So questions from the Panelists for
James.

Jenny.

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PANEL MEMBER QUINTANA: Hi. Thank you for that really great presentation and showing the power of what chemical analysis can do to really clean up, in effect, real-world problems. Thank you for that.

I had a question about the biomonitoring and how you felt the results communication went to the population that was biomonitored and how people interpreted their results, and do people vary in how they reacted to them, or just that whole experience that you had.

Thank you.

DR. McCORD: So in the case of the biomonitoring, there was a packet that went out, provided people with as much context as they felt was appropriate. So we tried to show, in the case of these chemicals, there's no health effects data associated with them. So we are trying to show people what their levels look like relative to the whole population.

There were a number of community meetings.

People are always unclear on how to handle data like this when it's inherently uncertain. And so most of the discussion sort of came back to we can't say very much about a lot of the emerging contaminants. And I don't necessarily think that inhabitants of Wilmington, North Carolina, where we did the study are particularly pleased with the amount of information that they've been given.

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There's always a lag in the actual like risk assessment for these emerging contaminants compared to their detection and even our ability to monitor them, because the chemistry progresses faster than you can do biology studies. So for GenX specifically, we were in a slightly better position, because it wasn't detected in anyone's serum. And it's the one that there is actual health effects data for. These emerging contaminants were kind of something that no one was expecting when they got their results back initially. So there's not as much community pressure about them, because they sort of came out of nowhere.

People were expecting to see GenX in their blood and to be upset about that. But knowing that there are other things in their blood and that the levels are decreasing, there was not as much of a -- sort of an uproar about it.

We are, both at EPA, and at the State level in North Carolina doing the follow-up risk assessment on the other emerging contaminants. But to a certain extent, the cleanup is already being done and it's going to be a very long process before we really know what the long-term effects are to know how to explain to people what their results actually meant or if there are any long-term consequences for them. So it's a -- sort of a fraught situation. I'm not sure if I can give any strong answers beyond that.

PANEL MEMBER QUINTANA: Well, you're at the frontlines of how to do this, so thank you to -- thank you very much.

CHAIRPERSON SCHWARZMAN: Tom.

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PANEL MEMBER McKONE: Thanks. That was a really fascinating presentation, very interesting. I guess the question I have is about the portability of the whole process. I mean, admittedly this was like a -- I mean, the whole thing was a wonderful learning experience, but I'm expecting that eventually people at EPA and other places are going to say, oh, this is great. Can you do this for us, you know, Boston at the Charles River, or Midland, Michigan? I mean, you could name dozens of places where people would like to start applying this sort of more comprehensive analysis to really answer questions

about what's -- what's in the water and what's getting into people. So my question is do you think this will be -- like you can set up a guide book of protocols, so that it can be transported elsewhere?

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DR. McCORD: So we have a team in RTP where I work that's exclusively dedicated to this type of system for PFAS analysis in particular. And we have about seven different states where we're doing this sort of thing. And it's, you know, Michigan, and New York, and West Virginia, and all these places. And there's more sort of queuing up all the time.

My hope is that we can develop a guide book where this could be applied in any state or area where people are interested in doing non-targeted analysis to really do this type of thing. I sort of infamously, when I had first joined the agency, in a conference call said that you could apply this technique to essentially any outfall for any industry and find something probably that was not on anyone's radar that they were exposing the sort of local area to, and so I would never be out of a job.

But we actually tried to move this into the State's arena. Here in North Carolina we tried to do some technology and methods transfer to the State laboratory. We helped them write a PR to get a QTOF to do the analysis themselves. We offered to like ship me over there for a

few weeks to train them and teach them how to do non-targeted analysis. And it was initially sort of tentatively approved by the State legislature, and then ultimately shot down, because it was deemed to be too aggressive of an environmental monitoring system.

So I don't know how much broad scale interest there is in applying these really like aggressive cutting edge monitoring approaches nationwide, but I definitely support the use of techniques like this for interested parties, if they are able to support the actual scientific effort to do so. And we have tried it in the past and we're willing to support it going into the future. That's sort of my -- my hope.

CHAIRPERSON SCHWARZMAN: Great.

Oliver.

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PANEL MEMBER FIEHN: So before we had another interesting presentation that said, well, the problem is two-fold, we don't have these qualification criteria and performance criteria to know if a laboratory or a method is really good in non-targeted analyses, right?

So if you now say, you know, you would like to see that non-targeted analysis would be used in other states, what is your take on guide -- guidelines or qualification criteria, accreditation almost for non-targeted analyses?

DR. McCORD: So it depends on what your end goal is in the approach that you're applying what sort of QA and QC you need to apply. So I think Jon is trying to solve a much harder problem.

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So in our case, in this particular instance, we're associated with a class that's high visibility, high interest, and people are generally of the opinion that anything that falls into the PFAS category is something that's worth following up on.

So we have the easy job of focusing on a relatively-ease-to-characterize class of chemicals and identifying novel compounds, which is difficult from an analytical perspective to do the like elucidation and everything. But once you do an identification, everything that falls out of that basically shunts back into the standard workflow. You get standards. You do risk assessment. You do quantitation. And the discovery is the sort of hard part.

In the case of doing non-targeted analysis, generally like Jon is saying, I think that there is a need to focus on like he says the accreditation and being sure that what you are finding is defensible. And I think right now in a lot of cases, just any consistent approach applied broadly is better than an inconsistent approach. So if you come up with some process of doing validation

and you can sort of show your work, then that's a good starting point and it's a point where you can build from.

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So just the fact that people can use, for example, the Schymanski Scale to talk about the confidence of their identifications and then store their data, I think that's a good starting place, because like Meg said as well, if you've done the analysis and you've assigned confidence and made identifications, you can always go back and improve them later.

So as long as you have a good handle on the limits of your methodology, going forward that data is always useful. So collecting the data, even if the results that come out initially aren't always optimum, you at least are in a starting point and you can improve over time.

CHAIRPERSON SCHWARZMAN: Thank you so much for that. Veena, I know you had a question. We need to break for lunch. And we have a discussion section later this afternoon. And so I want -- I just have note that you have something you want to add. And if it's a question for James, we can return to this. That's no problem.

Does anyone else want to get on the queue for later. Carl, you had one. Okay. And Ulrike. Okay. Great. So this will start off our discussion after the afternoon presentations. And I have the queue.

Thank you so much, James, for the very interesting presentation and we'll obviously return to it a little bit later, when we have a discussion.

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So I want to wrap-up the morning session.

There's also no -- sorry just to note that there were no audience questions also, that we checked for those.

Okay. So before we break for lunch, I just need to provide the usual informal Bagley-Keene reminder, which is that as for the -- directed at the Panel members, please comply with Bagley-Keene requirements and refrain from discussing Panel business during lunch or the afternoon break. We'll break for lunch now. We reconvene at 1:30. But I want to make sure that everybody is aware of returning no later than 1:25, so that the technical issues can be resolved and we can start right at 1:30.

Thank you very much to all our morning presenters and the staff who have been keeping this going and we'll come back at 1:30

PANEL MEMBER McKONE: Quick question.

CHAIRPERSON SCHWARZMAN: Yes.

PANE MEMBER McKONE: So my computer will probably shut off, because it goes to sleep. And I -- can we re-sign in the same way we did this morning? It's a technical question.

CHAIRPERSON SCHWARZMAN: Technical question of

staff.

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PANEL MEMBER McKONE: I assume I'm going to have to sign out. I mean, my computer won't likely keep this open, unless I sit here and keep it from shutting down.

MS. HOOVER: This is Sara. I can tell you from my own experience earlier when no one could hear me, I actually shut everything down, re-signed in using the same link and it was fine. So my understanding is that during the session, the links will remain live. I'll just pause for a second to see if Elizabeth or Marley want to comment on this. I think that that is the case.

MS. ZALAY: You'll be able to rejoin, if you have to restart or reboot from sleep mode.

MS. HOOVER: Thank you. Thank you, both Elizabeth and Marley who have just confirmed that that is the case. So please feel free to sign-off and rejoin at 1:25. Thank you very much.

Thanks.

MS. HOOVER: Okay. Bye.

(Off record: 12:42 p.m.)

(Thereupon a lunch break was taken.)

AFTERNOON SESSION

(On record: 1:30 p.m.)

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CHAIRPERSON SCHWARZMAN: Okay. We're going to start the afternoon session. And I will just begin by introducing the next speaker.

Dinesh Barupal is an assistant professor in the Department of Environmental Medicine and Public Health at the Icahn School of Medicine at Mount Sinai, New York. He's leading the Integrated Data Science Laboratory for Metabolomics and Exposomics. His research focuses on developing, integrating, and implementing novel computational methods for metabolic epidemiology, computational metabolic -- metabolomics, and chemical text mining, blood exposome, and metabolic bioinformatics.

Dinesh will present on, "Data Science and Chemoinformatics Tools to Support Exposomics and Metabolomics".

(Thereupon an overhead presentation was Presented as follows.)

DR. BARUPAL: Thank you, Meg. So I want to say some -- a few discussant points, and ideas, and data science for metabolomics and exposomics that could be interesting for the Biomonitoring Program.

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DR. BARUPAL: So I will cover three topics in

this discussion. One is what are the opportunities in non-targeted analysis with a focus on data science; chemical to publication mapping. You know, build a resource for chemical to publication mapping; and prioritization some more chemicals for hazard assessment.

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DR. BARUPAL: So opportunities in non-targeted analysis.

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DR. BARUPAL: I see NTA as the great potential in disease prevention strategies. We have -- on the left side, we have primary preventions that we're interested in on risk factors. And as you go towards the right side, secondary and tertiary prevention, where we're interested in the -- in the tertiary prevention, we're interested in finding new biological pathways or metabolomic reactions....that can be targeted by therapeutic options.

And if we fill in exposomics and metabolomics

NTAs, we can see that exposomics fits more on the

risk-factor side and prevention. And metabolomics fits

more on tertiary prevention, where we're interested more

in internal biochemistry and metabolomic pathways that can
be tinkered by drug targets.

And the question is how do we prioritize

different NTA assays? So as you know, that a combination of different chemistry or computational approaches, we have a battery of NTA assays that can be explored. And we need -- that is a need to prioritize that which assay -- NTA assay fits best for identification of risk factor in exposomics view, or which NTA assay fits best for identification of new metabolomic pathways when we want to have tertiary prevention strategies.

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We can also think, and this -- in a single prevalence way, that in genomics, a rare variance carries more risk. We see this often in common diseases. We have a similar situation for NTA assays, that sample frequency, as we go more from lower to higher, we are capturing more internal biochemistry and metabolic pathways. And it also more fit with metabolomics assays. Whereas, if you go on the lower side of the -- on the lower sample frequency or single prevalence, we are starting recovering exposures or fine, or precise sub-type, or group of population they're exposed to only specific type of chemicals.

And this way, exposomics fits on covering the chemicals that may occur with the low frequency, whereas metabolomics fits more on that. We cover more and more component of high prevalence.

In my view, that we should avoid just holding single prevalence when we generate metabolomic NTA

matrices. We see this all of a sudden during a quality control process or during the element generation that our thresholds applies on data matrices that ignore all the compound, if they don't occur in 50 percent sample or 10 percent of samples. I think those signals thresholds should be avoided and dropped for NTA assays. All the signals that we detect, even in one sample, that should be stored in a proper way in a database, so we can mine them, the way we mine electronic....records.

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DR. BARUPAL: And that can be achieved if we start indexing raw LC-MS and GC-MS data into enterprise databases. So enterprise database system -- enterprise database systems need to be developed for indexing raw spectral data. And once we have that all the data, whatever -- even if it takes from one terabyte to going from 100 terabyte. Because enterprise databases have original scalability, we can index all the data, and this way we can query directly on the database. We can generate tables directly through the databases, exactly the same way we mine literature data, chemical data, or electronic health data in a medical system. And these are all -- it's stored in an enterprise database system.

And NTA assays that needs improvement. Well, we need to work on indexing raw spectral data into enterprise

databases. That way, we will have better tables and we can do better analytics to find risk factors as well as finding new metabolomic pathways.

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The other serious issue we have, the annotation of LC-MS and GC-MS peaks is we know that compounds still remain unknowns. My view is we need an integrated holistic approach. So if we focus only on expanding mass spectral library, we will have a increment of progress that only small personal chemicals will be annotated.

However, if we combine the best innovation in all these four domain, like instrument, sample processing, andin workflow in mass spectral library over progress multiplies and so we cannot see good progress. If we combine all these four domains and the best innovation, the best ideas in a logical way, and then we have a notation approach, that will have a better -- more impact and that way we can annotate a lot of peaks that have MS/MS spectra. And also, if we have better instruments, we can get a lot of MS/MS spectra for peaks which lack MS/MS spectra, then the issue will be that how do we rank experimental or in silica evidences for peak annotations? These are open-ended questions that need more discussion when we process data or when we design NTA studies these.

I'll jump to the biological interpretation part.

NTA data cover way more number of compounds. And these

days, they can easily have 1,500 to 2,000 compounds. And it has been absorbed regularly that many of these compounds are not covered in current biochemical databases such as KEGG, Reactome, or MetaCyc. That creates a problem when bioinformatics or statistical approaches that you utilizes -- that were adopted from genomics and utilizes a hypergeometric test that depends on a background database. So in genomics this is clear, the total number of genes we know.

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But for metabolomics this is not clear. And quite frankly, a background database does not exist for NTA. And if we use existing bioinformatics approaches that use a hypergeometric test, we're introducing interpretational bioassay for NTA data. And interpretation is focused to only compound that are covered in the background database.

Assuming the statistical independence of chemical is false, it's false solution. And in applying FDR on raw p-value for individual chemical, and for metabolomics and exposomic data sets, I think it's inappropriate, because we do know that there are mixtures that are chemical groups and there are metabolomic pathways. And there's a correlation between these metabolite group. So it is inappropriate to apply FDR or by assuming the statistical independence on unusual metabolite level.

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DR. BARUPAL: One way to visualize NTA data is in a chemical similarity graph. So idea is that we have two chemical structures, xanthine and hypoxanthine as shown here. They're 90 percent similar. We could computationally map them using a coefficient called Tanimoto Similarity Coefficient that uses a -- utilizes a substructure decomposition metrics.

That similarity data can be visualized in Cytoscape software and we can overlay statistical results. We have used those natural graph in several studies that works great for 100 to 200 chemicals. However, today's NTA data can have more than that, so -- and I seem -- we can easily have thousand compound in non-targeted metabolomics data set. And it become harder to visualize that many chemicals in a single network plot and become even harder that how do we interpret those large-scale network visualization.

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DR. BARUPAL: So other approach that we developed is called ChemRICH, which uses medical subject ontology. It takes chemical structure by chemical detected in NTA data. And then it uses Tanimoto similarity coefficient formula to map them to MeSH ontology....

So in this plot, we have lipophilicity on X axis

that chemicals groups that are polar are shown on the left side, non-polar are shown on the right side. And on Y axis we low -- a negative log or p-value that we get from Kolmogorov -Smirnov test that is done on set level one using observed p-value for assay and comparing against a P uniform -- uniformed distribution of p-value. That way, we can rank the sets by their association strength.

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And I was reading this paper last week by a biomonitoring program identifying chemical groups for biomonitoring in EHP. I think....will be a great tool to be used for NTA data coming from biomonitoring studies.

And that way we -- it can prioritize and show the chemical classes that can be put into prioritizing some process.

That's still an open-ended question that how do we include unidentified metabolite into chemical set analysis. So this is all non-compound. We know the chemical structures, but I would include unknown compound, which is a large majority of peaks are still unharmed.

There are ideas like substructure or correlation modules or correlation modules that can be explored to include those unknown -- include those unknown in the chemical set analysis.

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DR. BARUPAL: So in summary, we have several well known issues with NTA data processing. That a large

number of signals remain unknown. We have a slow signal processing for large batch of samples. So whenever we're trying to process 1,000 or more number of samples, it's hard to -- it gets really slow. And then we have errors in peak grouping and deconvolution. We have correction of retention time drifts, for batch to batch. The presence of missing values are there. Low frequency signals are often ignored. We have presence of artifacts and background signals, issues with data normalization, challenging biological interpretation. There are ethical issues in data sharing for sensitive analytes such as illicit drugs.

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So these are well-known issues. We just need to know them. And there are ways to overcome several of them. But this need to be addressed and bring to the table when we're trying to process the data, as well as when we're trying to design -- do a new NTA study.

So those were my thoughts on NTA, and what new opportunities we can have, and what new ideas we can bring in for processing data and interpretation and what are the challenges on it.

I'll switch to now chemical to literature mapping.

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DR. BARUPAL: Why -- we need a resource in a

public domain where chemicals are connected to publication in an electronic way -- electronic way.

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So we report chemical names in a different section of our paper, like abstract, tables, figure, and paragraph, and supplemented data, and again converted to PDF or in XML, and they can have in a database. But there is no table where that this chemical was reported in this literature in public domain. So we wanted to have this resource, because it will allow us to build next generation tools for metabolomics data interpretation, as well as chemical prioritization. Because right now, the approach is we take one chemical and then we do -- run manual queries and it gets laborious. And if you have -- if you detect 1,700, 2,000 identified compounds, we're talking about easily million paper in -- to review for a -- just a single study. So to over -- to make it efficient, we need a chemical to literature mapping.

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DR. BARUPAL: We undertook such exercise, where we mined one million publications on measurement of a chemical in a blood specimen. And we learned that we could have almost 42,000 two-dimensional structures. And out of those, at least 15,000 have five or more number of publications. And those 15,000 can be utilized for expending mass spectral libraries as well as to prioritize

candidate structure for peak annotations.

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potential for non-targeted metabolome studies for blood to

expand the blood exposome database. So in 2008, there

DR. BARUPAL: And there is a -- there's a great

were 150 identified compounds a typical metabolomics data set generated by Metabolon Company. And now, there is a 12-fold increase that they can generate data sets with 1,700 identified compounds. Those are all known targeted analysis.

All those identified compound goes to this subset

D, which is NTA metabolomics studies added 1,250 specific compounds to the blood exposome database. There is a great potential for NTA assays in this approach. We need to make sure that existing mass spectral libraries have all these compounds in the public domain, so other people can use it.

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DR. BARUPAL: Now, it's chemical prioritization for hazard assessment.

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DR. BARUPAL: Why chemicals?

Because most exposures are for individual chemicals, whether they come from diet, drugs, or water pollution, or other sources. In fact, IARC monograph,

which is one of the best character -- characterized exposure database. Eighty-two percent are individual chemicals. And there are mechanisms in place to identify, monitor, and regulate exposure to a specific chemical.

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DR. BARUPAL: IARC monograph (inaudible) it will weighting evidences and based on -- based on a systematic review of the evidence by the working group a classification scheme for in this is proposed. For example, Group 1 has 120 agents.

And we took exercise to prioritize pesticide structure using chemical similarity, chemoinformatics, and text mining. So in this plot, we see organophosphorus on the upper panel, organochlorine in the bottom panel. And the size represent the number of (inaudible) papers on pesticide and cancer, and thickness of the blood -- this dark represent number of papers on cancer epidemiology.

So this way we can see that in organophosphorus which chemical pest -- which pesticides have literature evidence availability on cancer, epidemiology. And that lead to IARC Monograph meeting 112 and 133 on organophosphorus and organochlorine pesticides.

The idea is that chemically similar agents can be well weighted together, as they might have a similar toxicological profile. And I think we can develop a

similar approach for chemicals in the California
Biomonitoring Program list. And we can overlay text
mining -- to text mining, if there are any available
literature data for different chemicals in the list and if
they can help in the priorities.

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DR. BARUPAL: So in conclusion, non-targeted analysis assays are a great potential for detecting high-priority chemicals exposome research and biospecimens. However, proper combination of analytical chemistry and data science need to be planned ahead. And indexing raw data into enterprise databases and avoiding a signal threshold prevalence -- threshold are needed for exposomic assays. Computational text mining can improve the prioritization process by linking chemicals to publications.

And hopefully, in the future, we can mine full text data for expending the chemical to publication linking. An interpretation bias remains a major challenge in mining NTA data, that utilizing existing biochemical databases, which are proven and complete for NTA.

With that, I want to thank my current and former collaborator at IARC, UC Davis, and Icahn School of Medicine at Mount Sinai. A special thank you to NIH for funding these two initiatives.

And thank you. Happy to take any questions

CHAIRPERSON SCHWARZMAN: Great. Thank you so

much. We have a little bit of time for Panel questions

for Dinesh and we'll check for audience questions as well.

DR. BARUPAL: Hi, Oliver.

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PANEL MEMBER FIEHN: Thank you. That's, of course, very interesting. What do you think about using large repositories, like the GNPS or massive resources for mapping non-targeted spectra to compounds, to similar compounds, to exposome compounds, do you know anyone who's tried that?

DR. BARUPAL: Let's do it.

PANEL MEMBER FIEHN: Can you explain?

DR. BARUPAL: So it need to be done. It's a great idea. And we know the GNPS people are actually importing data from other repositories and they're indexing those data. And they are mining -- reanalyzing the data in their platform. But it is their platform and it's their computational approaches. And there are other computational approaches in other groups. Like you have your own ideas to mine and annotate. I have my own idea to mine and annotate.

And I think it will be complimentary, if those data sets are mined with multiple annotation schemes and annotation of algorithms. But the -- but I don't know

anyone who's actually pooling those data, except GNPS and mining it.

CHAIRPERSON SCHWARZMAN: Other questions from the Panel.

Eunha.

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PANEL MEMBER HOH: Eunha Hoh from San Diego State University. It's a great presentation. Lots of information. A lot of work. One question I always have that, you know, the exposome study that measuring metabolites untargeted. But in the non-targeted analysis of environmental chemicals in the same biospecimen, there is always a challenge because of the concentration, it's very -- like environmental chemicals are at concentration much lower than the metabolites, or nutrients, or, you know, other chemicals that humans are exposed to. Is -- I kind of want to hear your thoughts about that. You know, I mean I know that you're doing a lot of data mining and, you know, informatics, but if you have any thoughts on that.

DR. BARUPAL: It's -- I think it's a well -- it's a known issue of analytical chemistry, the sensitivity of an instrument. And so if you start with a hundred microliter plasma sample and so -- and you have your sensitivity by LC-MS, of course, there are peaks that will be below the detectional limits.

And so if you give me 500 microliter, we gave five times. And so it's a combination that starting material the sample preparation, and the sensitivity of the instrument, that what compounds will be above the LOD and what will not be. But then it's the same time exposure itself, because you can have some population in a cohort that exposed to five times more to a chemical than other, then we will be able to see it.

But the way I think is that it should be by a database approach, that we index everything. And even if a single subject is exposed to it five times more, I think it's important that we highlight that.

PANEL MEMBER HOH: Um-hmm.

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CHAIRPERSON SCHWARZMAN: Other questions?

And, Marley or Sara, are there any questions from the audience for Dinesh?

MS. HOOVER: This is Sara and there are no questions in the Biomonitoring California email and there's no questions via the GoToWebinar platform either.

CHAIRPERSON SCHWARZMAN: Okay. In that case, I suspect we'll be returning to this information in our afternoon discussion as we continue to pull all of this together. And thank you so much, Dinesh for that presentation and I will introduce our next speaker.

DR. BARUPAL: Thank you.

CHAIRPERSON SCHWARZMAN: Doug Walker is an assistant professor in the Department of Environmental Medicine also at the Icahn School of Medicine at Mount Sinai. His research focuses on advanced analytical strategies for measuring the occurrence, distribution, and magnitude of previously unidentified environmental exposures, and assisting in delineating the mechanisms underlying environment-related diseases in humans.

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He also leads the high-resolution exposomics facility at Mount Sinai, which was established to provide high-quality non-targeted screening of biological samples for nutrition, precision medicine, and environmental health research. Doug will present on, "A Multi-Platform Non-Targeted Framework for Measuring the Human Exposome".

(Thereupon an overhead presentation was presented as follows.)

DR. WALKER: Great. Thank you. Can you see my slides okay?

CHAIRPERSON SCHWARZMAN: Yes, we can.

DR. WALKER: Okay. Great. Excellent.

So today, I'm really delighted to talk about our work using multiple high-resolution mass spectrometry platforms to provide better measurement of the human exposome. I've been really impressed by everything that I've heard so far, you know, but describing both

environmental sample analysis and human sample analysis.

I'm going to do kind of a deeper dive into how we're using non-targeted analytical strategies to provide better measurement of human exposures and link those to disease.

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DR. WALKER: So just briefly today in my presentation, I'm first going to describe a critical role for untargeted or non-targeted assays in characterizing the human exposome. I'll next describe how we're using untargeted high-resolution mass spectrometry to generate new insight into understanding more classical exposures, such as persistent organic pollutants and volatile organic compounds.

And then next, I'll describe how we're incorporating these untargeted assays to study exposures of emerging concern, including things like microplastics, and also develop different strategies for assessing exposures in population studies.

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DR. WALKER: So I'd just like to briefly describe why we're really interested in using these untargeted methods and why we have a focus on the exposome. We know from a number of different studies, and, you know, of course, by speaking to environmental health scientists we're all aware of this, but we know that genes, or your

genetics, alone do not describe your disease risk. And this was shown -- you know, this is another example of how we can demonstrate this, and this is from a paper by Steve Rappaport that was published in 2016.

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But what this graph is showing is the population attributable fraction for a number of common disease phenotypes, if we account for gene or gene-by-environment risk factors. Now, the population attributable fraction is the percent reduction in a disease that would occur, if we could eliminate some risk factor.

So what this is showing us is that for most of the common disease phenotypes, if we remove gene or gene-by-environment risk factors, the prevalence of a disease is only being reduced by, you know, at most 30 to 40 percent. And so what we're really interested in is identifying what is driving the disease risk, what is that other 65 to 70 percent, and how can we develop new analytical methods to better characterize that?

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DR. WALKER: Our current estimates suggest that only about five to 15 percent of diseases can be attributed to heritability alone. The other 85 to 95 percent of diseases are derived from environment, lifestyle, or their interaction with the genome. So therefore, if we're considering heritability alone, we're

taking an unbalanced view of human health and disease.

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DR. WALKER: However, there's a discrepancy between our ability to characterize environment and our ability to characterize biology. We have incredible technologies that allow us to very thoroughly characterize different aspects of biology from whole genome sequencing through transcriptomics, or epigenetics, or proteomics. They allow us to provide deep characterization or phenotyping of different levels of biology.

However, even some of our best analytical methods that we currently have only allow us to characterize about ten to a hundred exposures in an assay or in a series of assays. So therefore, in order to make the measurement of environment or exposures more consistent with its importance to human health, there's a critical need to develop analytical frameworks that allow us to provide universal screening for measures of the exposome.

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DR. WALKER: We've approached this through measurement of the human metabolome, which can be defined as the complete collection of small molecule metabolites found in the human body. As was mentioned previously and has been discussed in the other presentations, this includes a very diverse series of different types of

chemicals. These range from things like the core biological metabolites that are needed for proper biochemical functioning and for life, microbiome related chemicals, nutritive chemicals in diet, as well as exogenous compounds like supplements and pharmaceuticals, commercial products, and environmental chemicals.

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DR. WALKER: One of the advantages of measuring the human metabolome is that it allows a -- the ability to characterize both that endogenous contribution, which is a measure of biological effect, as well as the exogenous contribution, which is a measure of exposure or internal dose to these different compounds.

Our current estimate suggests that if we wanted to have the ability to characterize a reference human metabolome, we would have to have the ability to detect roughly one million chemicals. I'm not trying to imply that if I were to take my blood sample and run it, you know, using multitudes of different platforms, we would be able to detect a million chemicals.

But if we want to assemble a reference metabolome that includes metabolomic measures from very diverse populations, collectively, we would have to have some system to categorize, catalog, and detect a very large number of compounds. Our estimates suggest that exposure

is likely to contribute to 400,000, or approximately half, of these compounds or more.

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DR. WALKER: So therefore, to really embrace and understand the complexity of the human exposome, there is a critical need to adopt analytical strategies and study designs that incorporate untargeted measures of exposure.

There is no way we'll ever be able to develop panels that allow us to characterize or biomonitor for compounds on that scale. So we have to really start thinking about how we can incorporate these data-driven methods for detecting exogenous molecules in human populations.

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DR. WALKER: The measure of the human metabolome provides us a comprehensive measure of the continuum from exposure to disease. And we can use this information to really understand the biological effects of exposure, and start developing new hypotheses to study about the length between exposure to disease. So we know that by measuring exogenous compounds, we have a measure of exposure or biomarkers of exposure in a biological sample.

Because we're characterizing the metabolome in our assays, we also able to characterize endogenous metabolites and how those are associated with those

exposure markers. So we have intermediate biomarkers of effect. And if we're looking at certain disease populations, we can also identify physiological manifestations of disease within the metabolic phenotype.

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When we think of ways of combining those, we can essentially develop a framework for what we like to think of as systems toxicology.

So as I mentioned just previously, we can use measures of exposure and to identify exposure to phenotyping and biological effects. If we work with disease populations, specifically nested case control studies, we can look at how metabolic or exposomic alterations are associated with the disease phenotype. And we can link these exposure and biological effects to start identifying hypotheses that we can study and model systems, and use more informed approaches for describing mechanisms linking exposure to disease.

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DR. WALKER: The way we've operationalized this concept of measuring the exposome is through a multi-platform approach that relies on both gas chromatography and liquid chromatography high-resolution mass spectrometry.

By combining these, we have a framework for doing a comprehensive exposome by metabolome-wide association

study of disease. Using the gas chromatography or gas phase analysis, we've really focused our methods towards detection of volatile and semi-volatile organic compounds. Many of these are the environmental exposures that were interested in studying.

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Our extraction method is really optimized to, you know, isolate these compounds from either tissue or blood samples. And we're primarily characterizing these on the gas chromatography system.

At the same time, every sample that we're analyzing on the gas chromatography mass spectrometer, we're also analyzing using liquid chromatography with high resolution mass spectrometry. This allows us to characterize biological response markers, as well as more polar metabolites, and environmental chemicals, and also just more polar exogenous compounds such as drugs and their metabolites.

By combining these two platforms, it gives us the ability to detect more components of chemical space and improve what we're measuring in our exposomic assays.

DR. WALKER: The gas phase or gas chromatography analysis is based upon the thermo GC orbitrap instrument. We've established a, you know, very robust and relatively high throughput workflow that allows us to do simplified sample extraction and then analysis in a fully untargeted

manner.

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As I mentioned previously, our sample extraction has been more focused towards non-polar, volatile and semi-volatile compounds. And so this allows us to target environmental compounds. We found that the sensitivity of the instrument is excellent, so even operating in full scan mode, we have similar sensitivity to what we would see with a GC triple quad. And we're able to routinely detect very low abundance environmental chemicals at the subnanomolar or subnanogram per ml range.

These include things kind of like the polychlorinated biphenyls, brominated flame retardants, as well as some of the new phosphoester flame retardants and other compounds that are relatively volatile.

Our workflow is completely untargeted, in that, we don't have any specific targets or MS/MS that we're trying to collect during the initial analysis. We've optimized the method to detect as many signals as possible. Once we've run samples from a study, we then go back and do our peak picking and data extraction. We run our bioinformatics and then do our metabolite annotation followed by identification.

Identification, of course, is based upon confirmation by comparison to standards or high confidence annotation by matching to databases or in silico data.

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DR. WALKER: The liquid chromatography high resolution mass spectrometry platform, which again we use for exposure of biomarkers and biological response measures is, you know, relatively traditional untargeted metabolomics platform. We use a Q Exactive HFX orbitrap mass spectrometer that's interfaced to a liquid chromatography system that has dual chromatography capabilities.

So every sample we're analyzing, we're actually analyzing two different modes. One is with positive electric spray ionization. The second is with negative.

Similar to the GC, we're operating in a completely untargeted framework, where we're collecting our data in full scan mode. And then after we perform our biostatistics and bioinformatics, we go back and do our annotation and identification using standards.

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DR. WALKER: I'm just going to go through a few of the studies that we're applying these methods to and how we're using this untargeted data to better characterize exposures that are being experienced by human populations.

One of the demonstrations of the usefulness in this technology was an initial study that we did to

characterize metabolome of Chinese workers occupationally exposed to trichloroethylene, which they were using in degreasing operations in a manufacturing plant.

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So within this study, we had collaborators who went to different regions of China and identified factories that were using -- only using trichloroethylene in the degreasing process as well as control factors that weren't using any volatile organic compounds in any of the processes that are ongoing in that factory.

After identification of these workers who were exposed and unexposed, they did an extensive shift monitoring of personal exposure levels, and collected blood and urine samples for characterization using, you know, a number of different assays.

We received blood samples from 95 unexposed workers ad 80 exposed workers and analyze them using the high-resolution mass spectrometry platform -- or framework described in the last slide.

We performed our metabolome-wide association study. And as a last step, we also tried to compare our metabolomic data to independently measured bioeffect markers that have previously been characterized in this population, and include things like immune, kidney damage, and exposure biomarkers.

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DR. WALKER: So the initial metabolome-wide association study identified 188 features at this -- they were features. These were not identified compounds that were associated with this exposure. We characterized these used in a number of approaches. But one thing that became very evident is we saw a high degree of features or high number of features that were very highly correlated with the exposure, but they didn't match any known TCE metabolites.

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When we characterized them further, we found that all of them had very consistent chlorine isotope patterns, suggesting that they could be arising from the trichloroethylene. But again, they weren't matching any of the known metabolites.

To characterize this further, we just looked at the biological response profiles associated with the exposure in these different metabolites. We found that a number of pathways were associated with the exposure that we would expect to see. For example, methionine and cysteine metabolism and bile acid biosynthesis were both associated with the exposure, which is what we would expect since these were -- these are the detoxification routes for this compound.

However, there is a number of other pathways that we weren't expecting, such as carnitine shuttle, which is

related to mitochondrial function, as well as purine metabolism and a few other immune-related pathways.

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Where the results really became interesting was when we started looking at the correlation with these other biological response measures, including immune responsive biomarkers, renal damage biomarkers, and TCE exposure biomarkers that were previously measured in urine.

So what we found first is that, you know, we were able to detect and identify a number of known TCE metabolites. This included trichloroacetic acid as well as a trichloroethanol. These were positively correlated with the urinary biomarkers that were again measured using a targeted lab. This is great, but it's exactly what we would expect. If we didn't see these correlated, I would be more concerned. So that in itself isn't exciting.

But what did become interesting is when we started looking at these compounds that we knew were halogenated chemicals, but they did not match any of the known TCE metabolites. And what we were able to see from this correlation network is that these identifiable metabolites were linked to the biological response markers that have previously been associated with trichloroethylene exposure. However, we didn't see any of these relationships with the known TCE metabolites.

So it suggests that there could be alternate pathways or relationships between these unidentifiable metabolites in these outcomes. And we're trying to explore this further and identify what some of these metabolites were using both MS/MS and cell culture studies.

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DR. WALKER: We've also been applying these methods in case control studies of different diseases. So this is an example of a study where we were examining how the exposome and metabolome was associated with primary sclerosing cholangitis and primary biliary cholangitis, which is a liver disease that really is un -- of unknown etiology. There is a few studies showing that there is some genetic risk factors for this disease, but the majority of the disease variance is really unknown.

So in this study, we wanted to apply the exposome assay, which is based upon gas chromatography and the liquid chromatography platform to characterize biological response and see how those were linked.

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DR. WALKER: The initial analysis using the gas chromatography platform allowed us to identify a number of different exposures that were correlated with the disease status, both for PSC and PBC. And for most of these IBD

status, which is a comorbid condition with primary sclerosing cholangitis really didn't impact the exposure levels.

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In addition to the identifiable metabolites that we were able to detect, and these were identified by comparison to standards using retention times and fragmentation patterns, there was also a number of features, which had spectra, and they were consistent with chlorinated or brominated compounds that did not match anything that we could find in our database.

So again, these are very interesting molecules, but we're trying to follow up with those and study them further to see what they are.

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DR. WALKER: The metabolomics analysis was able to identify a number of biological alterations or changes in biological pathways that were associated with the disease. It's very important to mention that these individuals were very, very early in the disease process. And so they had very minimal, you know, measurable changes in their phenotype, but we were still able to see very large differences in metabolic pathways for these individuals.

One of those was bile acid metabolism. And we were actually able to take our untargeted results and see

which metabolites were associated with the disease and with severity, and then we developed a targeted assay or used the targeted assay and were able to replicate many of the findings in both this population and a much larger independent PSC population as well.

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So that was a good example of how we can use the untargeted methods to, you know, guide our biomarker selection and use that for clinical characterization.

And finally, when we looked at linking these pathways that were associated with the disease to the exposure, we were able to find a number of different relationships. And what was most interesting about this is we kind of grouped our pathways based upon overall disease process. So we had bile acid pathways, oxidative stress pathways, and immune-related pathways.

And when we looked at the correlation between exposure and biological response measures, we were able to group those into different clusters. And they were largely consistent with the process itself. So we saw two clusters that were really enriched in bile acid metabolism and then we had a third cluster that was more enriched in oxidative stress and immune response.

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DR. WALKER: And so really what these untargeted assays allow us to do is provide a functional measure and

sensitivity needed to perform exposome by metabolome-wide association studies of human health and disease. There's always been this question about whether the metabolomic approaches provide the sensitivity to measure very low level environmental chemicals. But by using the gas chromatography platform we've really been able to demonstrate that we do have that sensitivity and it really expands what we're able to detect in some of these untargeted assays.

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DR. WALKER: And very briefly, I'll just spend a few minutes talking about what are some of the new exposures and new sample types that we're characterizing using these methods.

So one of the things that we want to be able to do better is measure exposures that have a relatively rapid half-life. And we can't necessarily characterize these in biological samples. So one of the things that we're working with are the silicone wristbands and badges that were initially developed by Kim Anderson's labs. We've shown that you an apply the untargeted methods to characterize these, and with a high degree of sensitivity actually isolate different microenvironment exposures even in different rooms in the same house. So they provide a high degree of sensitivity and they're relatively cheap to

mail out and distribute.

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We're also working on micro-needle patches that can be worn for up to 24 hours. These patches were vaccine patches that were essentially re-engineered to withdraw interstitial fluid rather than deliver a vaccine. And we can use these for biological response monitoring or internal dose monitoring.

The hope is to get these developed to the point where they can be delivered to participants in a cohort. They can apply them, where them for 24 hours, and ship them back.

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DR. WALKER: And finally, one of the things we're really starting to focus on now is developing these untargeted analytical strategies to measure microplastics in biological samples. There's been a lot of interest in characterizing exposures to these micro and nanoplastics. However, most of the work to date has been in environmental or food samples. And there's really no good methods for biomonitoring of these microplastics in human populations.

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DR. WALKER: So we're really trying to leverage all of our high resolution mass spectrometry capabilities and strengths into measuring these. We're using pyrolysis

high resolution mass spectrometry for quantifying known particles, as well as screening for a very wide range of different polymers and particulates that could be contributing to this burden.

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We're using depolymerization methods with liquid chromatography that allow us to characterize bound additives as well as screen for known particle monomers. So we're essentially destructing the monomers and trying to look for the -- or the polymers and looking for the building blocks of those particles. And we're also doing small molecular profiling to look for free plastic additives or things that could be absorbed and co-transported on these particles.

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DR. WALKER: And a lot of the projects we're focusing on now relate to early life exposure including studies where we're trying to measure these in both placenta and amniotic fluid from pregnant women. And this is in a collaboration with researchers in Europe at Utrecht University and Deltares.

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DR. WALKER: So just in conclusion, untargeted assays that allow us to profile the human metabolome have the potential to provide key insight into exposure and biological response and how that's associated with disease

outcomes by combining a gas chromatography and liquid chromatography high resolution mass spectrometry platform. We really tried to establish a unified framework for expose via metabolome by an association study of disease. And these technologies can be leveraged for developing new methods to better characterize emerging exposures, such as microplastics. And as we saw in the previous presentation, some of the PFOS.

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DR. WALKER: I'd just like to acknowledge my team at Mount Sinai and all the excellent collaborators I've had over the years, specifically Roel Vermeluen at Utrecht University and Dean Jones, Gary Miller, and Karan Uppal at Emory University.

Thank you very much.

CHAIRPERSON SCHWARZMAN: Thanks so much, Doug, for that. And we have a few minutes for questions. And we are going to break right at 2:30 for 15 minutes. So anything that we don't get to now, we can -- let's bookmark for the later discussion today.

Questions from panelists to begin with and then we'll turn to the audience.

Jenny.

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PANEL MEMBER QUINTANA: Hi. Thank you for that excellent presentation. And you showed the power of this

approach when you had applied it to very carefully characterized subjects. So one case exposure was extremely well characterized, TCE, you know, with good selection of subjects, and excellent exposure measurements, air exposures. Another case, you had a carefully characterized group of diseases -- people affected by a disease.

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And I'm just wondering if you could comment or give your thoughts about whether it's better to -- directly -- to use your resources on these kind of very targeted groups or to look like California Biomonitoring is doing a larger population that's not carefully selected, if you have thoughts about that.

DR. WALKER: Yeah. So we've always -- and, you know, both the mentors -- my mentors who I worked with at Emory and, you know, something I have still envisioned these platforms as is basically a universal chemical surveillance and biomonitoring framework.

So we've initially applied these to very well characterized populations with the hope of demonstrating that they have the sensitivity and it's feasible to measure many of these exposures. I really see the ultimate application of them is kind of to -- for population screening or even applying to, you know, very diverse disease groups to look at, you know, groups of

exposures that could contribute to either specific or non-specific disease processes.

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So I absolutely do believe that there is a lot of application more in population screening. I think these initial studies have demonstrated what's feasible using this technology for exposome analysis. And I think there's a lot of potential to apply them in larger populations.

Our goal has been to make the assays as high throughput as cheap and use as little sample as possible, so that there is the potential to apply these more broadly to, you know, cohort studies and other populations.

PANEL MEMBER QUINTANA: Thank you.

CHAIRPERSON SCHWARZMAN: Other questions from the Panel?

Let me check in with Sara and Marley to see if there's any questions from the audience?

MS. HOOVER: Hi, Meg. This is Sara, and I can tell you there's no question from the email, but there is one question from the audience and I believe Marley is going to share that verbally right now.

DR. ATTFIELD: Hello. Can you hear me?
CHAIRPERSON SCHWARZMAN: Yes.

DR. ATTFIELD: Okay. Hi. This is Kathleen Attfield. I'm in the CDPH part of the Biomonitoring

California program. I was wondering -- I just don't know the background of this. Is it possible to pair dose-to-animal studies with the human studies, such as the TCE experiment you talked about?

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DR. WALKER: So it's not -- that's pretty challenging. And that's one of the issues that we're really facing with the TCE studies. So if we go to a cell model, you know, we're essentially putting the TCE in solution. And that exposure isn't representative to how these people are being exposed. Their exposure was respiratory.

So, you know, there is ways to kind of correlate animal dose to what we're seeing in the human populations. However, you know, one thing that we're -- the reason we're using these occupational cohorts is because, you know, they are very high exposures. These are higher levels than you would ever see in -- you know, in just a regular population, but they allow us to kind of cleanly define the human response to these exposures in a way that, you know, couldn't characterize in an animal model, and that you could never test, you know, using a controlled exposure.

So, you know, these are very unique populations.

And I think applying these methods first to these high exposure populations provides, you know, important insight

into how the exposure affects the biology or how it affects the pathways.

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I'm a -- you know, not a -- I have some toxicology training, but not a toxicologist by -- you know, through and through. So I would have to speak with some of my colleagues about what the best way to extrapolate that human exposure to an animal model would be.

CHAIRPERSON SCHWARZMAN: Ulrike.

PANEL MEMBER LUDERER: Yeah. Thank you for that very interesting presentation. The question I have for you is you -- you know, the platforms that you -- that you are using that you've talked about obviously were designed to maximize the types of, you know, both exogenous -- metabolites of exogenous chemicals, as well as endogenous, you know, metabolites that you can measure. And I was wondering if you could say anything about what of the universe of, you know, the metabolome in humans do you think you may be missing with these platforms that you have?

DR. WALKER: I mean, I would still say we're missing the majority. You know, I think we're moving forward with baby steps. You know, we're going from being able to detect, you know -- I don't want to -- you know, we can detect a lot of features, but, you know, when we

deconvolute the features and convert them into spectra and do all the filtering, you know, we went through a measuring about, you know, 15 to 2,000 compounds on the LC to now being able to detect an additional, you know, 10,000 or so on the GC. And again these are not features I'm talking about. These are actually, you know, de-convoluted spectra that we believe are signals that could be arising from the sample itself.

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But, you know, we're still not even close to measuring the totality of chemical space. I mean, it's an improvement, but relative to what's out there, it's an incremental improvement. But, you know, as long as we're moving forward, I feel good about that.

I think we have pretty good detection of a lot of the things that we're interested in, you know, again for the GC, the semi-volatile and volatile compounds. From the LC, you know, we capture really good -- most of the endogenous pathways some representative -- some representation from those, as well as lot of drug and environmental chemical metabolites.

But, you know, it's challenging to really put a number on that, especially when we don't know what the number is in terms of how far along we are, but I think we still have a ways to go.

CHAIRPERSON SCHWARZMAN: Great. Thank you so

much, Doug, for that really interesting presentation. And we're going to have a 15-minute break. We'll restart right at 2:45 for our final presentation of the day followed by an afternoon discussion session.

So see you then.

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(Of record: 2:31 p.m.)

(Thereupon a recess was taken.)

(On record: 2:45 p.m.)

CHAIRPERSON SCHWARZMAN: Okay. We are going to restart. And just to give you a sense of where we're going. We have one final presentation now and then the afternoon discussion session. So I will introduce our speaker -- our final speaker of the afternoon is Rachel Morello-Frosch who's a professor in the Department of Environmental Science Policy and Management and the School of Public Health at UC Berkeley.

Her research examines how social determinants of environmental health and environmental chemical exposures interact to produce health inequalities in diverse communities.

OEHHA and CDPH collaborated with Rachel to develop and test Biomonitoring California's results return template. And so she'll be presenting on that today on, "Chemical Suspect Screening as a New Approach to Biomonitoring: An Application in Firefighters and Office

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             presented as follows.)
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             DR. MORELLO-FROSCH: Thanks. Can everybody hear
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   me?
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the next slide showing.

If you're working with an external monitor, also sometimes you just have to swap them.

DR. MORELLO-FROSCH: Yeah.

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PANEL MEMBER SINGLA: Or you might -- you might try turning off presentation mode.

DR. ATTFIELD: Or choose duplicate.

DR. MORELLO-FROSCH: What do you see now?

CHAIRPERSON SCHWARZMAN: We see the same thing as before, the presentation mode, but from the presenter's view.

DR. MORELLO-FROSCH: Sorry about this. I tested this and it worked before. And now it's not working.

CHAIRPERSON SCHWARZMAN: That's okay. And don't worry about your time. We'll give you the full time for your presentation and just work it out.

DR. MORELLO-FROSCH: No, I'm more worried about your time.

MS. HOOVER: Hi, Rachel. This is Sara. We could -- we have your slides, we could set them up and advance them, if you want to say next slide, if you're not able to get this to work.

DR. MORELLO-FROSCH: What are you seeing now?

MS. HOOVER: The same thing, presenter's mode.

DR. MORELLO-FROSCH: Okay. I don't know how to fix this.

MS. HOOVER: Okay. So why don't you close yours.

CHAIRPERSON SCHWARZMAN: There you go.

MS. HOOVER: There you go.

(Multiple voices at once.)

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DR. MORELLO-FROSCH: How is that?

MS. HOOVER: There you go. All right.

DR. MORELLO-FROSCH: Okay. I really apologize for that.

CHAIRPERSON SCHWARZMAN: The only thing Rachel, there's -- the control panel for GoToWebinar, it appears to us to be in the middle of your slide. Can you move it to your other monitor?

There you go. Perfect. Thank you.

DR. MORELLO-FROSCH: Okay. Thanks for your patience while I work through this. So today I want to talk to you about a study that I've been working on with a bunch of collaborators applying non-targeted methods to examine environmental chemical exposures among women firefighters in San Francisco. This study originated among firefighters themselves in the City of San Francisco.

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DR. MORELLO-FROSCH: Our fire department there is unique in that it has the largest number of women firefighters out of all urban fire departments in part

because of the consent decree awhile ago to desegregate the fire department. And so women have been with the Department for a while moved up its ranks. The last two chiefs have been women. And this person on the upper left Jeanine Nicholson, who is the current chief, before she became Chief, actually was very public about her experience going through breast cancer.

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And so increasingly women were concerned about their exposures through environmental chemicals on the job and the -- trying to understand the extent to which it might be linked to what's going on in firefighting.

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DR. MORELLO-FROSCH: Most studies that have looked at exposures to environmental chemicals among firefighters have been conducted almost exclusively in men, in part because it is predominantly still very much a male dominated profession. Although, it's diversifying in many areas. And a lot of those studies have shown that firefighters have higher exposures to a variety of compounds associated with firefighting and other aspects of their jobs, including PFASs, PAHs, flame -- different kinds of flame retardants, dioxins and furans as combustion byproducts.

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DR. MORELLO-FROSCH: And many of these chemicals

have been shown in animal toxicology studies, and this review was done by my collaborators at Silent Spring Institute, many of these chemicals that firefighters are exposed to have been shown in toxicology studies to be mammary carcinogens. And so they definitely warrant further study and it presented us an opportunity to characterize the extent to which women firefighters might be exposed to some of these compounds that have applications for Mammary development in breast cancer.

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DR. MORELLO-FROSCH: So in collaboration with the firefighters who approached our science team with Silent Spring Institute, as well as UCSF, colleague Roy Gerona, who was doing the initial non-targeted work, we created the Women Firefighter Biomonitoring Collaborative. And we also had environmental health advocates working with us on this to do some of the research translation piece, which I will not be talking about today.

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DR. MORELLO-FROSCH: And together we created this study. And our primary aims were to characterize chemical exposures among women firefighters and compare them to a control group, in this case women office workers who also work for the City and County of San Francisco, but who are not first responders. We are also assessing potential

impacts of the chemical exposures that we have characterized on upstream biomarkers of effect. And I will not be talking about that today.

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DR. MORELLO-FROSCH: So our inclusion criteria were we wanted women over 18 who were non-smokers. For firefighters, we wanted women in the fire service who had had at least five years of active duty. And we ended up with 83 women firefighters and 79 women office workers in our study population.

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DR. MORELLO-FROSCH: And we collected biospecimens, serum, urine, and whole blood. And we did targeted and non-targeted analysis in parallel in our studies. So there was definitely a focused interest on PFASs, but also on legacy and current use flame retardant OPFRs. Again, I won't talk about that today. We have some papers in the works and the PFAS papers published. And then we did in parallel non-targeted chemical analysis. And the biomarkers are thyroid hormones and telomere length.

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DR. MORELLO-FROSCH: We did an exposure assessment interview looking at occupational and work activities, diet, personal care product use, and consumer

product use in the home and in other places.

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DR. MORELLO-FROSCH: And just as a little plug for potential collaborators, we have building -- in addition to the samples that we've collected in the study, we sought to build a bioarchive that would facilitate future analyses as analytical chemistry methods improved. So we have a very nice repository. And we are definitely eager to collaborate with others who might want to take advantage of the samples that we currently have banked. And we also had added nurses which I'll talk about in a second.

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DR. MORELLO-FROSCH: So as I said, we've had two papers out. And this one on the upper left, which is in the list of references in your packet and on -- available on the SGP website. You can read both this -- this paper that I'm going to talk about today.

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DR. MORELLO-FROSCH: So our study demographics for the office workers, which is this column on the left and the firefighters on the right. From a racial and ethnic point of view, they were very similar. We had differences in educational attainment where the office workers had higher levels of educational attainment and

the firefighters had higher levels of income.

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DR. MORELLO-FROSCH: And so we set about developing an in-house chemical mass spectra database for our project. And in this case, we were limited to chemicals that would ionize in the negative mode. And we -- so we started with a baseline, a chemical library, of what we call environmental organic acids, which we had used for another study, characterizing exposures among pregnant women. And then we added to that library of chemicals. Had a lot of conversation with the firefighting community, but also based on that review that Silent Spring conducted adding chemicals that were shown in toxicology studies to affect mammary gland development. And the endocrine disruptors that were an initial promoters of mammary tumors. And then we added additional chemicals of specific interest for firefighting. again, we focused on compounds that would ionize in the negative mode.

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DR. MORELLO-FROSCH: And so similar to the other workflow diagrams that you saw in prior presentations today, we ran our samples through LC-QTOF/MS electrospray negative mode. And you get this -- these chromatograms, which then we processed to narrow down the kinds of look

at -- we look at the features and then try and narrow it down to chemicals, chemical compounds, and -- that match formulas and then we sort through the different isomers by comparing retention times.

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And then we go through a process of prioritizing which ones we want to confirm and measure using targeted methods. And because of resources, we have a priori criteria for deciding which ones we're going to actually quantify.

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DR. MORELLO-FROSCH: So this is an example of the cumulative number of environmental chemicals that we detected using LC-QTOF. Each one of these lines represents an individual participant in the study. And the X axis is the number of chemical hits or candidate compounds. And the color coding is the different chemical classes that these chemical candidates fell into.

And so you can see there were quite a few phenols, phthalate metabolites. And a lot of this is also driven by the library itself. So I think that's important to point out. And we could see that the hits were fairly similar although slightly higher in firefighters.

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DR. MORELLO-FROSCH: And then we embarked upon a scoring and ranking of these chemicals based on the

suspect screen. And as I said, we had an in-house library of about 722 chemical formulas. And we were able to narrow it down to isomers that were matched to 300 chemical formulas or -- and then with retention time correction, we had about 622 putative compounds, many of which were isomers.

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Here, we applied different kinds of inclusion criteria based on whether or not, for example, peak areas were higher in firefighters, differences in detection frequencies, between the two groups. Maybe a novel compound was ubiquitously detected in the entire study population. And then, of course, chemicals that were of interest because of their implications for mammary gland development in toxicology studies, their estrogenic potential, and also chemicals that had not been previously biomonitored in NHANES or Biomonitoring California.

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DR. MORELLO-FROSCH: So this is a partial list of the candidate chemicals that we sought to validate, just so you can kind of get a sense of how we did this. And we -- this is looking at the differences in the detection frequencies of these chemicals, in terms of between firefighters and office workers. And then we compared the mean peak areas between the two groups, the average mean peak areas. And so where you see stars and symbols, this

indicated a significant difference, particularly in terms of peak areas.

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DR. MORELLO-FROSCH: We then applied a scoring approach, based on those criteria that I just talked to you about in that workflow, so whether or not they were flame retardants, differences in detection frequencies, whether or not they were statistically significant, whether this was a novel compound that hadn't been biomonitored before, genotoxic potential, estrogenic potential, whether or not that something was not found at all in office workers, but found in firefighters only, and then whether or not it was a mammary carcinogen. Again, it's another attempt to prioritize our chemicals.

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DR. MORELLO-FROSCH: And then we used, where there were commercially available standards, we sought to validate the presence of these chemicals. And so you can see, you know, we had different kinds of chemicals, some of which had several different types of isomers with different mean retention times in our serum samples. And we compared that to the lab standards. And so the ones that were -- that have the check were the ones that we were able to validate in the QTOF. And these ones below we were not able to validate because of a retention time

mismatch.

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validation process, then we were able to prioritize which

chemicals we actually wanted to go in and measure and

quantify, based on targeted methods. And so this is

DR. MORELLO-FROSCH: And then based on that

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descriptive statistics of the select chemicals that we measured from the NTA, and then we actually quantified them using targeted methods. And as you can see here, we didn't see huge -- huge differences, but we did find some interesting compounds, including Dinoseb, which is a banned pesticide.

It's also, it turns out, present in some dietary

medications. So we're in the process of finding out from

some of our participants whether or not some of them take

some diet medication that maybe might explain the presence

of this compound in some of our participants.

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DR. MORELLO-FROSCH: So we are expanding our cohort and we have been adding nurses. So we have 60 nurses that we recruited, additional office workers, and firefighters, a subset of the firefighters in our study.

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DR. MORELLO-FROSCH: And we are looking at them, these firefighters, after a fire event. So they basically

contacted us 24 -- within 24 hours post-fire and were collecting blood and urine temporally and conducting interviews. And then for the nurses versus office workers, we are collecting blood and urine from them and doing a comparison between these two groups. And in this case, instead of doing targeted, and non-targeted, in parallel, the non-targeted analysis for both of these studies is going to drive what we're going to actually look at in the targeted analysis.

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So it's providing us a method for prioritizing those compounds that we want to confirm, and quantify, and see if we can detect novel compounds that haven't been measured before in these study populations.

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DR. MORELLO-FROSCH: Finally, I want to get back to this issue of reporting back. As I think this Panel has already heard about the many ways in which we've done digital exposure report back that interface these tools to return study results to study participants. And we collaborated a lot with the Biomonitoring Program to do this kind of thing in our study populations and in particular with firefighters.

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DR. MORELLO-FROSCH: And so this is kind of the screen shots of what these kinds of things look like for

targeted analysis, which we've already returned to our participants. So this gives you examples of like what's -- the information on the chemicals and then participants can actually see what their levels are compared to others in this study population.

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DR. MORELLO-FROSCH: But we are now in the process of doing some report back on what we found out from the non-targeted analysis. And this is kind of language which you can kind of study when you look at the slides more carefully. But it poses some interesting questions about how do we communicate how this technology works to study participants, because people are used to kind of traditional explanations that we think what we're going to measure first and then we go and measure it, and while this is a very different approach.

So -- and the decision amongst the group, because this is a community-based participatory research study, was to report aggregate results rather than individual level of results for these preliminary non-targeted measures that we had in the study population.

So in terms of thinking about how we return results for non-targeted analysis to study participants, I think this is going to become an increasingly important thing to think through how we communicate to our study

participants. In other words, how do we describe this method in a meaningful way? How do we explain how non-targeted differs from targeted methods and what are some of the advantages and disadvantages of this approach? How do we explain and distinguish between chemical suspects versus confirmed and quantified compounds when we're trying to communicate the different steps that we undertake to elucidate novel chemicals that maybe haven't been studied before in these populations.

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And do we report things, for example, you know, the number of chemical suspects found in each participant, because, you know, like that graph that I showed you with the hits and the different chemical classes, do we provide it by chemical group?

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DR. MORELLO-FROSCH: So do we do kind of a suspect profile for each participant to kind of walk them through how this process works or is that information overload?

And then how do we convey our decisions about the criteria that we applied for choosing which chemicals we were ultimately going to validate and quantify in our study population?

Because I think one of the challenges that has been the subject of today's conversation is, you know, how

do you narrow it down, and what criteria do you apply, and how do you -- how are you transparent about those decisions, particularly when you're doing a collaborative project with your study participants or community-based participatory research? How do you come up with those criteria for choosing those subset of chemicals that you're actually going to measure using targeted approaches?

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DR. MORELLO-FROSCH: So I will leave you with that. I just want to thank our illustrious team. This is definitely a community-based participatory research project. It's data driven in terms of non-targeted analysis, but also community driven, in terms of how we've designed our study protocols and sought to analyze and disseminate our data.

Thank you.

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CHAIRPERSON SCHWARZMAN: Thank you so much,
Rachel. We really appreciate that. We have a few minutes
for questions for Rachel and we'll start with the Panel.
And just a reminder, you can just raise your hand and I
will call on you. And then we'll go to the afternoon
discussion. We can just sort of blend into that.

Okay. Questions specifically for Rachel? Oliver.

We see you Oliver, but we don't hear you.

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PANEL MEMBER FIEHN: I needed to find the panel. It's like where is the Panel?

Thank you for that presentation. So you and several other participants have stated something about in-house panels. And I really wonder about why is it in-house? Why -- can you make things public? Can you make spectral libraries public, retention times public? I mean, do we all have to buy the same compounds again, and again, and again? Somebody else said something about, you know, the reproducibility of spectra, you know, because it is important to share information. And also, not only that, but at some point also share raw data, if we can. I mean, Dinesh Barupal talked a little bit about the problems in sharing raw data. And I guess that's another discussion. But at least for spectra, we should be able to share them, shouldn't we?

DR. MORELLO-FROSCH: Yes, I totally agree with you. And as I mentioned really quickly in my conversation was we developed, you know, our own spectral database, not because we were trying to keep it proprietary, but we used as a baseline another spectral database at the time that we had used for something else and then built upon it.

But it's something definitely that we seek to share and expand. And it's definitely not something

that's proprietary. And now, for example, we -- now that we've added nurses, we are collaborating with the Department of Toxic Substances Control. So we're using even more -- you know, a much larger spectral database ionization in the positive and negative mode. So I agree, these are things -- this is information that you will want to share, so that other people can see, you know, how you built your database and how that impacted what you found, because it is limited by the database that you're comparing it to.

CHAIRPERSON SCHWARZMAN: Yeah. Oliver, do you have a follow-up?

PANEL MEMBER FIEHN: Follow-up question. So you also presented preliminary annotations that turned out not to be correct by retention time metric, right?

DR. MORELLO-FROSCH: Yeah.

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PANEL MEMBER FIEHN: But still, I mean, they are very likely very similar, right? I mean, they have the same -- a good spectrum and it matches nicely to your phenols, just not that exact isomer. How do you --

DR. MORELLO-FROSCH: Right.

PANEL MEMBER FIEHN: How do you suspect -- how do you suggest we should handle those, those isomers -- an unknown isomer of?

DR. MORELLO-FROSCH: Yeah. Well, again, this was

sort of a proof-of-concept paper. So just to kind of walk us -- walk ourselves through this process to figure out a prioritization scheme of how we might make these decisions. And I think there is now kind of more sophisticated statistical strategies that we could undertake to make some decisions and not sort of arbitrarily be eliminating isomers. I also think that again this data that you generate, the raw data, can be mined again and again. And so, you know, those -- you can revisit those isomers and other kinds of things to redo your analysis or to re-mine the data again.

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CHAIRPERSON SCHWARZMAN: Rachel, I'm just curious how -- how the study participants kind of related to the non-targeted aspect of this study? Did your -- sort of what did they want from it -- your expectations(inaudible) with NTA in its current form? What was that like including that portion of the analysis in the study, as opposed to the sort of more traditional and easily understood targeted analysis?

DR. MORELLO-FROSCH: Yeah, that's a great question. So when we -- when we co-designed the study, it was -- it was decided that we -- that they -- they were very interested in applying this -- what was then a more novel technology add now has proliferated a lot more since we started. But they also wanted to make sure that we did

targeted analysis for chemicals that they were very specifically interested in, in this case, current and legacy flame retardants, PFAS, and some of these other kind of usual suspects that have been studied in other populations in order to compare what has been found -- what would be found in women to other studies.

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And I -- but there was also a lot of excitement about being able to apply this technique to elucidate chemicals and to maybe discover something new. They also knew the risks that maybe, you know, this -- this strategy might not necessarily reveal, you know, a smoking gun, if you will, of some -- something novel that's very specifically related to firefighting.

So there's a lot of conversation, a lot of description of -- really visual descriptions of how time of flight technology works, how you -- how you scan the samples, the challenge of the isomers, how you narrow it down, all that kind of stuff. So that was a lot of conversations in sort of our weekly science meetings was explaining this technology and to apply it and then thinking together about the kind of criteria that we want -- we would want to apply to narrow down the chemicals that we would measure directly, because we also had -- they wanted us to measure some of these chemicals that we've discovered through non-targeted analysis

directly. That was an important part of it for them.

And, of course, you're limited by resources in terms of choosing which ones you're going to measure.

CHAIRPERSON SCHWARZMAN: Yeah. Great. Thank you.

Eunha, you had a question.

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PANEL MEMBER HOH: This is Eunha Hoh San Diego State University.

It's very interesting work and I'm glad to see the project and really appreciate that presentation, and also very interesting subjects. Non-targeted analysis, you show the results based on the LC/QTOF, and -- but I'm thinking about, you know, considering that the firefighters are much more exposed to combustion, you know, products, you know, like more chemicals associated in combustion. Do you plan to analyze the samples on more GC side of analysis -- non-targeted analysis?

DR. MORELLO-FROSCH: Yes. We -- we are -- we would definitely like to be table to do that. We are -- we need to get more resources in order to do that, but yes, absolutely.

CHAIRPERSON SCHWARZMAN: And, Veena, you had a question also and then we'll go into the discussion period.

PANEL MEMBER SINGLA: So hi, Rachel. Good to see

you.

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DR. MORELLO-FROSCH: Hi.

PANEL MEMBER SINGLA: Great presentation.

My question was about, you know, in terms of some of the primary health concerns the firefighters had, and especially breast cancer, how do participants feel about the total information that they got from the targeted analysis and non-targeted analysis? In relation to their health concerns, did they feel they got the information they wanted? And also, did they -- did the participants have any suggestions for what they might want to see done next or what should be done about the information?

DR. MORELLO-FROSCH: Yeah. So -- so one of the things when they -- when they came to us with wanting to do a study to try and understand the extent to which exposures might be associated with breast cancer, a lot of the conversation before we even sat down to write the grant for this was why an epidemiological study is not feasible in this population, issues around statistical power, all kind of things.

And so it became abundantly clear that, you know, an exposure study was going to be really trying and get at their question more indirectly and could also potentially be more efficient in terms of lifting up chemicals of exposure and their sources that they could act upon now.

So I think there was a lot of enthusiasm. But again, it took a -- it was a process to really have that conversation and decide what kind of study design was, A, feasible, and would also get at some of their concerns.

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And as a result of a lot of this work, there have been practices that have changed at the -- at the fire department to try -- try and reduce exposures and efforts to do not only education but also some standard operating procedures that have changed there as a result of some of the results that we've -- we've been seeing.

CHAIRPERSON SCHWARZMAN: Thank you so much for the presentation, Rachel. And I'm sure these issues will arise again as we continue the discussion. I hope you'll be able to be around for it.

And I want to transition into discussion section now. So a couple of reminders as we go through this just about how to participate. So as before, Panel Members, please raise your hands if you want to speak and I'll call on you. Guest speakers and Program staff go ahead and turn on webcams and raise your hands, if you want to speak also. For attendees, who wish to speak during the discussion session, please alert us by using the question feature or the raise hand feature, and go to webinar, and we'll call on you individually. And at that point, you can unmute yourself and ask your question or provide your

comment. And then once you finish, please, of course, mute yourself again.

I just saw -- oh, no, I see the view just changed. We're all good.

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Okay. And then during discussion -- so Panel members, just please keep your webcams showing unlike pausing them as -- as we have for the presentations, unless you need to step away. For everyone else, please keep the webcam off, and your -- your microphone muted, once you've finished speaking.

So webinar attendees can also submit written comments or questions via GoToWebinar as before or by emailing biomonitoring@oehha.ca.gov and we'll read them out loud an paraphrase them if need be, if we lack time.

So I want to start the discussion -- I have some discussion questions that I want to highlight, but also I want to make sure that we return to some questions that didn't get answered before we ran out of time. And on my -- please tell me if my list is correct, I have Veena first, Eunha, and Ulrike.

And do you still -- if those questions are still relevant, we can just start with Veena, and we'll go through that list, and just go ahead say if your question isn't relevant. It's okay. We can move on.

PANEL MEMBER SINGLA: Okay. Thank you, Meg. And

James, thanks so much for the excellent presentation earlier. I had -- I had two questions. One was in relation to the information we heard from John about the kind of wide range of coverage that he saw with, you know, different methods and different labs. And I just wonder if you had any sense of the coverage of the specific method you were using on -- with PFAS and if -- if you had any like ideas or estimates of what you might still be missing from the -- you know, the new compounds that have been identified?

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And my second question was about if we have any sense of if there might be wider exposure to these new chemicals you've identified beyond the specific sites, if we know anything about potential long-range transport of these chemicals, or if these chemicals are present as contaminants or otherwise in fluoropolymer -- polymers or the kind of end-use products of -- of those fluoropolymers?

DR. McCORD: Okay. So I'll answer the easier question first, which was the second one. So when we start to identify these new materials, we tend to both go back and look at places where we already have data that we've banked for things that we might have missed, where they were below a threshold that we would use for examining things. And in some of these cases, we do have

far-ranging samples that we can compare to. And then as we do more studies going forward, we always look for things that we previously discovered.

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So in the example of New Jersey, we have far field soil samples from as far out as New Hampshire, which is quite far north in the prevailing wind direction. And we were able to identify that material as far north as basically into another State and region. And it's not in samples that we've collected from the non-prevailing wind direction, where we no reason to believe that it's a thing.

For stuff like GenX, we've only ever really found them to an appreciable degree in industrial impacted locations. And it seems like because of this proliferation of new -- different types of materials, every different manufacturer seems to have switched to new things. So unlike the legacy PFAS compounds, like PFOA, which is ubiquitous, because everybody had -- had this convergent evolution onto a very narrow chemistry, because we've proliferated to all sorts of different new things, it appears like the problem is very site specific.

Everywhere that we go that we do an investigation like this, we find a different fingerprint, a different combination of chemicals. And depending on who the major producer or user is in that site, they're using whatever

chemical is best suited for their purposes. So like I said, we've got six or seven different states that we're working on.

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And in each site, there's a different major driver chemical. So, you know, Michigan is dealing with 6:2 FTS as a replacement for PFAS. New Jersey has these chlorinated compounds as a replacement for PFOA.

Fayetteville was -- has GenX and related chemicals as PFOA replacements. West Virginia is another Chemours site that's also dealing with both the GenX HFPO-DA compounds, but then other things. And it does seem to be very stratified in terms of what's a problem where. It's no longer a universal PFOS -- or a universal PFAS question.

And I've Completely forgotten what your first question was, if you could repeat it.

PANEL MEMBER SINGLA: Sure. No problem. If you have any sense of the coverage of the -- your method for PFAS?

DR. McCORD: Yeah. Okay. So we apply a couple methods. So like in the previous talk, we do both sort of positive and negative mode LC methods. Most of what we see are negative mode compounds again because they're kind of drop-in replacements for a lot of the legacy PFOA type compounds. So they're frequently carboxylic acid perfluorinated type compounds, especially around

fluoropolymer manufacturing. In specifically AFFF impacted locations, there's a lot more zwitterionic compounds, sulfonamides, and positively charged compounds that show up only in the positive mode. So we look at both sorts of modes.

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We know that we're blind to volatile compounds because we don't have a GC method that we apply. We sort of just focus on water surface, water extractable organic fluorine type compounds from soils and other stuff that's LC-able, because that's the only high res instrument that we've had for a while. But we've recently added more high res GC capability and we're hoping to move into that space.

Specifically, we know, in North Carolina and in West Virginia, as well as in New Jersey that there is a substantial air deposition portion to the story. And we see it in soil and in, you know, surface water that's impacted by things coming out. But there are volatile fluorinated products that are released via the air route that we can't measure at all. And some of them, we can determine -- we know what they turn into environmentally.

We can see the end products in the water.

Others, they're not known to -- you can't see them by

LC-MS and we know that we're missing them.

And so that's one of the reasons that outside of

my group, but in the EPA in general, they're working on, you know, total organic fluorine-type methods to try to get a handle on everything that's perfluorinated, so that we can fill in some of these gaps. So we're trying to be as broad as we can in our coverages by applying lots of different methods. But we do know that we're missing things and we try to be aware of that.

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I'd say we get -- I don't -- I don't know. I can't speculate. But we get most of the stuff that's important in water I think. Non-ionized stuff we're missing a lot and I don't know how much, because there's no way to know.

CHAIRPERSON SCHWARZMAN: Eunha, I think you're next.

PANEL MEMBER HOH: This is Eunha Hoh. I have -actually, Jon had to stop the presentation, because of the
time. But one of the slides that, Jon, you planned to
show that it was the loss of detection or identification
through the sample preparation or something like that. I
don't remember, was like solvents. You know, you added
the chemicals into solvent, and the next -- the other
stat, another stat, so we're kind of losing the detection
of the chemicals.

Jon, would you -- I think that's very important, you know, for non-targeted analysis in the rear samples,

you know. Would you give some comments on that or, you know, your thoughts on it. Yeah.

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DR. SOBUS: Of course. And I apologize again for having to basically skip those slides. I think the slides are available to the Panel members and the public. And we did go ahead provide some of the references of the publications. So if folks want to do a deep dive into that information, you should have access. If you don't, please feel free to reach out to me and I'll get it to you.

In a nut shell, that work was led by our colleague Seth Newton, where we basically took that SRM2585, which is a house dust -- a composite house dust sample. And Seth did a number of additional experiments. So he did experiments where he did multiple dilutions, simply in solution. So that was basically the best case type scenario, where you would expect performance to be optimal. Then he did experiments where he took that same spiked solution and added in dust extract basically to provide that matrix.

Then in another experiment he spiked at a very high concentration the dust and performed an extraction and a cleanup.

And then in the final experiment, he spiked at an environmentally relevant concentration into the dust and

then did the extraction and the cleanup.

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So basically, that slide was showing that we started with about -- let me see, I've got my notes. We started with 365 spiked chemicals. The best case scenario for his experiment just in the solvent spike, we identified 134. So we lost 63 percent off the bat, even in the best case scenario.

Once we took into account the matrix effects, we lost ten percent of the compounds just because of matrix interferences. When we took into account the extraction procedures and the cleanup procedures, we lost another five percent. And then when we took into account the concentration issues, we lost an additional nine percent.

So we went from 134 that could be identified under optimal conditions to 49 that could be observed under real-world experimental conditions. So it's just kind of making that point that, yes, the ENTACT mixtures is a really good way to kind of evaluate benchmark methods. But it's critical that we move into that kind of real matrix, real sample appropriation space, because it is going to drastically affect the performance of a given method.

PANEL MEMBER HOH: Thank you.

CHAIRPERSON SCHWARZMAN: Okay. And our -- last on our list of built-up questions is Ulrike.

PANEL MEMBER LUDERER: Hi. Thanks. Thank you, again for those really interesting talks. I had a question again also for James. And that was -- so a lot of these new PFASs are supposed to be less persistent than the -- some of the legacy ones. And I was wondering if you're -- in your analyses, you were able to determine whether you were finding some evidence of environmental breakdown of any of these chemicals, or whether you were really just -- you know, you were identifying only the chemicals that were actually being manufactured at these sites?

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DR. McCORD: So as I mentioned, we know a little bit about the chemistry from our discussion with the companies that are there. What we are detecting in a lot of cases are the terminal environmental Provex from things that they're emitting via air or other sources. They fairly rapidly breakdown, and so what we see are the terminal products.

And there's really no indication thus far that there's any further environmental breakdown. They are designed to be less persistent. But our experiments that we've done for the most part don't really bear out that they breakdown substantially more than a perfluorinated compound, specifically the structures that we've seen. So we did a little bit of sort of top assay type

experimentation, where you can hit them with, you know, strongly oxidizing conditions to force them to another terminal product. But all of the perfluorinated ether type compounds that we've seen, and even the chlorinated ones, would pretty much stay as they are, even under pretty harsh like top assay type conditions.

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We've seen evidence that things like ADONA, which is a polyfluorinated, and then things like 6:2 FTS obviously, they have chemical breakdown pathways that you can do under top assay type conditions and then also biological breakdown pathways. But it seems like the emerging contaminants they have the same types of general behaviors as the other contaminants do.

Now, they drop off in the environmental matrices that we're measuring, because, for instance, in the Cape Fear River when you turn off the effluent, they're the only source. And then the water dilution basically dries the concentration way down over a very short time frame.

So if you remove the source, the total concentration in the environment is so low that it drops below the limit of detection within a month or two. And then it seems like the clearance rate in humans is on the order of months to years. So the exposed population starts to get rid of it as well, but that just means that it falls below our analytical levels, not that it's

necessarily breaking down.

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And we've tried to do some sort of studies where you can put them in soil and do different things, and they don't really do much.

PANEL MEMBER LUDERER: Thank you.

CHAIRPERSON SCHWARZMAN: So I see there's a couple questions. I saw Oliver's hand and Carl's. I want to take a moment here just to go over the goals for the discussion question -- the -- sort of the prompts for the discussion session to make sure that we're getting the Program the feedback that they need, as well as kind of getting our questions answered.

So I'm just going to take a minute here to read the discussion questions and then set us loose on that.

And if Oliver and Carl have questions or comments related to that, we'll get to those next.

So some things that the Program is interested in hearing from us about are, number one, possible next steps for the Program in terms -- in the area of non-targeted analysis. Are there pilot NTA projects that you could envision that you think the Program should conduct? And what are the challenges or the difficulties that you would envision and sort of foresee as Biomonitoring California starts to try to integrate NTA into its studies? So that's the first question is next steps and potential

challenges.

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The second question is looking for opportunities to work with other groups that are already doing non-targeted analysis or collaborate with those researchers in ways that support the Program goals, and specifically to some of the guest speakers, can we arrange trainings with your groups for Biomonitoring California staff to sort of deepen staff's understanding of the methods that we heard about today?

And the third question from the Program is are there emerging chemicals or chemical groups that NTA is identifying that should be reviewed by the Program as potential designated chemicals? So if they're not on the chem — designated chemical list, but they're really kind of rising to the top in some of the NTA studies that are being done, should we propose those to the Program for consideration?

And finally, any suggestions for the Program about returning results from NTA studies to participants? And that gets back to some of the questions that we asked of Rachel in her study.

So those -- that's what the Program wants input on. Oliver, did you have a point you wanted to make with regard to that? Please go ahead.

PANEL MEMBER FIEHN: Yes. My question was

related to that to some extent it's both question and also comments for next steps and so on -- opportunities. So I found it very interesting today. It's very much to my heart, as you can imagine. But there is a problem that is shared between metabolomics and exposomes research in the sense of how to convey the coverage as well as the confidence in annotations, when you find a chemical and, you know, how sure are you, especially now when you report back to participants in a study.

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That relates to the question how we report confidence. Several speakers said they would use the Schymanski levels, but Jon made very clear that, in his presentation, that when his -- in his ring trial, you know, participants reported like they had every confidence. Schymanski level one with MS/MS and retention time and everything is fine, and it actually wasn't correct. So that was interesting.

And I see this both as a question to Jon how he would envision, you know, these confidence levels calculated or better maybe transparent to the public and to participants, as well as for us, including the biomonitoring program the State laboratories, you know, to think about the ring trial, because Jon specifically said they would invite other lab stories to participate as well.

And I think that would be a great opportunity for, you know, our laboratories, if they want to use NTA methods in the future to see, you know, how they would stack up to that challenge, right?

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Apparently, that was -- you know, it's very clear -- the study design is clear. It is possible to participate. It is -- the samples are ready, both the chemical mixtures, as well as the, you know, spiked matrices. And I think this would be a great opportunity for our laboratories to see how they stack up before they then dive into further, you know, resources, and further studies. So that was my question, as well as next step, and opportunities.

CHAIRPERSON SCHWARZMAN: Jon, if you want to respond or add anything to that, you're welcome to.

DR. SOBUS: Sure. That's a series of great points. And I agree with Oliver on pretty much all of them. Let me see if I can remember some of the points I wanted to make there.

First is kind of confidence levels in reporting.

Oliver and I had attended together a metabolomics workshop as ASMS last year. And something that really stuck out in that evening work group, probably attended by a hundred,

150 people, is how many people said how frustrating it was that even when they reported the confidence levels, when

they passed along that information, folks that were the recipients of the information didn't quite know what to do with it.

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In some cases, if they didn't know what to do with it, they would just kind of leave it off. So there's both the -- from the standpoint of the recipient not understanding the confidence levels, and not being able to interpret them and then there's perhaps something equally or more troubling, which is what I've experienced is the experts who are actually annotating some of these compounds with the confidence levels that, in my experience, are getting it wrong, right?

So you have some people that aren't using confidence levels, some people that send the confidence levels forward to have them misinterpreted or not interpreted, and then some people that send them forward but they're actually the wrong confidence levels to begin with.

So I think that's something that is going to be addressed through education, and through consensus, and through adherence. I don't necessarily think the solution is making confidence levels more refined or more complex. I think that may have the effect of pushing people away. But we -- we had I think and hour and a half or two hour discussion exactly on that topic and there was no

consensus, at the time, amongst a community of experts.

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So the confidence level reporting thing is extraordinarily complex. And, Oliver, like you said, even if you report something in a confidence level, it's very difficult to know whether you can accept that as truth at a particular confidence level. One of the things I'm doing, but I didn't talk about is trying to assess performance not only for a specific method or a specific lab, but the performance level at a specific reported confidence level.

So the difficulty here is is you need to answer those questions because they're critical, but the more I dig into these performance metrics, the more nuanced they get, the less accessible they get to my target audience. So it's incredibly challenging. And I think the solution again is to have as much discussion as possible, as much education as possible, and just transparency in what's being done.

So that's -- that's kind of the confidence levels and reporting confidence levels in terms of performance review. Again, I think I understand what you're saying. And if I do, I entirely agree with you that in my mind, you know, for targeted analyses right now, there are -- there are bodies that do credentialing, right? Some specific targeted methods have to be credentialed in order

to do work. I see that as the future for environmental NTA.

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And part of what we're trying to do through the ENTACT project is put the research community on a path towards credentialing. And this -- these are my opinions. But in designing ENTACT the way we did, we kind of had that in mind to say what would a study need to look like to be able to fully and usefully evaluate performance?

Now, some of the drawbacks in the way that we've done it is we started it in 2018. It's now 2020 and I've still got a long way to go in terms of the evaluation. I think the participants would agree unanimously that it was more work than they thought it was going to be when they agreed to participate.

So I think my recommendations right now would be, A, this needs to happen. There needs to be some type of samples that can be used for evaluation and the contents of those samples need to be kept secret. I think there needs to be an independent third party that is appropriately resourced to manage these projects and to efficiently do the data analysis that I'm trying to do right now.

It's incredibly difficult to find the time to clean the files, to process the files, to match it, and to constantly shift thinking and have to redo things. So

whomever that third party would be that would be in charge of the credentialing, again would have to have the appropriate resource to be able to work efficiently and get responses to these laboratories to get them credentialed as soon as possible, so that they can support actual experimental work.

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CHAIRPERSON SCHWARZMAN: Dinesh, I think you have something to add here?

DR. BARUPAL: Yeah. So I have a comment on this one that to whom we'd want to communicate the result. So if it's a question of development of a database or publishing a manuscript, I think we can use different scheme of levels for annotation. But moment we want to share those with participants, or clinical decision making, or some other hard decision making, they need to be passed through that they will -- annotation were confirmed by pure standards, because any misannotation can be translated into wrong decision-making.

So after database development or if we want to just publish some good manuscript, it's okay to use these levels and discuss about that, the pros and cons about. The moment we want to give it to a participant, we want to be really sure that it has been confirmed by pure standards.

CHAIRPERSON SCHWARZMAN: Carl, did you have a

point, suggestion, or a question? You were next on the list, I think.

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PANEL MEMBER CRANOR: Are you talking to me, Meg, I'm sorry?

CHAIRPERSON SCHWARZMAN: Yes. I thought you had raised your hand earlier.

PANEL MEMBER CRANOR: I have, yes. First of all, I think this is a great set of presentations. I hope that it is useful to us and to the State. I want to -- let's see, what could be next? Well, I think the generic idea behind non-targeted testing is to identify things that are maybe not yet seen to be a problem and then to send them through an appropriate system as quickly as possible to see whether they -- people ought to be protected from them. That's point one.

Point two though is that non- -- in some sense, non-targeted testing and biomonitoring are, what I would be tempted to call, legally post-market, but let's think of it as not fully preventative of heath effects to people, because they're already out there, and now we're discovering them, and tying to do something about them.

So a second point I would make about our next steps, is there a way to design non-targeted testing, so that they can be used in a much more preventive manner?

Now, I thought the case study from Cape Fear was

very interesting in that sense, because you were looking at substitutes for the perfluorinated compounds. That raises a couple of -- well, it raises a couple of legal questions, but also a couple of preventive questions.

Were those, the second- or third-generation substances, something that had been properly approved by the U.S. EPA? I suspect not or didn't get much scrutiny. But they're out there -- the suggestion was that they're still very persistent and guide -- I'd seen a few papers that suggest that there's toxicity.

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So to what extent can our non-targeted testing not only pick up new things that we should worry about, but can -- can be pushed into the existing legal structures, for example, to hold companies accountable if their substances that they're using haven't gone through the appropriate EPA presumably pre-market approval that is supposed to be part of the new TSCA?

So I do worry about both non-targeted testing and biomonitoring as being -- having preventive opportunities but very late in the game. And so I would urge people to think about how to make them more preventive upfront to the extent possible and see where we can go that way.

DR. McCORD: I can -- I can comment on it. I don't know if I'm supposed to raise my hand first or not.

CHAIRPERSON SCHWARZMAN: Please, go ahead.

DR. McCORD: Okay. So I am not an expert in TSCA, but I'm going to give some of my understanding of what has gone on specifically in the Cape Fear and sort of more broadly.

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So GenX, the compound that we were able to identify a standard for was sold as a product by Chemours as a replacement for PFOA. There was the sufficient TSCA approval and available toxicology data, both in America and in Europe for that compound -- particular compound.

In that particular instance, the State of North Carolina actually had prohibited the company from emitting GenX from their manufacturing process at the facility. So they made that particular compound in one of their processing lines in a closed loop process that generated no waste. However, it turns out, it's also produced as byproduct of their polymer manufacturing process and used internally within that process, in addition to the fact that they make it closed loop and then ship it to other places where they use it.

And all of the other compounds that we identified in the Cape Fear are also byproducts of their mostly fluoropolymer manufacturing process. And it makes sense from an industrial perspective if you can use the byproducts of your polymer manufacturing as your processing aids and everything is internal, then you don't

have to make products all on their own. And they're also exempt from TSCA regulation, because my understanding is that TSCA does not cover intermediates and other products with no sort of commercial purpose. They're simply waste products and then they are byproducts and used internally, because they're never designed to be sold or emitted in large quantities or anything like that.

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So the company ultimately was responsible for emitting GenX, because they have a specific rule about non-emitting it and were actually emitting it as a byproduct, but the other compounds are not covered under the same type of TSCA regulation, just because of the nature of the types of chemicals that they are.

Likewise, in New Jersey, the chlorinated compound has some toxicology studies and it has approval as a food contact material under ECHA. And I don't think it has any TSCA data that I have access to. It might be filed under CBI, which I don't typically have.

So some of these compounds, they -- they do have toxicity data, if it is ever anticipated that they are going to be emitted. But a lot of the compounds that we find fall into this sort of gray area. They're intermediates and other products that they're never supposed to get out. They're only ever found as trace contaminants of the finished product or side products of a

reaction that wind up in a waste stream, and then they expose 75 miles of the surrounds for a particular manufacturing facility. So they fall into this legally questionable area.

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The question of how you can apply non-targeted analysis to the TSCA process is something that I'm particularly interested in. And we have a project that we're in the very early stages of of trying to figure out how we can do non-targeted analysis particularly on things that are mixtures and that might have byproducts and other trace contaminants within TSCA. However, I think that we are a very long way away from even getting non-targeted analysis data incorporated into the TSCA process for characterization, because in many cases, TSCA approvals and significant use rules -- significant new use rules and things like that, there's no experimental data that doesn't come from the company.

So until a consultant has a QA problem -- that we can solve a QA problem, where that data can be part of a regulatory like data submission package, I don't even know how we can get people to look at that data. So solving the QA problem and figuring out to like deal with that type of data is I think the first step in getting regulatory bodies to look at it and being able to interpret that data.

And then there's the challenge of figuring out how do you do rapid toxicology characterization on unknown compound if you do detect them, because the goal, I think, is to avoid turning environ -- and the goal, it depends on who you think is really in charge of things. But the idea would be that you would like to avoid having to characterize every potential chemical in existence in order to have approval for anything.

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CHAIRPERSON SCHWARZMAN: Thank you for that. But I want to interrupt here for -- just for a sec to say that -- to allow enough time for this discussion to really carry through, we're going to do a small change to the agenda, which is move the public comment period from the end of the day up to 4:05 p.m., and then return to this discussion until 4:20, when we move on to sort of wrap-up the day.

So just to say we have more time for this discussion. If anybody -- the reason I wanted to mention it now is if anybody wants to make a public comment, they should please prepare it and either indicate to us that you would like to speak, or send it into the email address, or indicate it on GoTo meeting and we will come back to that at 4:05 p.m.

MS. HOOVER: Meg -- Meg, let me just chime in real quick. This is Sara. I just wanted to clarify that

there are a bunch of public comments right now that are relevant to the NTA discussion, so we could call for those basically now.

CHAIRPERSON SCHWARZMAN: Right. That's what I was going to say.

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MS. HOOVER: And the 4:05 -- just to clarify to the audience. What we're saying is normally at the very end of the meeting we call for open public comment on any topic. And what I'd like to do at 4:05 is just hear is there any open public comment, given the fact that we haven't had any -- or much of any public comment all day. So we'd like to keep this going, because it's such a rich discussion and lots of topics. So anyway, continue. Thank you, all.

CHAIRPERSON SCHWARZMAN: Yes. So I'm going to do the comments and questions that have come in over email now, but I just wanted to give people a heads up about the public comment period will be at 4:05, not at the end of this discussion or we'll check for public comment at 4:05.

So with that, I want to turn to the comments and questions that are coming in on the GoToWebinar. And I want to invite Marley to share those.

CHAIRPERSON SCHWARZMAN: We can't hear you, Marley.

So if that's --

MS. ZALAY: Can you hear me now?

CHAIRPERSON SCHWARZMAN: Yes. Good.

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MS. ZALAY: Okay. This is a question from Susan Hurley with Biomonitoring California for Doug. Can you elaborate more on the silicone badges, are they ready for prime time?

DR. WALKER: So I would say they serve a very specific purpose. You know, it makes it very difficult to take what we're measuring on the wristbands and actually extrapolate that to an exposure level. We use them as more of a screening tool to try and identify what exposures are occurring and what we can detect. I think they are ready for use in that context. The nice thing about them is they're very cheap to produce. They cost about \$0.25 to buy the wristbands. And then we have to clean them to remove impurities that were added during the manufacturing process. That costs about another four to five dollars.

But, you know, they're cheap enough where you could actually produce a lot of them, send them out to cohorts, collect them, and then analyze subsets from those that you distributed. So say you want to provide these wristbands to a population and then study a certain outcome, you could easily send out a thousand or two thousand of them, collect them, and then re -- you know,

only analyze the samples from people who ended up, you know, developing an outcome or a disease. So in that context, they're very useful.

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But you have to be careful about how you're interpreting the data, because we can't, like I said, quantify what exposure is. We can just use it as a screen to identify what exposures are occurring.

MS. ZALAY: Thank you. There's one more question and one general comment.

I'll read the comment first and then I'll read the question. Miaomiao Wang from the Environmental Chemistry Laboratory at DTSC, the Department of Toxic Substances Control, commented that among the labs that participated and finished the ENTACT trial, they were among those -- those labs. And they also participated in the EPA workshop in 2018 and they're working on a manuscript and look forward to performance feedback from EPA.

And then lastly, there's a question from Martin Karozy saying NTA can be used to measure exposures, but it can also be used to measure biomarkers of disease. Can anyone talk to examples of this?

DR. WALKER: So --

CHAIRPERSON SCHWARZMAN: Can you just say something about that relevant -- let's take it within the

context of what Biomonitoring California does, which might be to make some of these links between exposure and disease. But please go ahead, Doug.

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DR. WALKER: Yeah. So we kind of see the disease biomarkers as an intermediate between the exposure and the outcome itself. You know, that's the advantage of using the untargeted metabolomic platforms is you can characterize all these biological processes and link them to the exposure biomarkers that you're measuring. But I mean that -- that is a very good point that was made by the commenter question. You know, a lot of these untargeted exposomic methods that we're using now were kind of borne out of more traditional metabolomics analysis. And those have been used in a number of applications for more disease biomarkers.

So there's -- you know, there's that kind of historical use, but another important thing to recognize is that the exposome or the exposures that we carry and potential development of exposome risk scores could also act as biomarkers of disease once we start better understanding, you know, how our exposure profiles overall contribute to our health outcomes, or, you know, other things that we're addressing and studying.

CHAIRPERSON SCHWARZMAN: Okay. Thank you for fielding those.

Jenny.

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PANEL MEMBER QUINTANA: In terms of the Panel giving guidance, to Biomonitoring California, I was wondering if we should have some discussion about the benefits of non-targeted analysis.

So, for example, what I've been hearing is that we can use it for finding regrettable substitutions. It's very valuable where people are changing what's in products. Another use I heard was byproducts, finding byproducts, which aren't necessarily known. The third might be finding unrecognized pollutants in lots of people that we didn't know was a big -- very widespread. And then fourth was risk from specific exposures like firefighting.

By the way, someone has their microphone not muted that's typing. Just as a -- let you know.

So -- and speaking about risks from specific exposures, like the firefighters, I did also wonder if there's a role for other samples that could really drive the non-targeted analysis, like, for example, air samples during a firefighting event. Maybe identifying chemicals there might help drive what you might look for in people.

So I just thought I'd maybe bring up should we talk about what would be the biggest benefit to Biomonitoring California, and then we could decide how to

achieve that first -- second.

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CHAIRPERSON SCHWARZMAN: Yes, Jenny. I think that's excellent. What we're trying to do is sort of summarize this and make recommendations to the Program where relevant. So if you have thoughts to start that. I mean, thank you for your -- for getting that started.

Did you have recommendations you wanted to add to that, Jenny?

PANEL MEMBER QUINTANA: I wasn't sure if "you" was the Panel or "you" was me.

My recommendation would be for substitutions. Finding those regrettable substitutions would be a very important use, I think, and also byproducts. So I think those two things would be very important. But apart from that, I think I would really vote for looking at specific populations. And the specific populations in addition to what's already being studied, I would say should be disadvantaged populations, whether it's a community or whether it's workers in a certain industry that -thinking about California, what makes California unique, perhaps compared to other states, is our high proportion of refugees, and immigrants, our diverse industries. think we should think about California also and what makes us unique as well and our proximity to the border. case of where I live, people exposed to pollution from

open burning in Tijuana. You know, there's -- there's a lot of unique features in our a population and a lot of agriculture, for example.

So thank you.

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CHAIRPERSON SCHWARZMAN: I agree, Jenny. And I think the examples of, you know, what's been done in Delaware and the other examples from the same presentation that are sort of site specific. And we're looking at, you know, what's -- what's in this area are really interesting models for us to sort of think about adapting to our state.

Dinesh, you had something to add there and then -- and then Carl, sorry. Just to let you know I've seen you.

DR. BARUPAL: Thank you. NTA has a great advantage in all -- all these -- some different scenario is the number of analytes. So you -- sometimes you only have hundred microliter plasma sample. And the question is do you measure ten compounds using -- using a targeted approach or do you measure 5,000 compounds with that hundred microliters and store all the data.

So -- and in this setting, NTA really has a great advantage that we should do NTA if it has the right combination of analytical chemistry, sample preparation, and the data science approaches.

CHAIRPERSON SCHWARZMAN: Okay. Thank you.

Tom.

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PANEL MEMBER McKONE: I forgot to reach for that mute button.

Thanks. I actually want to continue on the ideas that Jenny brought up. And I think -- I was thinking of it a little more broadly though, which is -- I mean, if you look at the history of environmental health sciences, we've always tended to look backwards and do a better and better job of detecting the chemicals we already know about.

And I think, you know, we've done some of that here. We're always building our list. But to the -- we want confidence and we're picking chemicals that we're going to find. And we want confidence that, you know, something somebody else has already done.

So what came up for me today, especially on the non-targeted assessments, is part of a broader issue of how do we really start being a little more proactive in finding chemicals that we haven't quite looked at well enough or don't even know about yet. I mean, there's -- there's this whole idea that there's -- you know, we heard really big numbers, hundreds of thousands of environmentally-related chemicals that might be, you know, in the metabolome or the exposome. And it's just a --

it's kind of overwhelming.

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So I thought, you know, in terms of strategy, I think non-targeted assessments is one component of maybe a broader effort to look at this is a really good tool. We learned a lot about it. But I also think the Program should be looking at kind of a portfolio of tools to find -- to make sure we're looking forward, that we're really anticipating what's in the population now or might be in the population in a year, instead of really still tending to look backwards about, oh, well, we just found out from other people that this is an important issue.

I mean, it's kind of what we did with flame retardants, so -- so -- and it's a funda -- I think what really brought this up is when I asked Jon Sobus about, oh, is this all you do or do you have a way of really targeting it. And he's -- he mentioned it. He said, oh, it's too much to talk about today, but that there's a whole series of activities to go through to try and build lists and then narrow them down, to then do their non-targeted analysis, not on everything in the world, but really things that show up. So there are these methods for kind of triangulating among different ways of looking at it.

CHAIRPERSON SCHWARZMAN: I'm going to use Chair's prerogative here to insert a comment, that it kind of

connects to what Tom just said on this issue of sort of finding replacements and the evolution of the market as it changed. This has always been such a weakness is our inability to anticipate given the sort of policy structure in the U.S. where there isn't public disclosure of where an industry is going and what chemicals they're putting in what.

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And so I even want to move beyond Jenny's kind of depiction of regrettable substitution, because often they're substituted with chemicals for which we have no toxicological data and so we don't know if it's an improvement or a worsening.

And Biomonitoring California has made such a tremendous step in enabling designation of chemical classes. And sometimes those classes are chemically based and sometimes they are more functionally based, or at least groups will -- class might be putting too fine a point on it. But sometimes there's a functionally based sort of group that's assessed, like where is the universal flame retardants moving from, you know, halogenated to non-halogenated flame retardants.

And so I think that's a particularly exciting use of non-targeted analysis and could be sort of combined with the other themes that have come up about kind of place-based investigations, like the ones that James was

describing, and in selecting those, identifying particularly hard hit communities or populations to understand what they're being exposed to.

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Those are the main things that kind of rise to the surface for me. Nerissa, do you have a response to that or something you wanted to add?

DR. WU: I do. And I agree with all of this. I think this has been a really exciting discussion. And it feels like the future of certainly little "b" biomonitoring, if not Biomonitoring California.

But I do wonder how this fits into our Program in terms of what our priorities are. We're small a Program.

We all know kind of the limits of what we can do. We have a mandate to do surveillance work. And there's this kind of dichotomy between really targeted, whether it's community based or geographic based, studies where there might be more motivation to look at a particular group of chemicals, and some tolerance or some uncertainty of what we're looking for versus surveillance, where we already know we have -- you know, there -- there's a lot of concern about giving your biological samples over to be analyzed. And if there's a thought that you might be looking at, you know, kind of a limitless range of chemicals, people might be less inclined to participate in something like that.

So as I always sort of find myself sort of thinking about how do we prioritize? We are legislatively mandated to do surveillance work. We are legislatively mandated to return results to people, and in a way that people can understand, and are useful to them, and are -- you know, that are educational and how do we balance that with this exciting work, which I think is really needed on this more targeted community basis, either the community is targeted or maybe the list of chemicals is a little more targeted. So it's just not a wide open swath of results that might be returned to people.

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CHAIRPERSON SCHWARZMAN: I wanted to continue this line of discussion, but I also want to stick to on our promise to check for public comment at this time. And so let's just put a pause on that for one sec, while we check for public comment, and then continue that line of conversation.

So Marley and Sara, is there any one indicating the would like to make public comment at this point?

MS. ZALAY: This is Marley Zalay. There's no additional questions from GoToWebinar.

MS. HOOVER: And I can confirm there's no questions in the email either, so please continue with this discussion.

CHAIRPERSON SCHWARZMAN: Great. Then we have

until 4:20 for this discussion.

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I appreciate the -- Nerissa bringing us sort of to the practicalities of the Program. And also one of the things it made me think of is the one way that -- by -- non-targeted analysis can even increase the surveillance aspect of biomonitoring work in ways that targeted analysis may be even is just much more limited. But I say that in the context of having heard this interesting thing that Rachel said, which was that the participants were less excited about NTA than they were about looking for the particular chemicals that had sort of risen to their consciousness and level of concern.

And I really appreciate the point that you're making, Nerissa, about sort of the palatability of these kinds of studies to participants.

Anyway, I'll step aside and let somebody else.

I think, James, did you want to respond to that?

DR. McCORD: Yes. So I've been associated with this problem in a lot of different cases. So we've worked with biobank materials and we have some ongoing studies related to that where we're doing both targeted and non-targeted work. And also, we've had to deal with reporting to communities that are affected this, where we've done targeted and non-targeted work.

And to also piggyback of something that Dinesh

kind of said earlier about choosing like when you're going to do targeted and non-targeted work. In a lot of cases, the work that we've done, we've sort of extracted targeted information out of non-targeted experiments. You can do a full scan type non-targeted experiment and also use it to generate some quantitative data, if you have standards and can run calcurves in conjunction, and you have the right method that gives you decent chromatograph peaks and things.

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So you can process a sample in many cases the same way that you would do a targeted experiment, still get targeted data, and then have all this excess data lying out. It depends on exactly the type of information that you're looking for in your targeted data. Some methods are very specific and will only -- sort of like Jon said, he only makes certain chemicals all the way through the process. But if you have generic enough preparation methods for certain classes of chemicals, you can definitely do both the targeted approaches that people are used to and expect and then also collect non-targeted surveillance data.

In many cases, when we've done this, our IRB and our subject's participation agreements have basically said that we hope to do this type of surveillance work in order to identify new materials, but we've only ever reported on

specifically the targeted data back to the participants, because only targeted data is associated with real risk assessments where we felt like we could give an individual any information related to their results or the associated health risk.

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And then we would only report the aggregate data for things that are non-targeted where there's less actual information. Because as I kind of mentioned when we dealt with the Wilmington participants, it's very difficult to make someone happy when you say we found a lot of different things and you have no idea what it means, because it's difficult to be comfortable with sort of the ambiguity of that process, right? And you'd like to avoid the anxiety associated with not knowing, if -- for the -- for the large majority of cases.

So it depends on what your exact legislative mandate is in terms of what you cannot do.

MS. HOOVER: Yeah. Perfect seg -- perfect segue, James. This is Sara Hoover. I'm just going to chime in. And this is -- this is a great setup for one of our questions, which is we have to return NTA results. So anything we measure on participants officially, we have to return.

Now, we've done a pilot project. I shared with everyone the pilot packet that we developed. Rachel had

some comments about, you know, working on participant returns. So that will be a fundamental part of any project we do. Except for the comment about biobank samples, that's definitely an opportunity where we can't return the results. So we could do more broad types of screening without having to think about results return.

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But I just wanted to throw that in there, because that kind of is a foundation of our Program. And so we don't have the luxury of -- we have to return all results. Anything that is a result we return. And you'll see how we did that for the very small pilot project.

Again, I think someone else just mentioned too -it's a very good point - I think Nerissa raised as did
others -- focusing in on a class of chemicals of a great
concern -- of a great interest -- was what we did. So we
didn't do actually non-targeted. Even though we called it
that, it was actually targeted towards just PFASs. So
that simplified some of the report back.

CHAIRPERSON SCHWARZMAN: I want to make sure that we hear -- capture for the Program all the Panelists' suggestions. So I want to scan who has something to contribute, in our -- in our last six or seven minutes. So, Veena, and is there anyone else that I should stack up lined Veena. Veena and Eunha. Okay. Go ahead, Veena.

PANEL MEMBER SINGLA: So I really appreciated

Jenny's kind of overall suggestions on thinking about specific populations, especially vulnerable populations and also appreciate the challenge of balancing that with the Program's mandate for surveillance.

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You know, I think I'd just say in the current moment when the pandemic has really laid bear the racial disparities that contribute to health disparities that it's really important to think about how we can better understand the cumulative burden of environmental exposures on vulnerable populations.

And I wonder if non-targeted analysis could help us do that, both in terms of chemical and non-chemical stressors and biomarkers of stress. And if that information could be helpful in making decisions -- actual policy decisions, because there is a really interesting New Jersey bill that's focused on cumulative impacts and integrating cumulative impacts into siting decisions before making siting decisions.

So I think it's something to think about how non-targeted analysis could help us better understand cumulative exposures and also how that information could be used to inform decisions.

CHAIRPERSON SCHWARZMAN: Great. Thank you, Veena. And Eunha.

PANEL MEMBER HOH: Yes. I think I'd like to

think about the -- like what we want to do in rare situation, what we can do, what is the feasible thing, you know. I mean, I'm running a laboratory as well that I don't -- I know how difficult it is. Like, we constantly adding more, and more, and more with a limited resource. It's a very, very difficult task.

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I was thinking about the non-targeted analysis, based on the biomonitoring samples like the very limited amount of blood samples. It's -- I'm not sure if really have a lot of resources. It's probably eventually that's the way to go, but I think it's something that can be more feasible, that like Jenny mentioned like something like more environmental samples in a certain community, you know. That could be done for the non-targeted analysis, if we see the chemicals that we completely overlooked, you know.

So that's what I kind of -- kind of can vision, you know, how we may want to put it into that, you know, the current Biomonitoring Program.

CHAIRPERSON SCHWARZMAN: Thank you for that. I want to put out a question to all our guest speakers about -- so the Department is -- or the Program is asking us, you know, are there ways that they should be deploying non-targeted analysis in realistically, currently in studies? And I guess I have an open question about sort

of the viability at this moment of using NTA methods -- of the Biomonitoring California Program, using NTA methods given, for example, all of the sort of method issues that Jon Sobus presented.

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So if the Program is asking us are there pilot studies that they should consider doing, I'm curious if the guest speakers have any brief, because we're almost out of time, input about what actually is possible right now?

DR. MORELLO-FROSCH: This is Rachel. I think in terms of addressing interests around disproportionately impacted communities, the approach that was taken, for example, to focus NTA strategies on chemical classes of concern is a nice way to start, especially because of the report-back requirement baked into the Program.

And so that gives you opportunities to do surveillance of interest and to see what's going on with chemical substitutes when you know that certain things are being phased out and yet products -- certain products are continuing to be manufactured. And it also allows for a meaningful way to do community collaboration where appropriate, either place-based or occupationally-based or whatever, in terms of speaking input and people understanding kind of what are the benefits and pitfalls of a non-targeted analysis.

But your -- you have sort of a -- focusing on chemical classes, your -- you have your known unknowns basically. And that's probably more doable than sort of completely non-targeted analysis.

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CHAIRPERSON SCHWARZMAN: Thank you for that. Doug, did you come on because you have a response?

DR. WALKER: Yes, I did. Thank you. You know, one thing the we should also think of, whenever there's an opportunity to compare targeted and non-targeted platforms, I think there's a lot of advantages to doing that. What we've been able to show for our assays is that we do get comparable detection to a lot of the more targeted platforms.

And so it supports using these non-targeted platforms and anchoring in what we know. And then in addition to that, you can start screening for, as was just mentioned, you know, other compounds from certain chemical classes. So in terms of pilot studies, if there is an opportunity to apply a non-targeted method to samples that have already been characterized by targeted analysis, I think that's a really excellent opportunity.

CHAIRPERSON SCHWARZMAN: Great. Thank you.

We need to turn toward our final summary and wrap-up of meeting. And I guess I would just invite, I think the Program is always open for input and eager for

input. So if there are ideas that we haven't captured today, I think the Program would welcome them by email in follow up.

So with that, I want to turn this over for a moment to Vince Cogliano who's the Deputy Director for Scientific Programs at OEHHA. And he will be providing a brief summary of the input and action items from today's discussion.

Vince, please go ahead.

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DR. COGLIANO: Thank you. Thank you very much, Meg. I'm unmuted. Okay.

So, let's see, we heard quite a bit -- and first of all, I'd like to congratulate all the speakers and participants who made comments. This has been a very useful meeting for me to understand this Program. And there's just a lot of exciting work going on here. And I think it will really help us advance risk assessment and environmental protection over the next several years.

I guess we heard quite a few things. We learned a lot of -- we heard some summaries of some of the programs from California Department of Health or the Department of Toxic Substances Control that are contributing to this. We learned some of the activities going on at the U.S. EPA about validating or coming up with methods that can be used to figure out which -- which

new methods are suitable. We've seen some applications of NTAs in exploring PFASs in North Carolina, and where they found many other chemicals that are related to the PFAS exposure outcomes, but which hadn't been identified before. And also the New Jersey case study that came up with emerging compounds we should look at.

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We learned about some cheminformatics tools to support exposomics, and metabolomics, and framework for measuring the exposome. In the morning, we had a specific item recommended by the Panel recommending that the Biomonitoring California Program design an NTA pilot project in a specific population, for example, in a disadvantaged community, which we heard quite a few of the comments in the last half hour, or in a specific occupational group, or refugee group, or another group relevant to California's unique population.

It would be important to examine the cumulative burden of exposures, chemical and non-chemical, in these heavily impacted communities and to suggest that Biomonitoring California looked at ways to use non-targeted analysis to address and inform these decisions that we make at these -- in these populations or at sites.

There was also encouragement during the day for the Program to look for ways to use non-targeted

assessments to be more proactive in finding emerging chemicals, including -- including regrettable substitutions in chemicals which almost nothing at all is known.

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In my long career in environmental health, I now have witnessed quite a few times when -- in my first ten years of my career we banned or phased out some substance and then ten years later we were concerned about the new substance that was being used. And I think these non-targeted analyses could really give us a handle on what chemicals might be better or worse for substitutes.

We have quite a few different tools and methods being developed, so we should be using a portfolio of tools to find out what is in the population now and what might be appearing in the future due to shifts in the market.

So there's a focus on looking forward and not looking back. You know, there's the comment made that we often refine our methods to look at chemicals we know about and get better and better methods of identifying them, and -- instead of looking at the chemicals that are actually emerging.

And then there was a suggestion to think about a practical and feasible way to tar -- apply targeted non- -- non-targeted analyses using environmental samples,

for example. Also, to ensure that we keep monitor

California's major priorities and participant feedback in

mind when we would consider future projects.

We also have to grapple with the result -returning results for any project, in case -- in
California, based on the statutes that we have here in
this state. So that's another thing that whatever we
find, we have to disclose.

So I think that's my quick summary of some of the recommendations and things that we've heard today.

And I'll turn it back to you, Meg.

CHAIRPERSON SCHWARZMAN: Thank you for that,

Vince. I appreciate the wrap-up.

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MS. HOOVER: Meg.

CHAIRPERSON SCHWARZMAN: I'm sorry, Sara, what?

MS. HOOVER: Yeah, I just wanted to toss in one morning recommendation just to capture the action items and that was just the item about the Panel would like OEHHA to figure out if halogenated carbazoles are already captured as part of any chemical group that is already on a designated list. If not, OEHHA should track this group of chemicals to consider for a possible preliminary screening.

And like I always do, I'll put up -- CHAIRPERSON SCHWARZMAN: Oh, we lost you, Sara.

You said, like I always do. So, Sara, I can't understand you anymore. Do you want to try again?

MS. HOOVER: Let me try again.

Nope.

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CHAIRPERSON SCHWARZMAN: That sounds better.

MS. HOOVER: Am I back?

CHAIRPERSON SCHWARZMAN: Yep

MS. HOOVER: You guys are frozen.

Okay. Sorry about that. Just real quick. I'm putting out a pitch for anybody who can hear me now to send in emerging chemicals of interest. Anything you become aware of in your work or in your NTA projects. I'm having an unstable internet connection. Apologies for that.

CHAIRPERSON SCHWARZMAN: We can hear you though. That's good. So for all -- everyone who's involved now in doing non-targeted analysis to keep in the back of your minds, if something interesting rises to the fore to shoot an email toward biomonitoring to say, you know, we're seeing this new interesting thing that you might want to take a look at.

So with that, I want to announce that a transcript of this meeting will be posted to the Biomonitoring California website when it's available. The meeting was also recorded and I'm not clear what's

happening with that recording, but it may be that it's being used by the transcriber.

The next SGP meeting is on November 12th of this career. And it will also be held as a virtual meeting.

I want to really thank the Panel and the speakers for everything that you've contributed to this meeting, as well as the audience. And a tremendous thank you to the Program leadership and Program staff, because there was significant legwork involved in making this meeting happen and transitioning it to a virtual format. And I think it was shockingly smooth, given all of the potential hurdles. And it's really kudos to a lot of time and effort that the Program leadership and staff has put in to making this work.

So thank you to everybody who participated and I'll adjourn the meeting.

(Thereupon the California Environmental Contaminant Biomonitoring Program, Scientific Guidance Panel meeting adjourned at 4:29 p.m.)

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CERTIFICATE OF REPORTER

I, JAMES F. PETERS, a Certified Shorthand
Reporter of the State of California, do hereby certify:

That I am a disinterested person herein; that the foregoing California Environmental Contamination

Biomonitoring Program Scientific Guidance Panel meeting was reported in shorthand by me, James F. Peters, a

Certified Shorthand Reporter of the State of California, and thereafter transcribed under my direction, by computer-assisted transcription.

I further certify that I am not of counsel or attorney for any of the parties to said meeting nor in any way interested in the outcome of said meeting.

IN WITNESS WHEREOF, I have hereunto set my hand this 28th day of July, 2020.

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JAMES F. PETERS, CSR

Certified Shorthand Reporter

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